

## **A method for the synthesis of spiro-1,3,4-thiadiazolines**

**Alexander V. Komkov<sup>1</sup>, Leonid G. Menchikov<sup>1</sup>, Andrey S. Dmitrenok<sup>1</sup>,  
Natalya G. Kolotyrkina<sup>1</sup>, Igor V. Zavarzin<sup>1\*</sup>**

<sup>1</sup> *N. D. Zelinsky Institute of Organic Chemistry, Russian Academy of Sciences,  
47 Leninskii Ave., Moscow 119991, Russia; e-mail: zavi@ioc.ac.ru*

## **SUPPLEMENTARY INFORMATION**

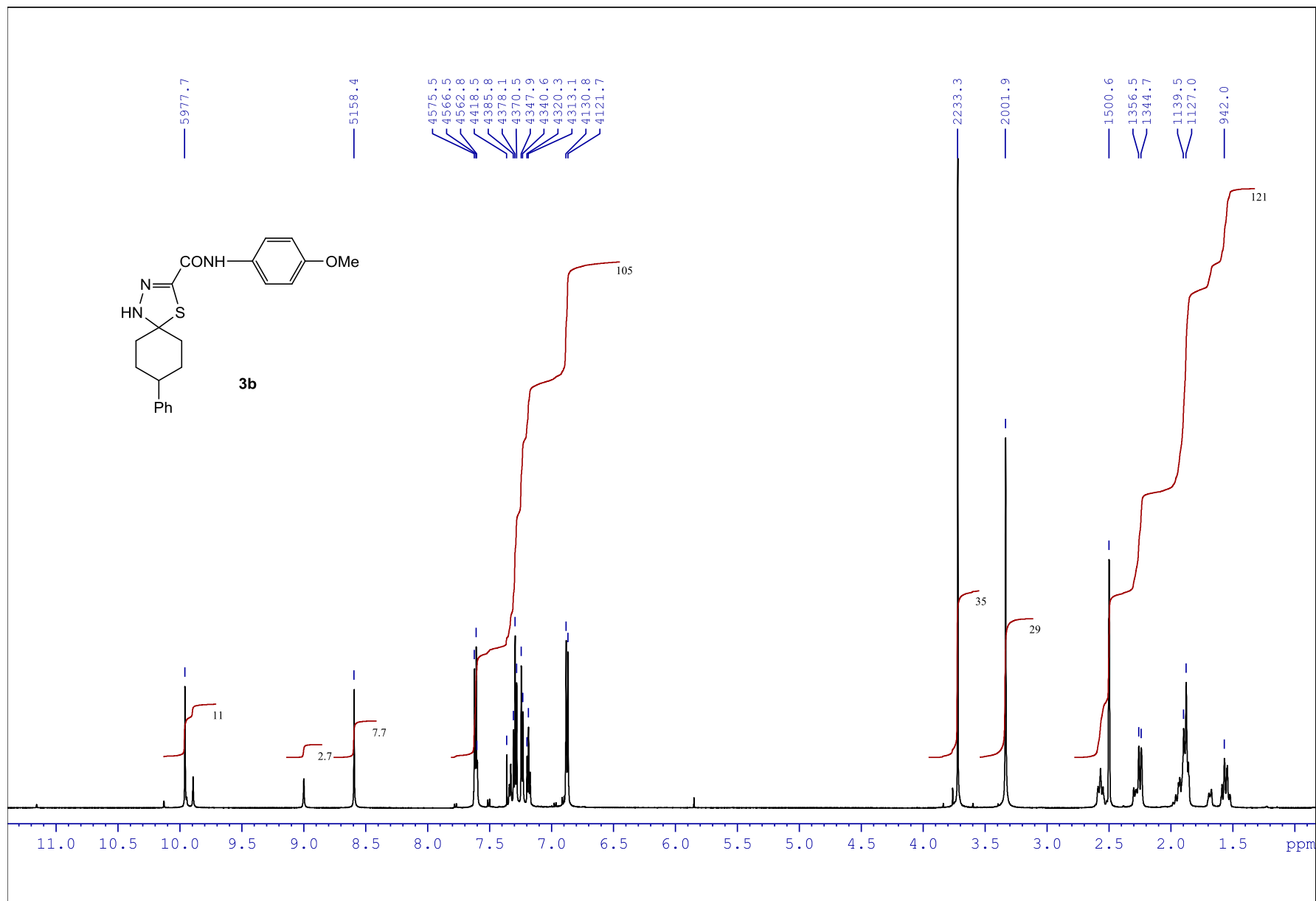
### **Table of contents**

1. Experimental Section.....	S2
2. NMR spectra.....	S3
3. IR spectra.....	S79
4. Mass spectra .....	S87

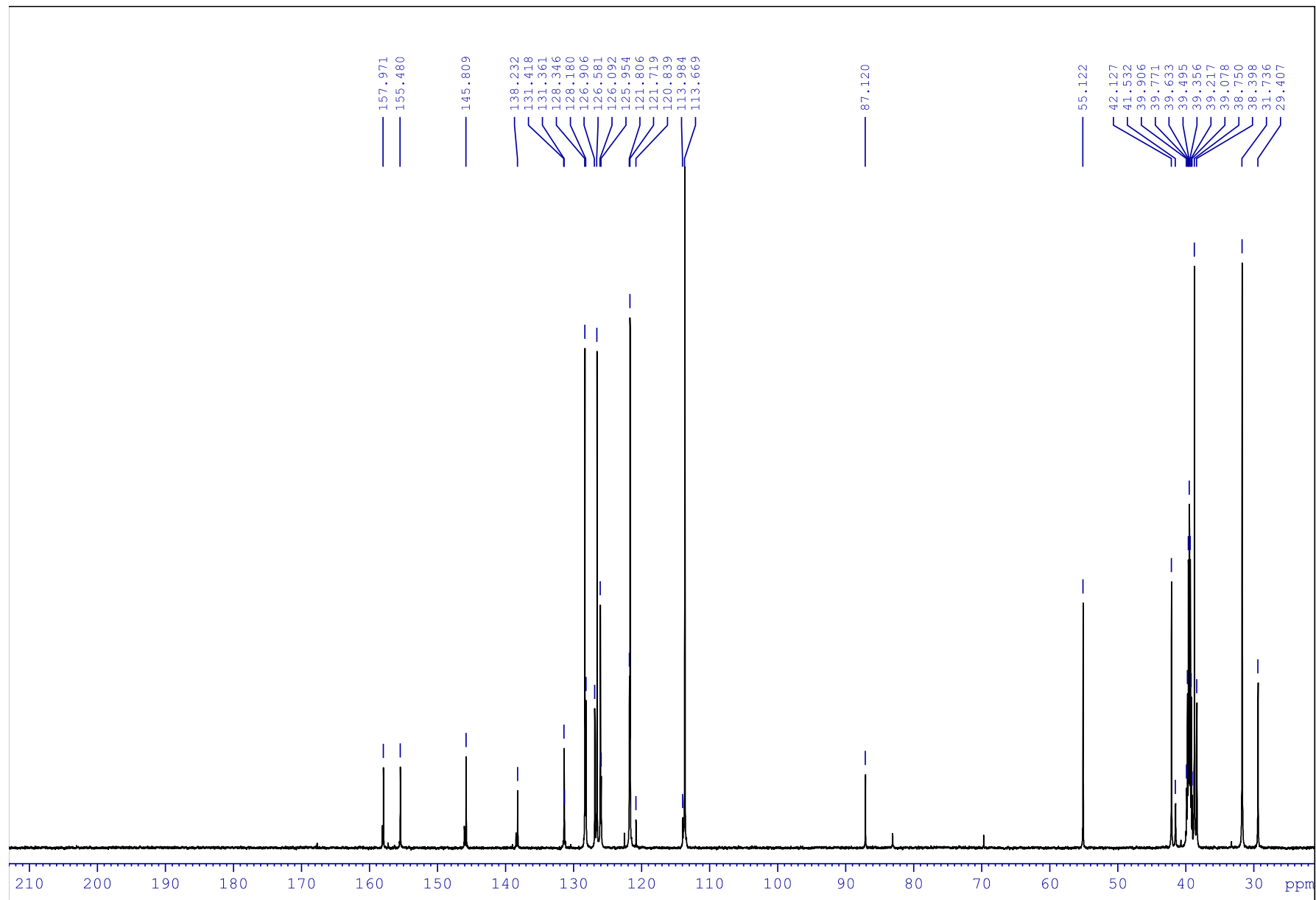
## 1. Experimental Section

$^1\text{H}$ ,  $^{13}\text{C}$ ,  $^{19}\text{F}$  NMR, 1D NOESY NMR, 2D NMR HSQC, HMBC and ROESY experiments were recorded on Bruker AV-600 (600, 565 and 151 MHz, for  $^1\text{H}$ ,  $^{19}\text{F}$  and  $^{13}\text{C}$  respectively), Bruker AV-400 (400 and 100.6 MHz, respectively) and Bruker AM-300 (300, 282 and 75 MHz, for  $^1\text{H}$ ,  $^{19}\text{F}$  and  $^{13}\text{C}$  respectively). The chemical shifts ( $\delta$ ) were expressed in ppm and referenced to DMSO- $d_6$  (39.5 ppm) for  $^1\text{H}$  and  $^{13}\text{C}$  NMR, respectively. The coupling constants ( $J$ ) are in Hertz. The assignment of the signals in the NMR spectra was based on the 2D NMR data. IR spectra were recorded on a Bruker Alpha spectrometer as KBr pellets, significant band ( $\nu$ ) reported in  $\text{cm}^{-1}$ . High-resolution mass spectra were obtained on a Bruker MicroTOF mass spectrometer by electrospray ionization (ESI) using Q-TOF detection. The melting points were determined on a Kofler hot stage apparatus and are uncorrected. TLC was performed using Silicagel 60 F254 plates. The chromatograms were visualized with an UV lamp (254 and 365 nm) and  $[\text{Ce}(\text{SO}_4)_2/\text{H}_2\text{SO}_4]$  developing solution. Column chromatography was carried out on silica gel 60 (0.063–0.200 mm, Merck). Commercial reagents were used without further purification.

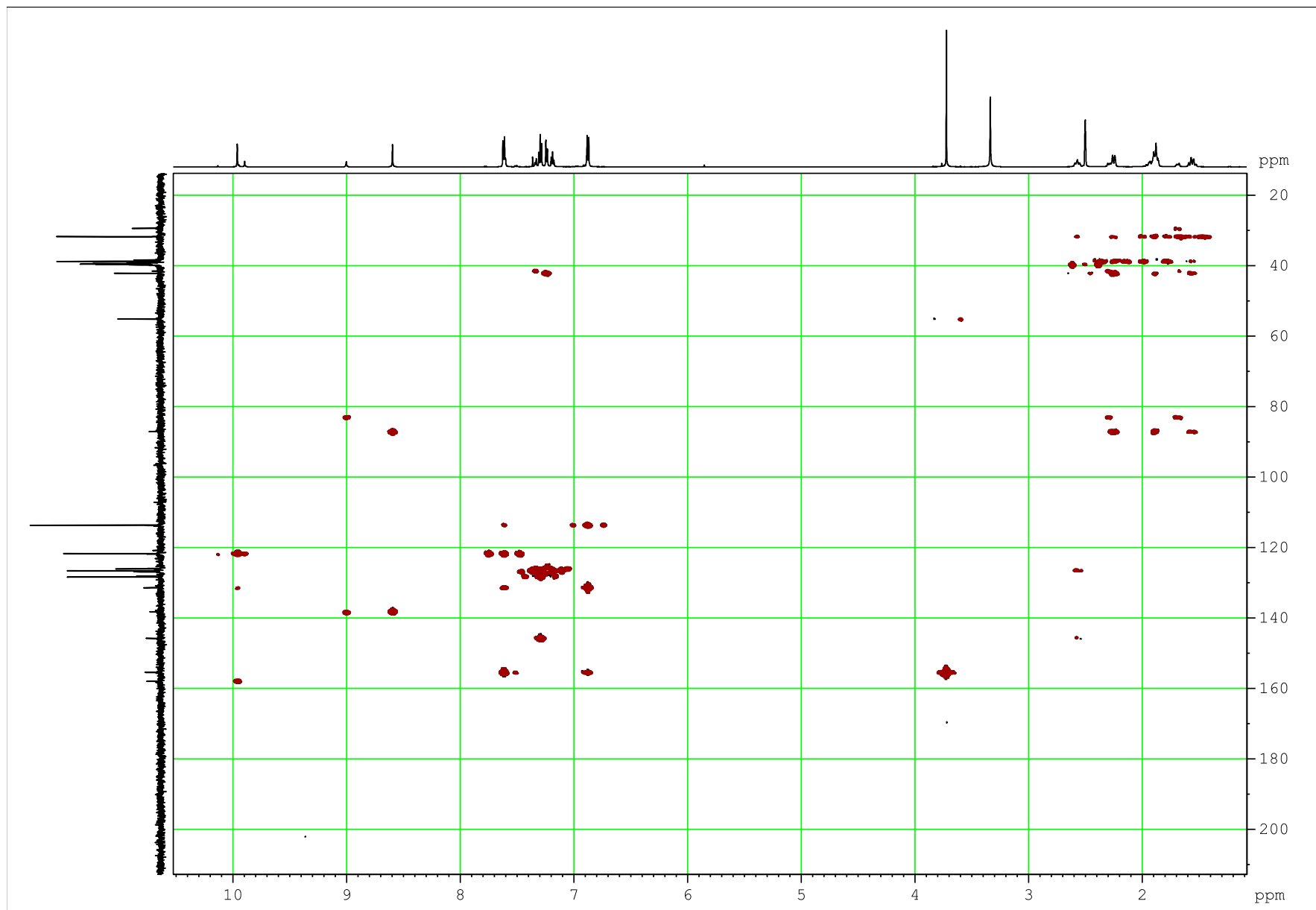
## 2. NMR spectra (Bruker AV-600)



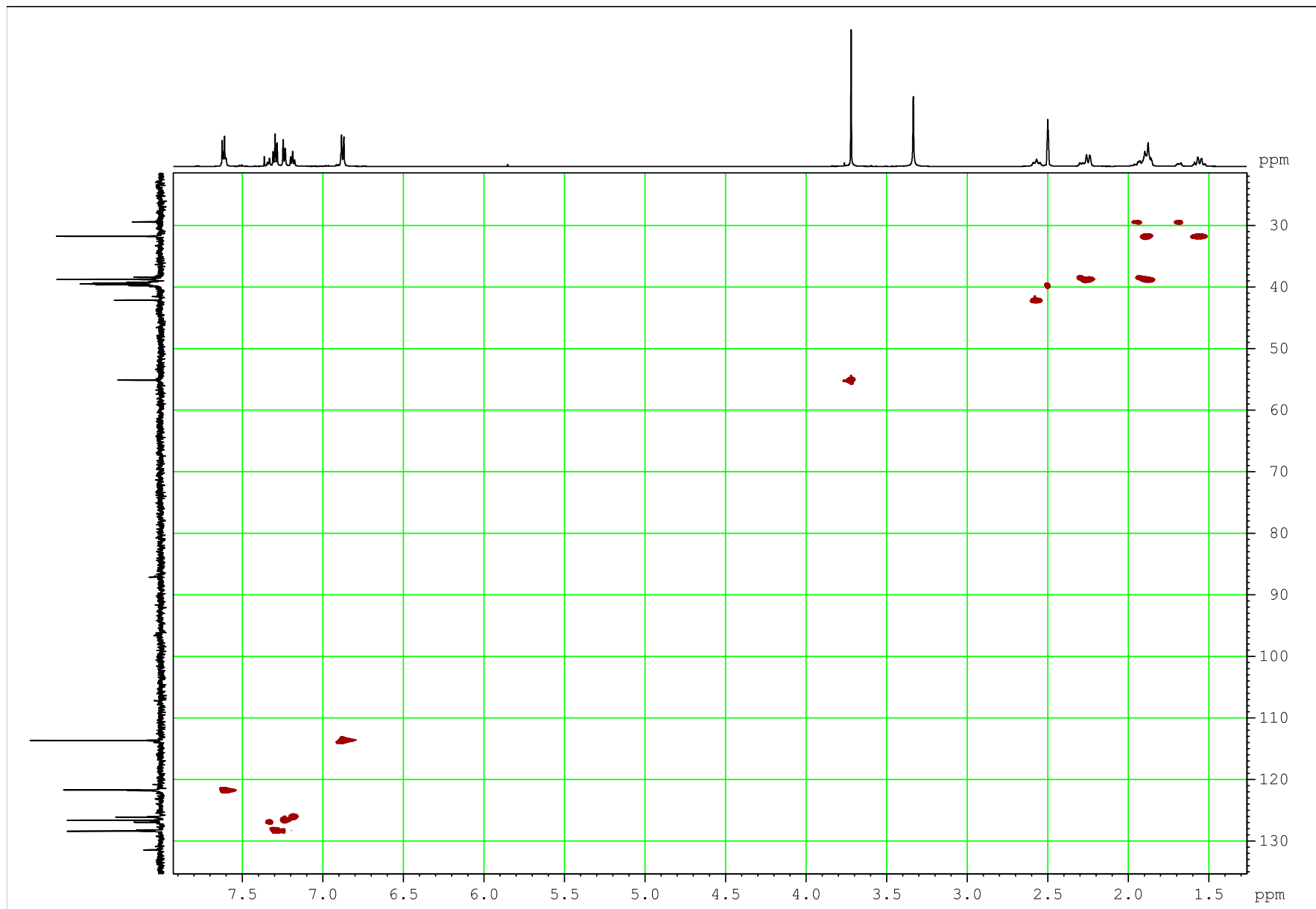
<sup>1</sup>H NMR spectrum of **3b** (DMSO-*d*<sub>6</sub>).



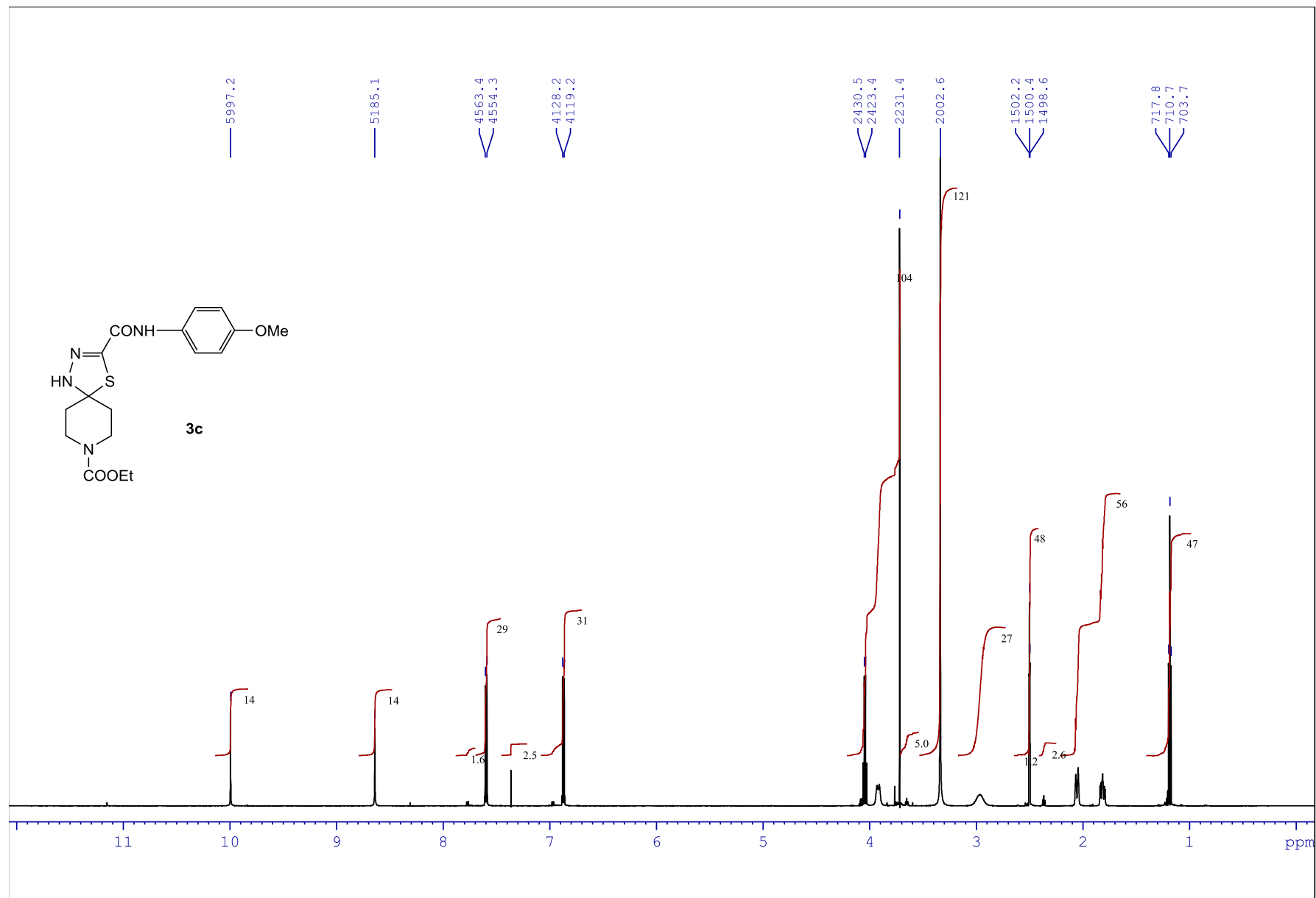
$^{13}\text{C}$  NMR spectrum of **3b** ( $\text{DMSO-}d_6$ ).



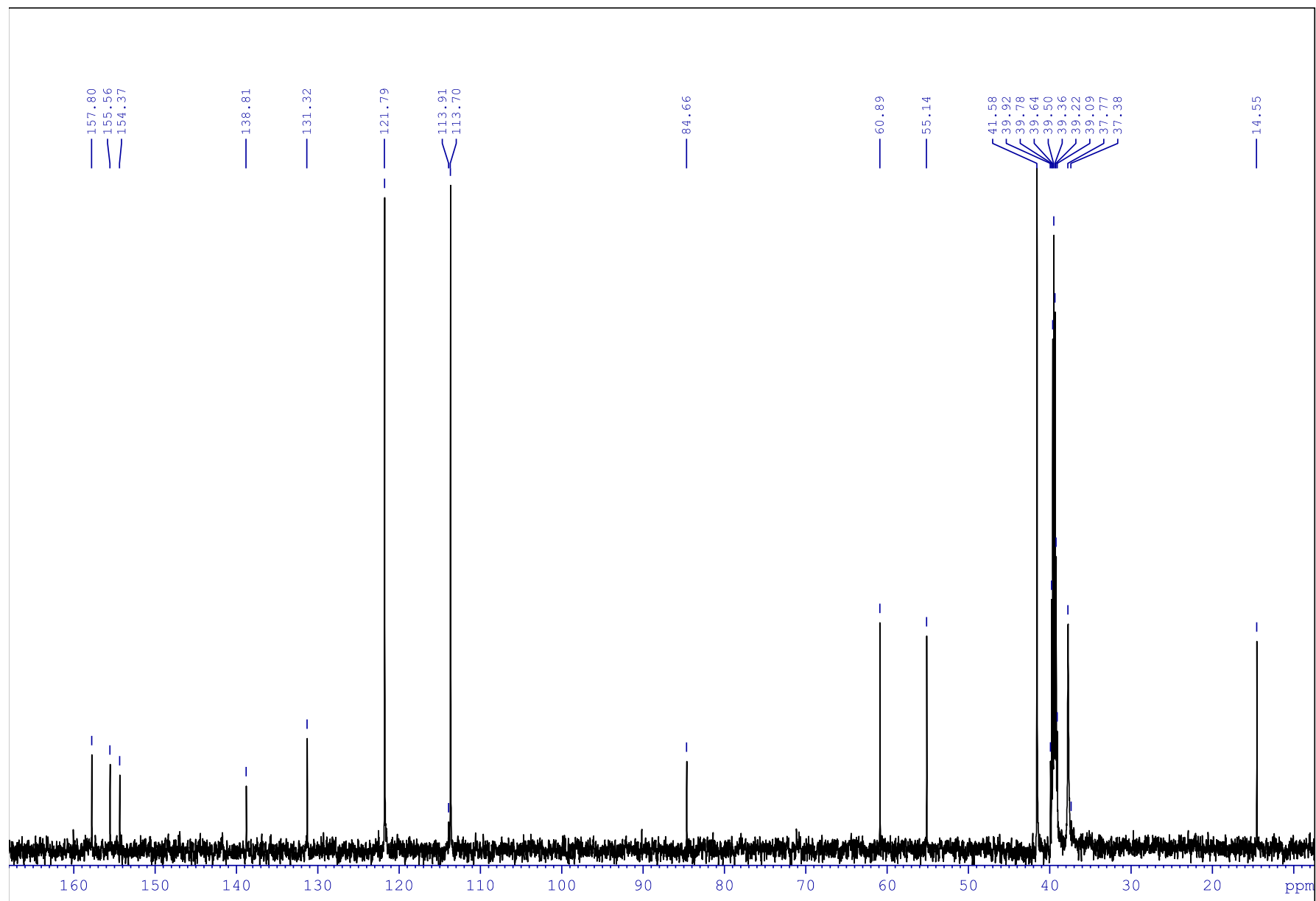
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3b** ( $\text{DMSO-}d_6$ ).



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3b** ( $\text{DMSO-}d_6$ ).

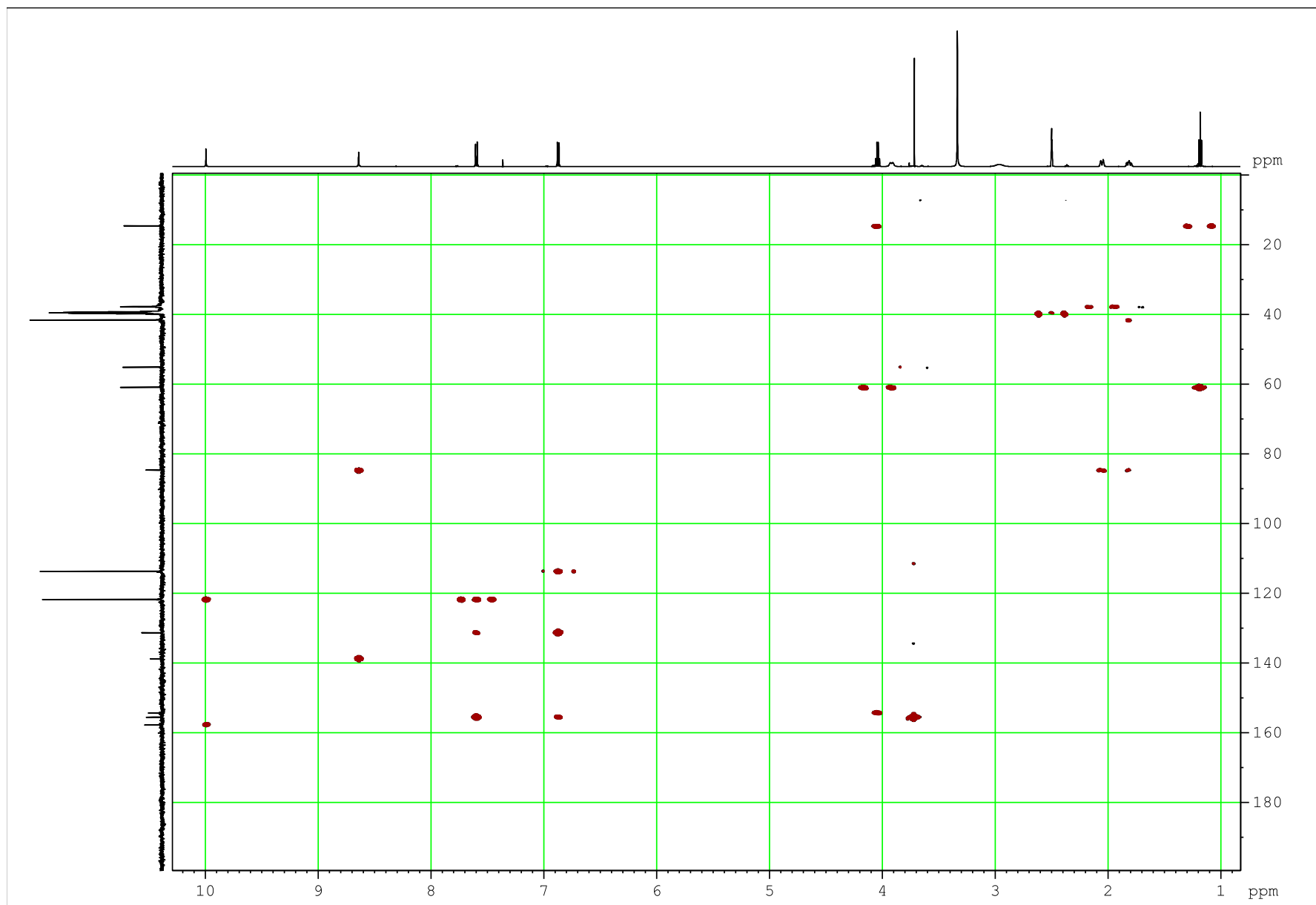


<sup>1</sup>H NMR spectrum of **3c** (DMSO-*d*<sub>6</sub>).

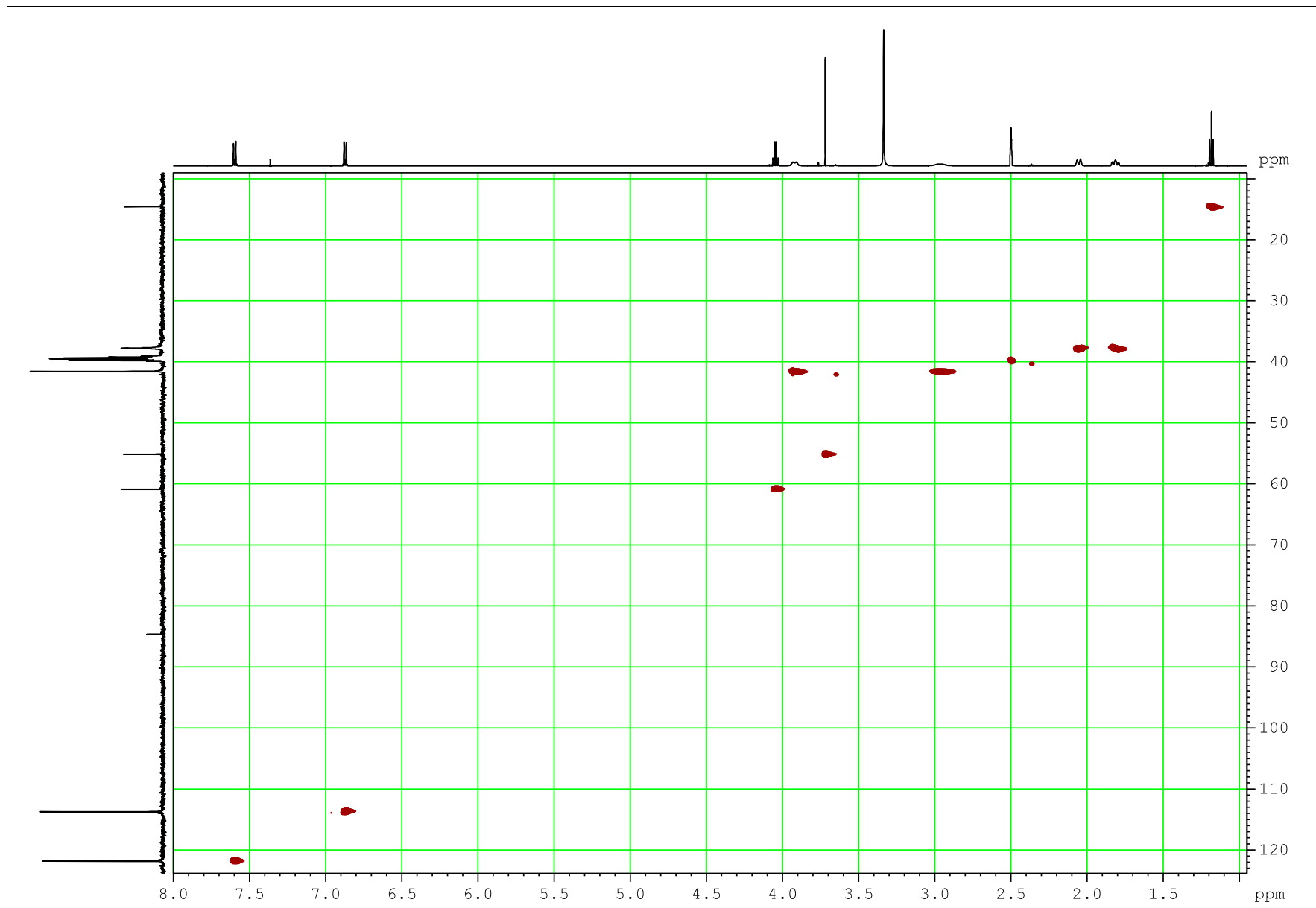


$^{13}\text{C}$  NMR spectrum of **3c** ( $\text{DMSO-}d_6$ ).

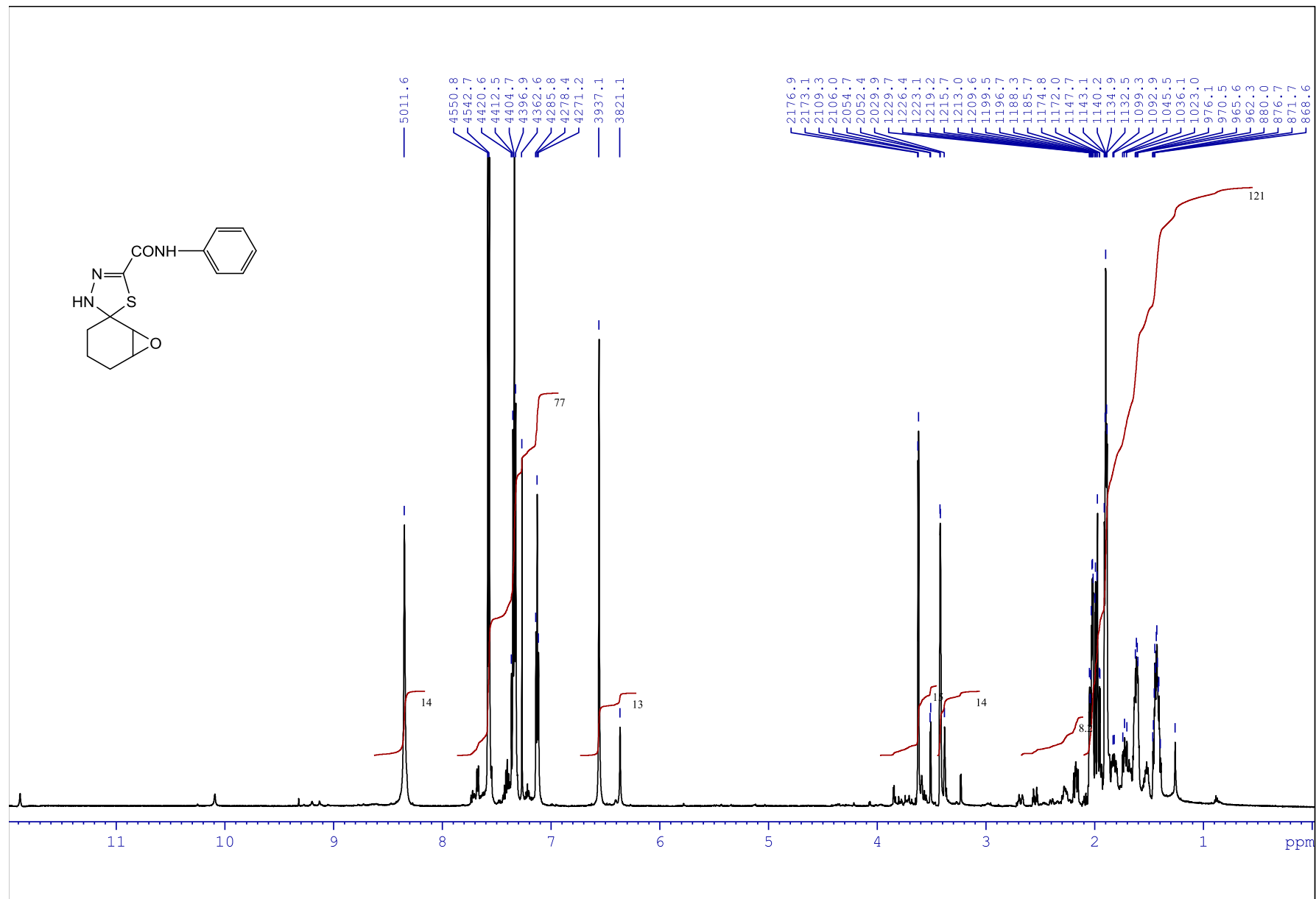




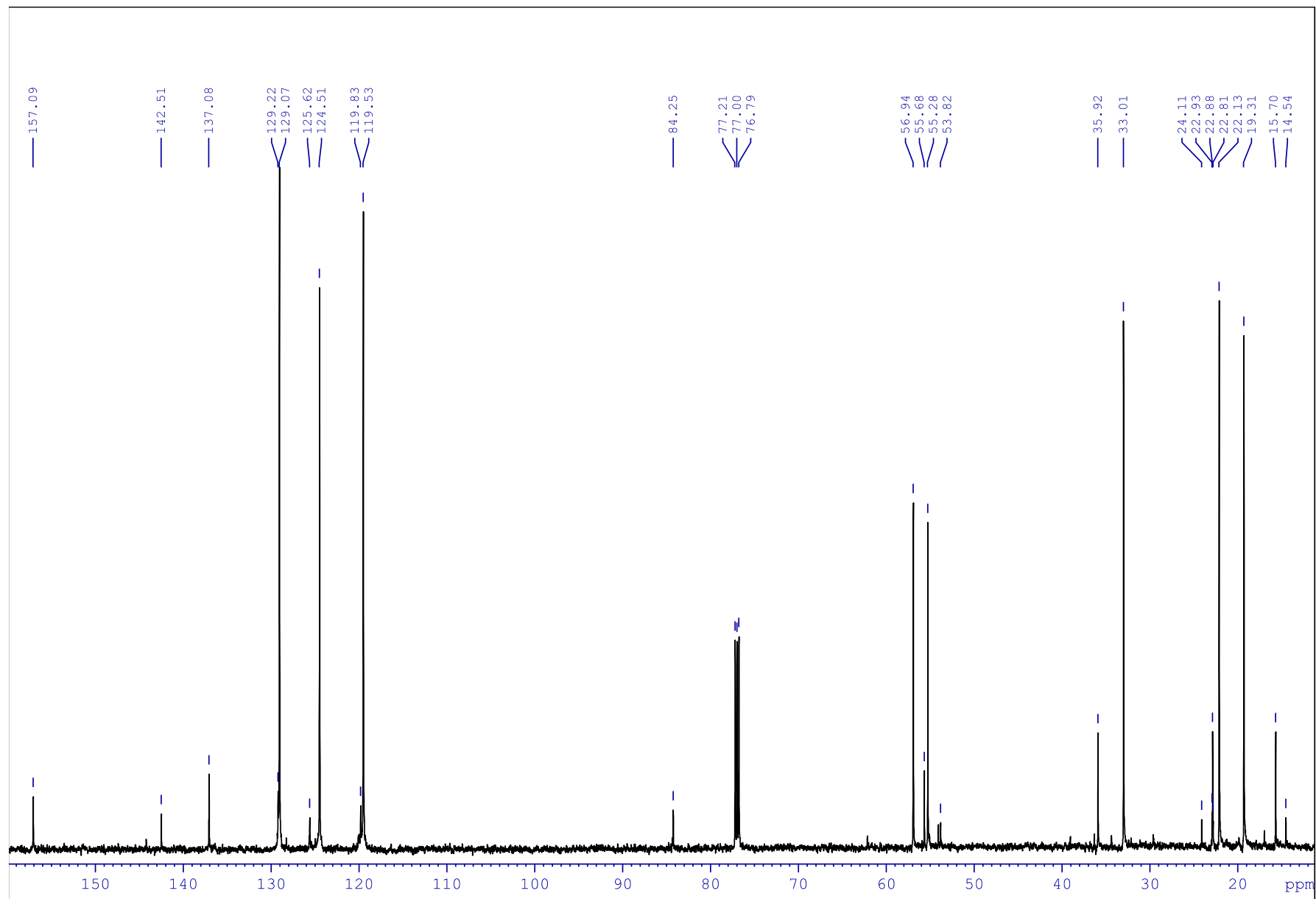
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3c** ( $\text{DMSO-}d_6$ ).



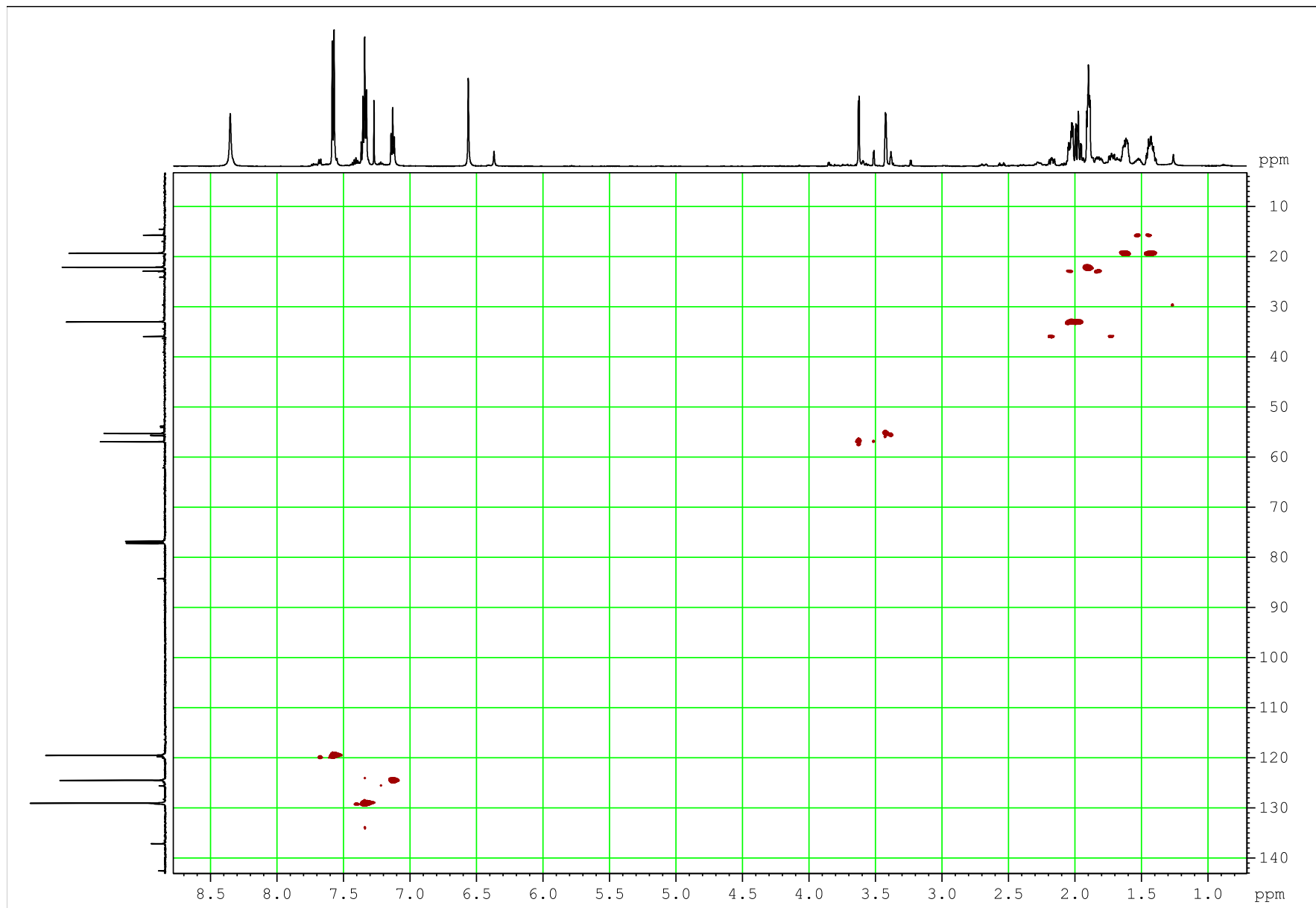
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3c** (DMSO- $d_6$ ).



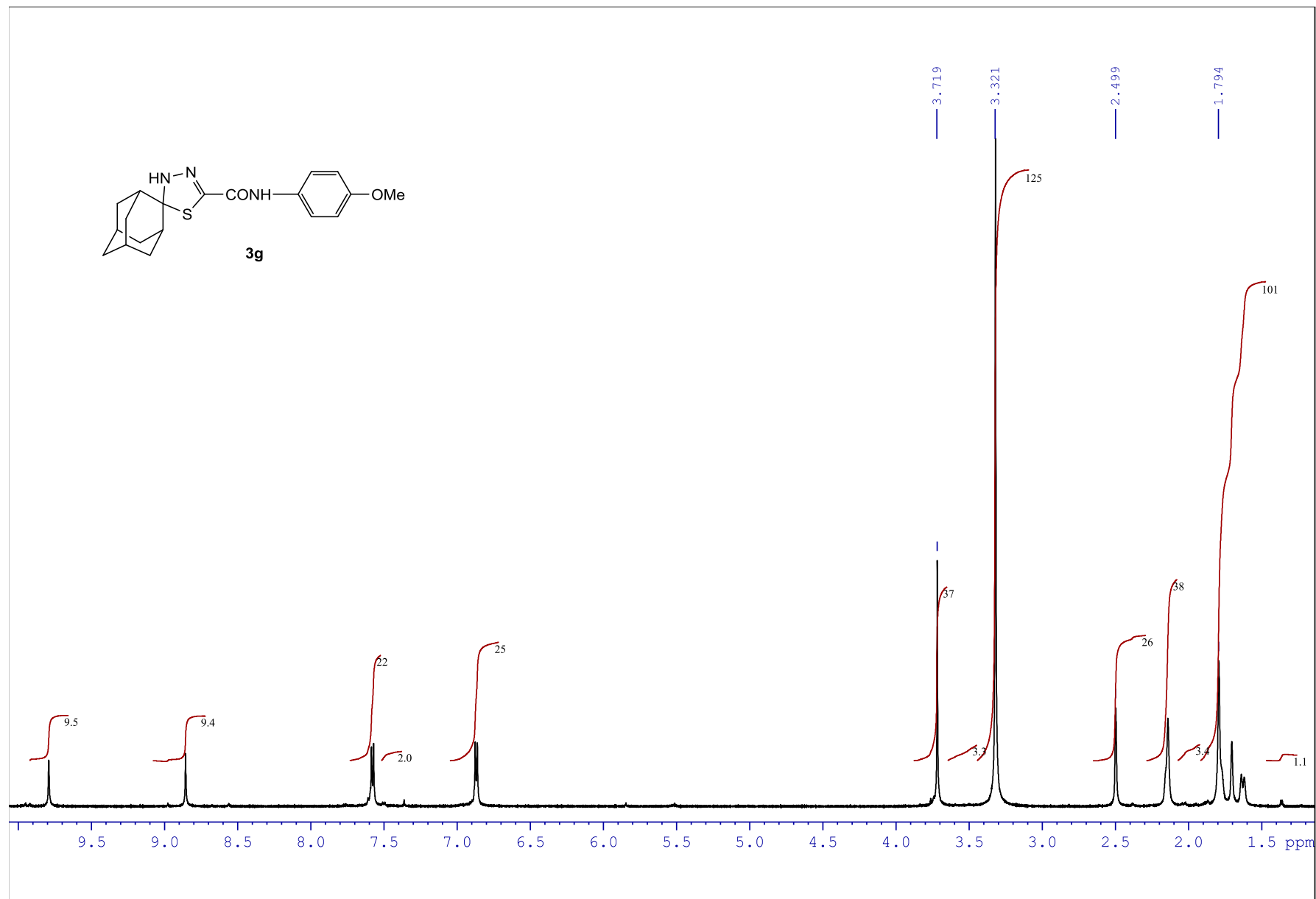
<sup>1</sup>H NMR spectrum of **3f** (CDCl<sub>3</sub>).



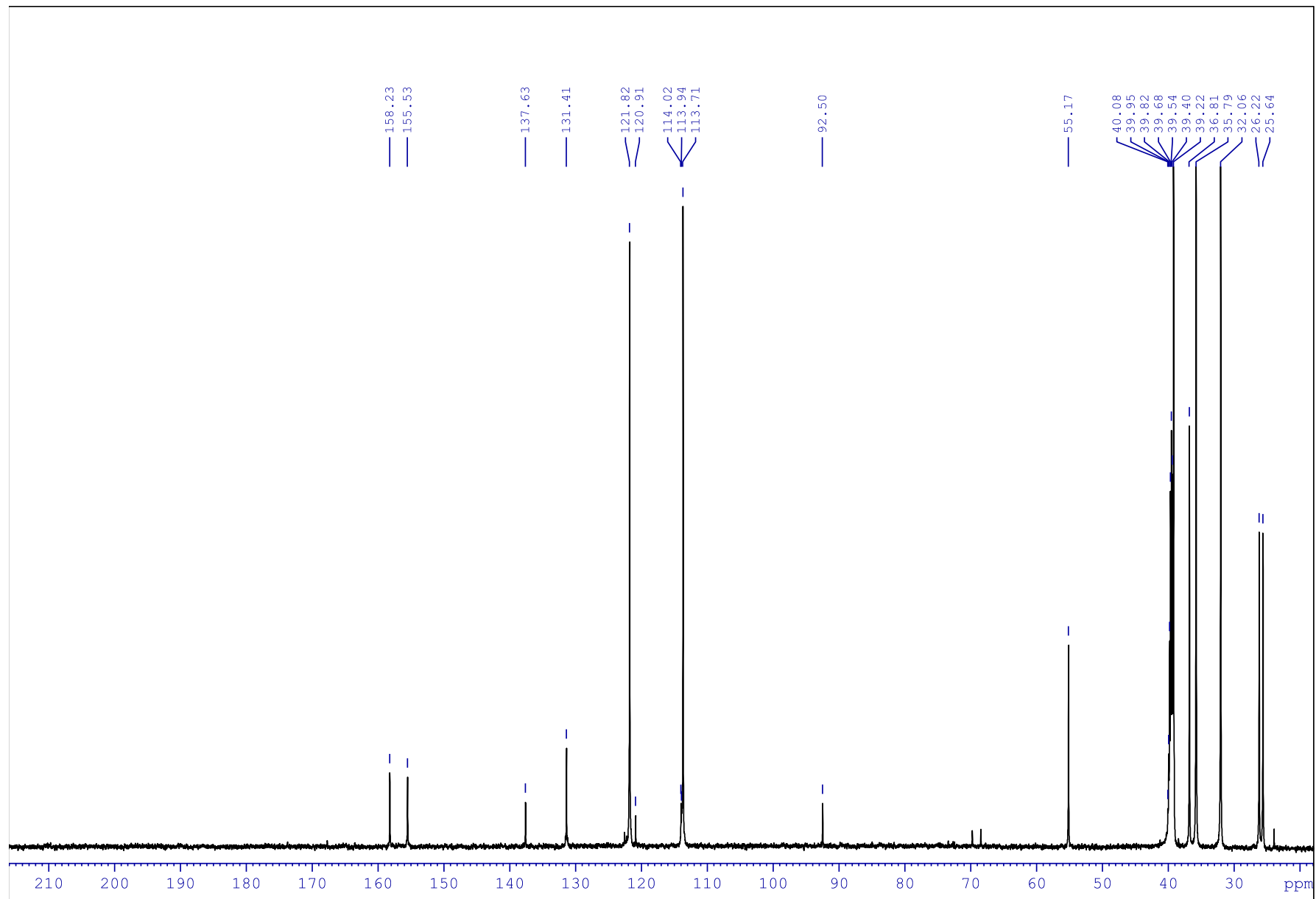
$^{13}\text{C}$  NMR spectrum of **3f** ( $\text{CDCl}_3$ ).



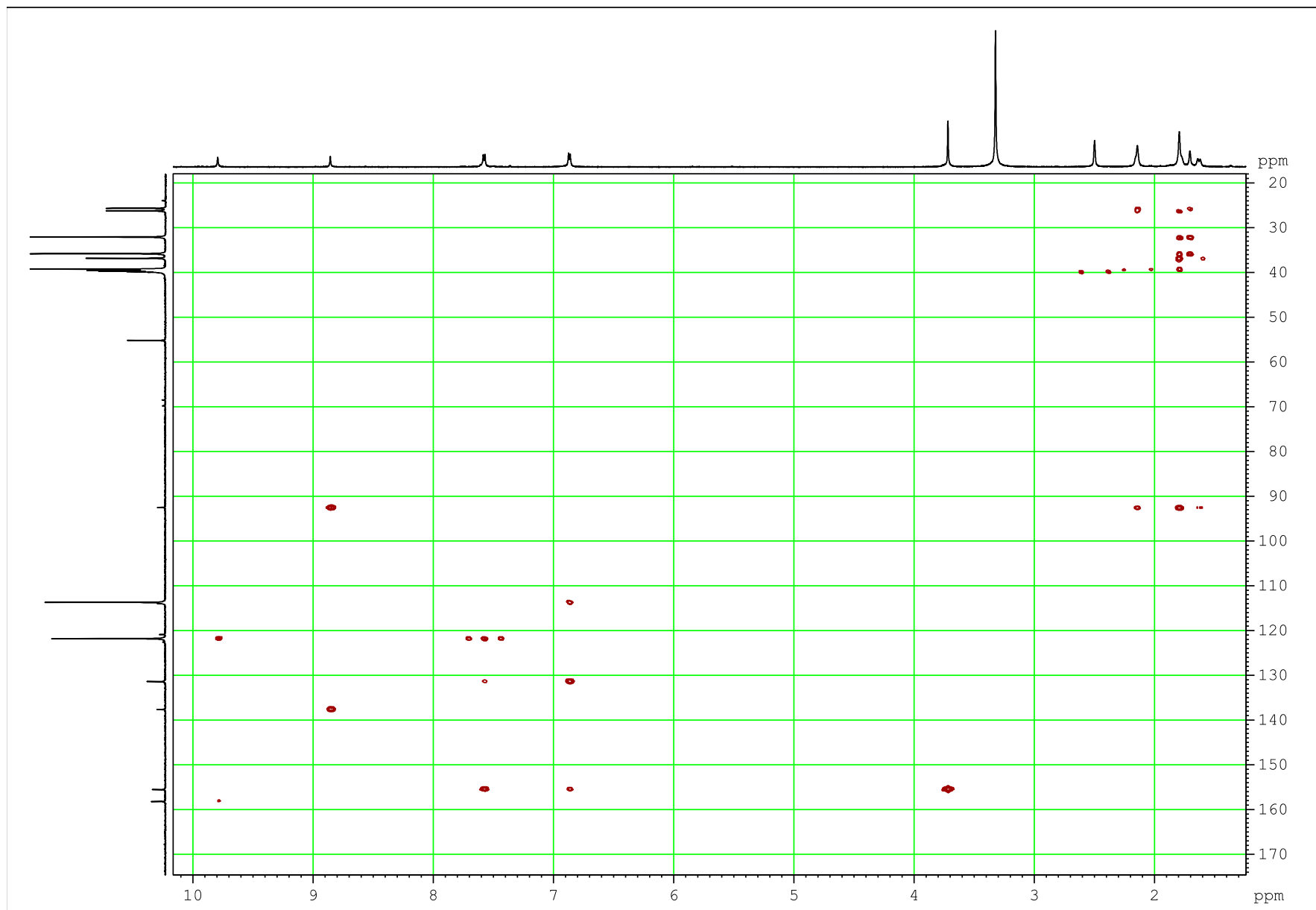
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3f** ( $\text{CDCl}_3$ ).



<sup>1</sup>H NMR spectrum of **3g** (DMSO-*d*<sub>6</sub>).

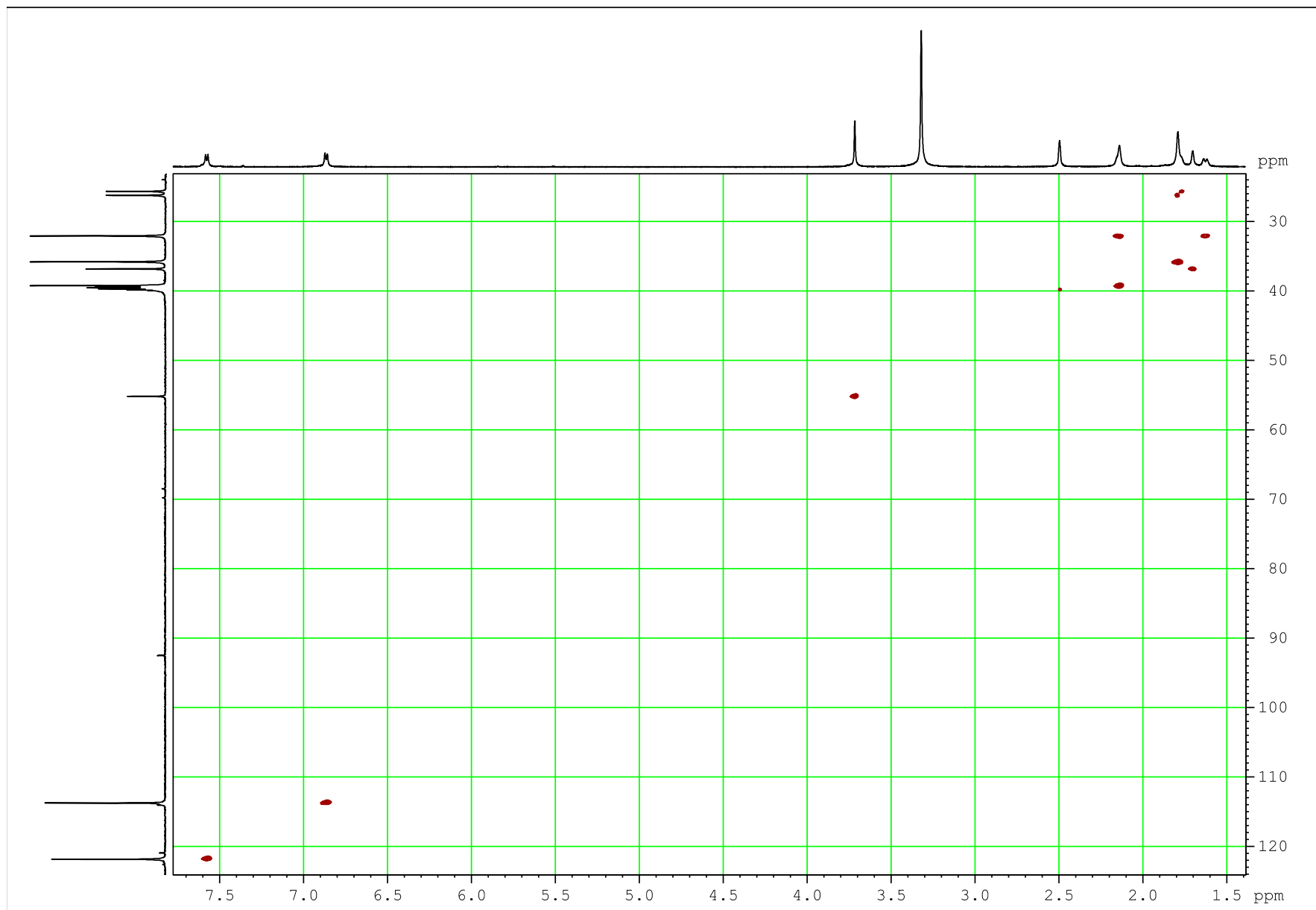


$^{13}\text{C}$  NMR spectrum of **3g** ( $\text{DMSO-}d_6$ ).

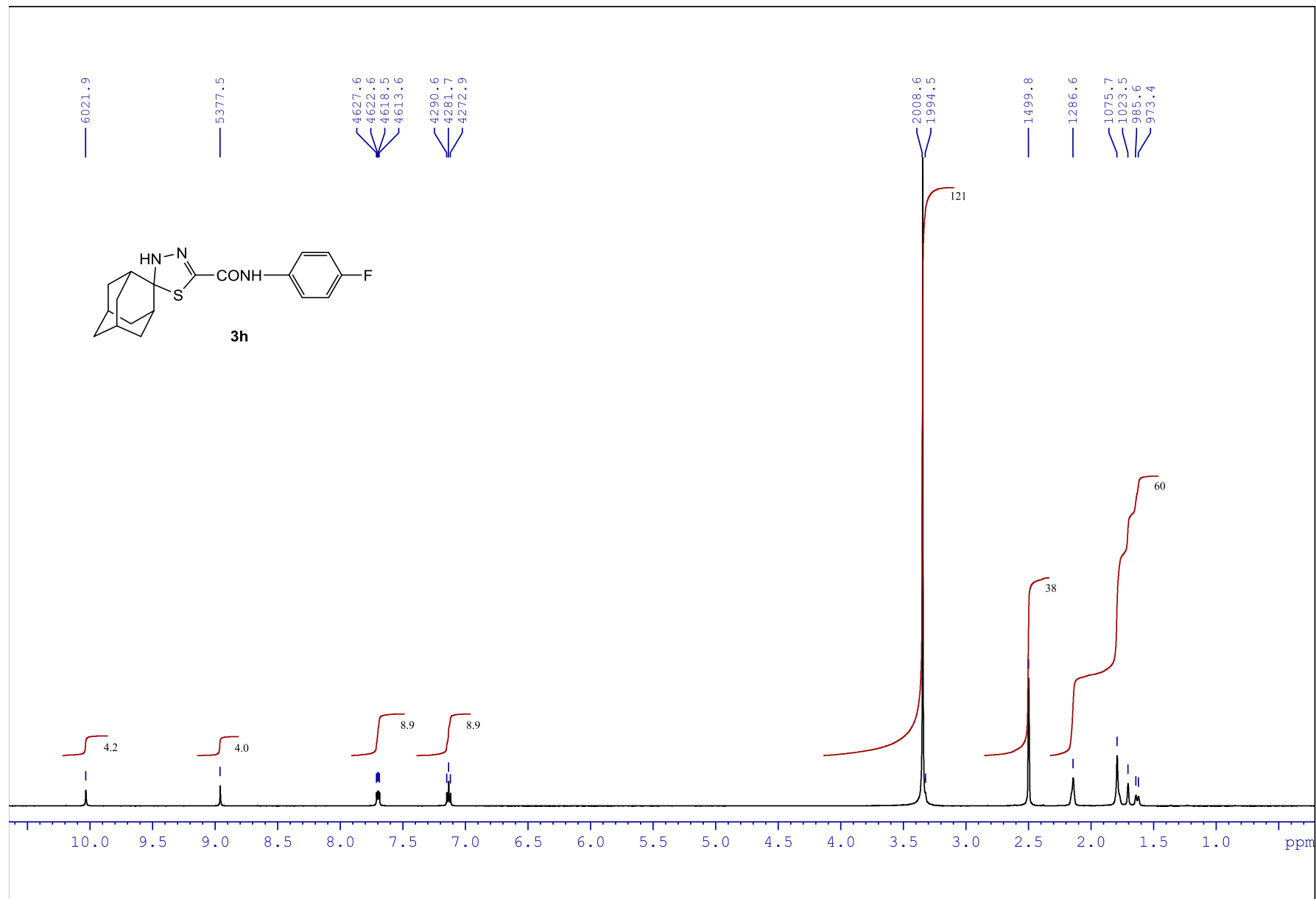


2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3g** ( $\text{DMSO-}d_6$ ).

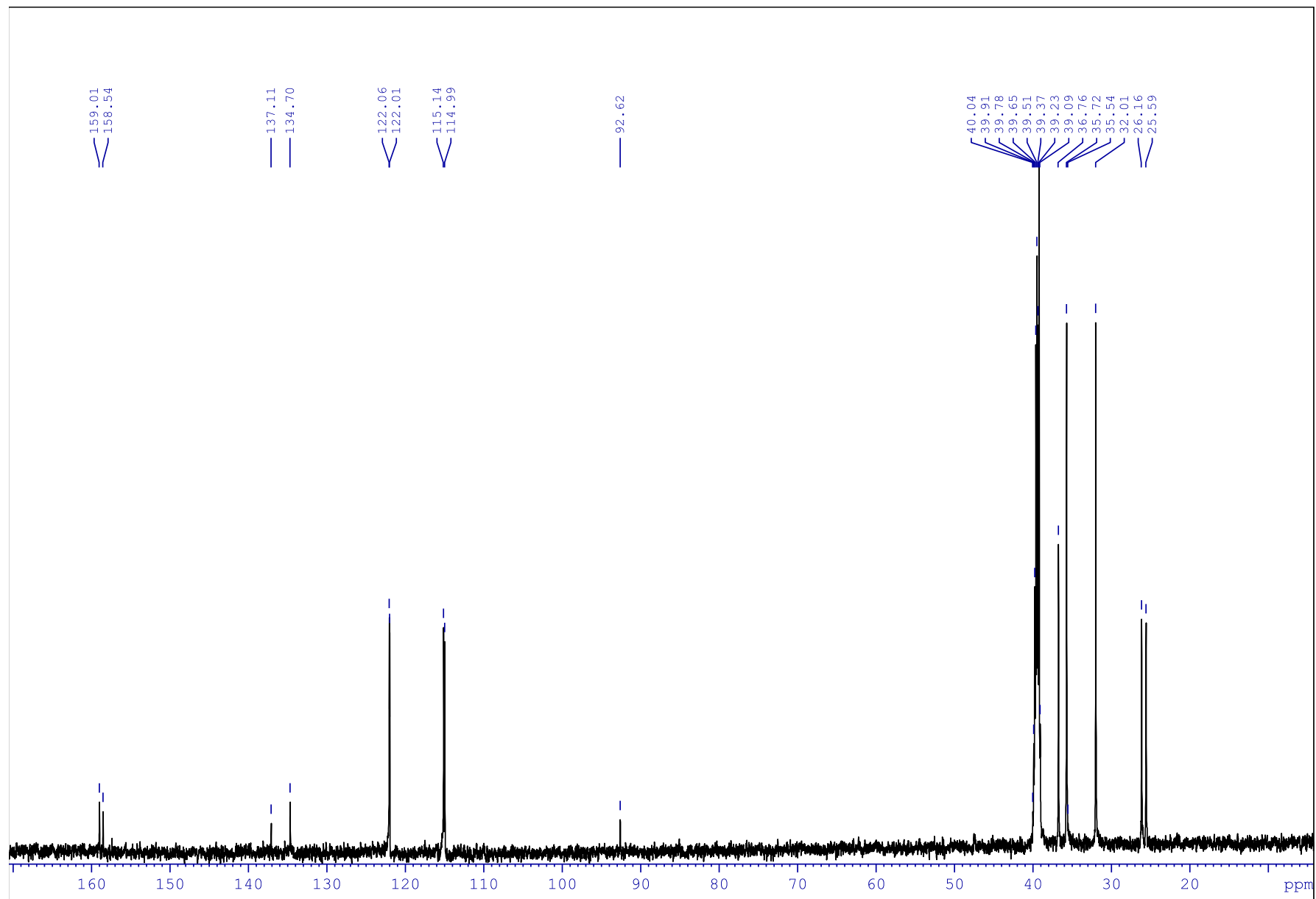




2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3g** (DMSO- $d_6$ ).

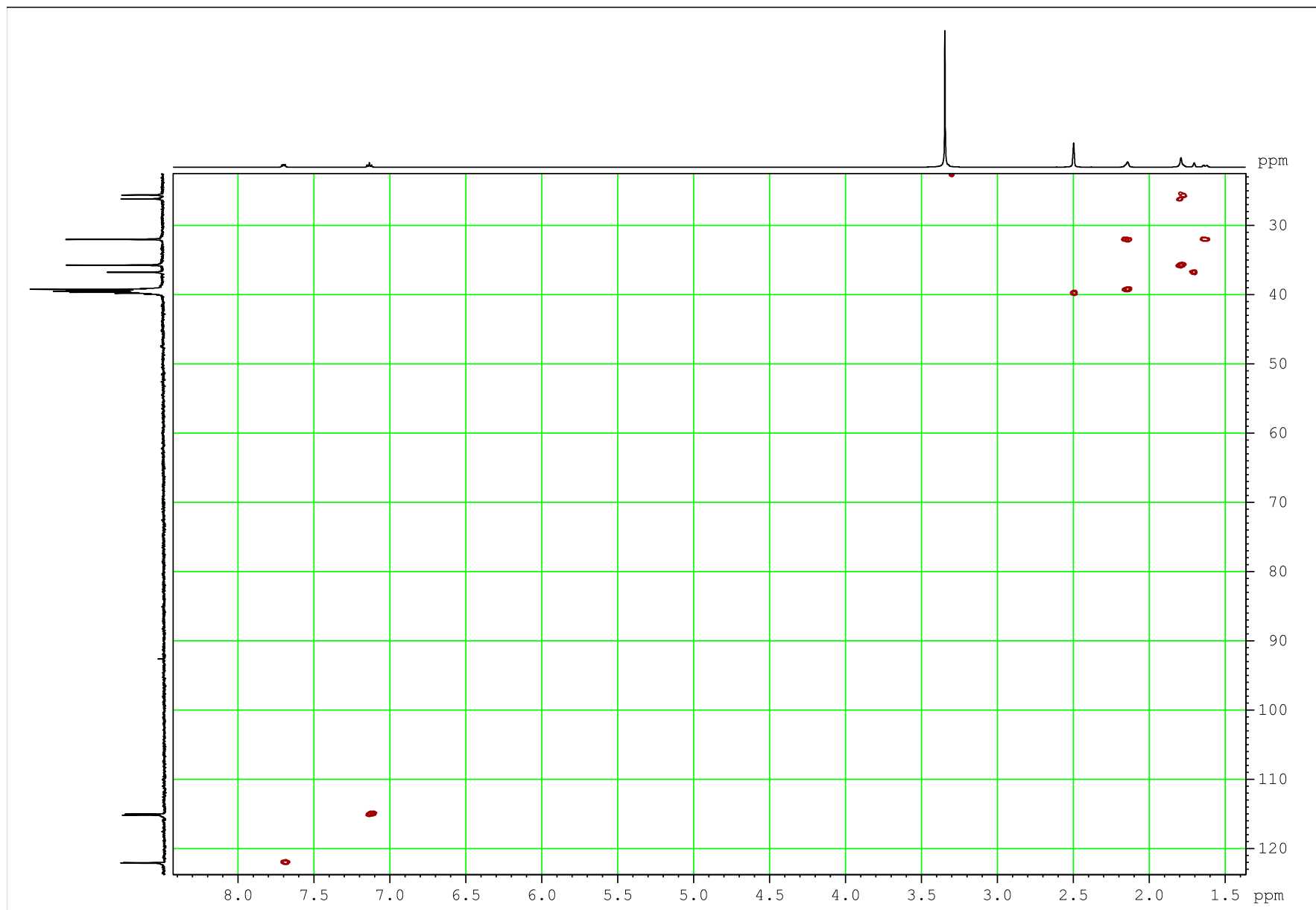


$^1\text{H}$  NMR spectrum of **3h** (DMSO- $d_6$ ).

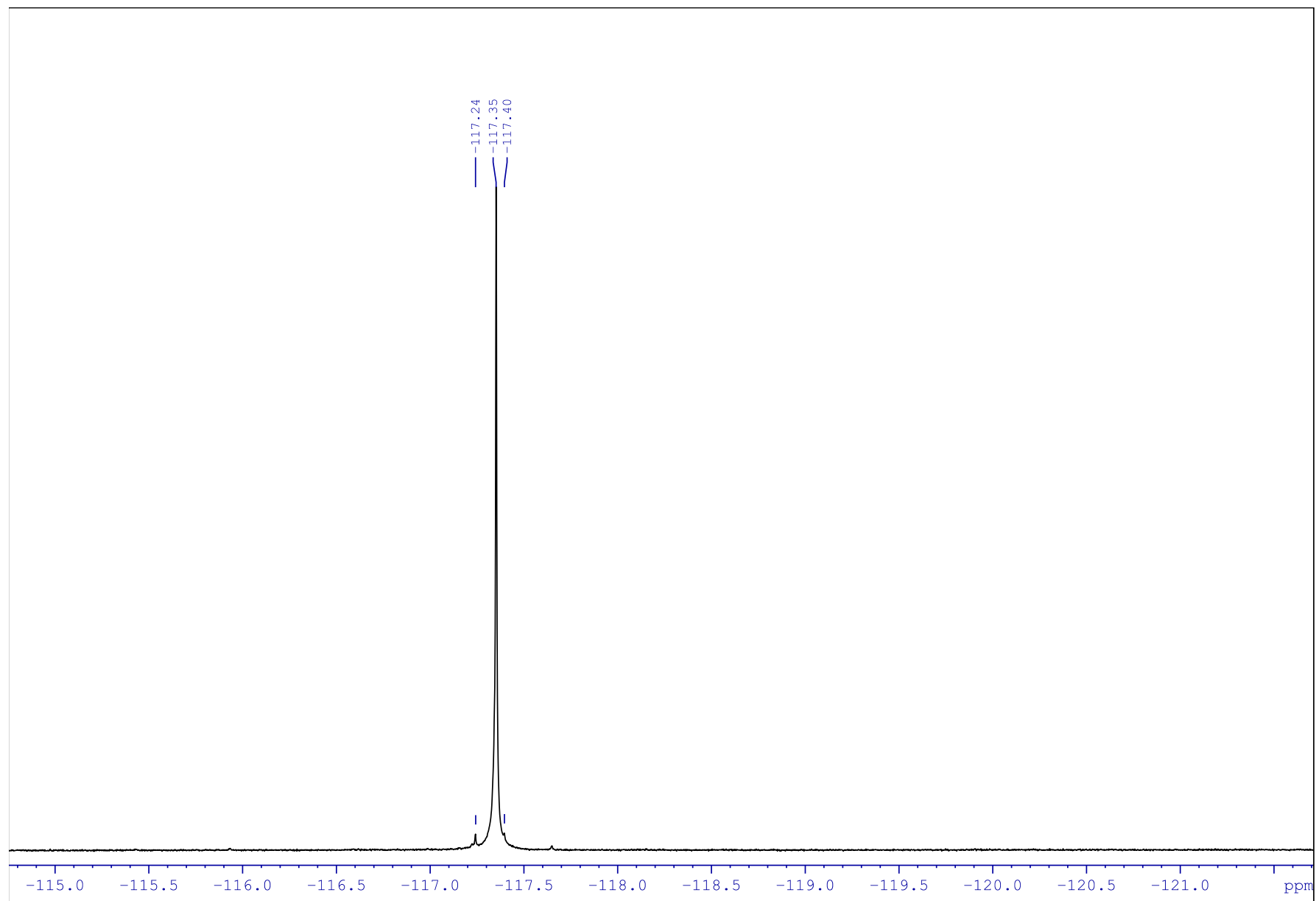


$^{13}\text{C}$  NMR spectrum of **3h** ( $\text{DMSO-}d_6$ ).

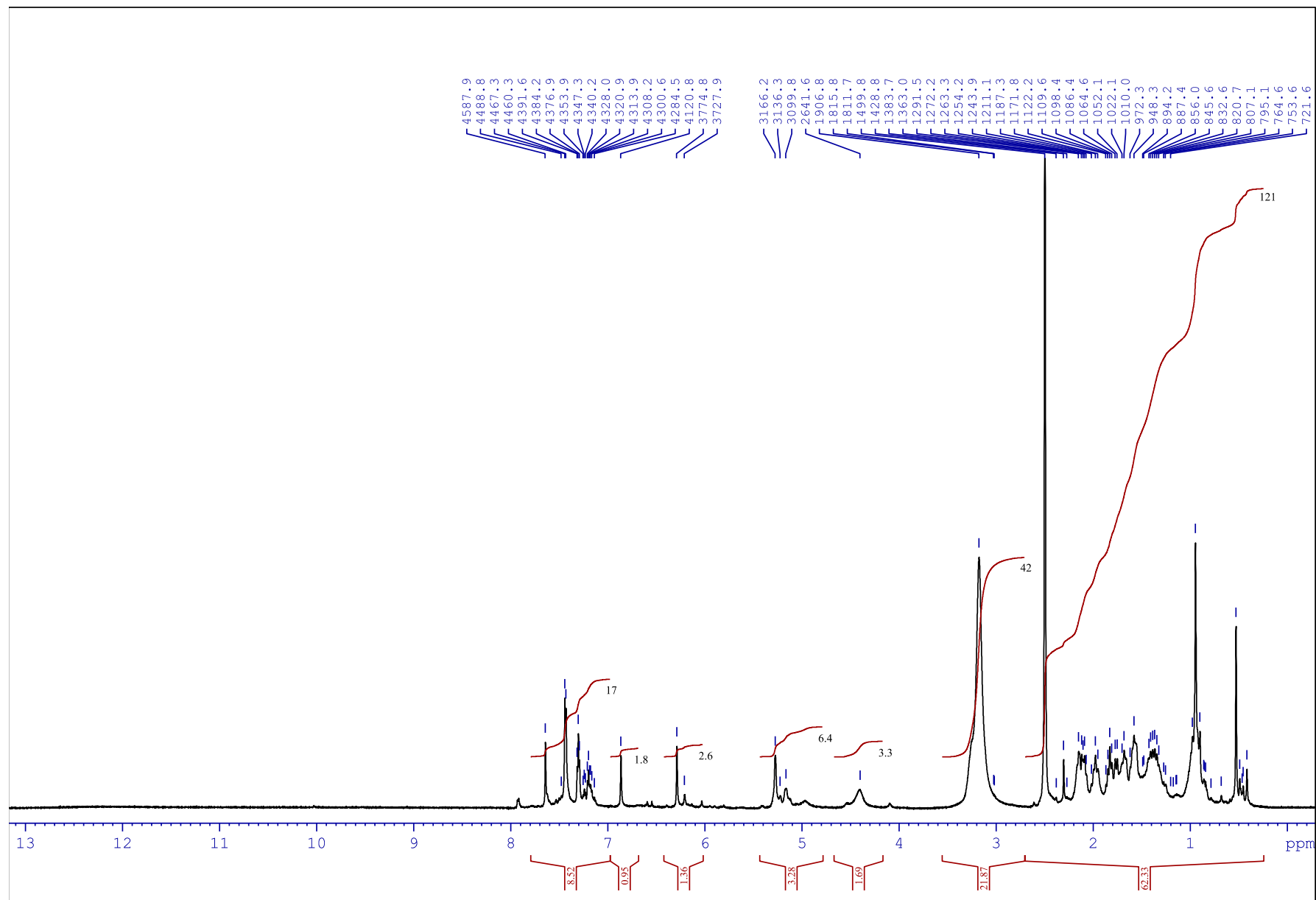




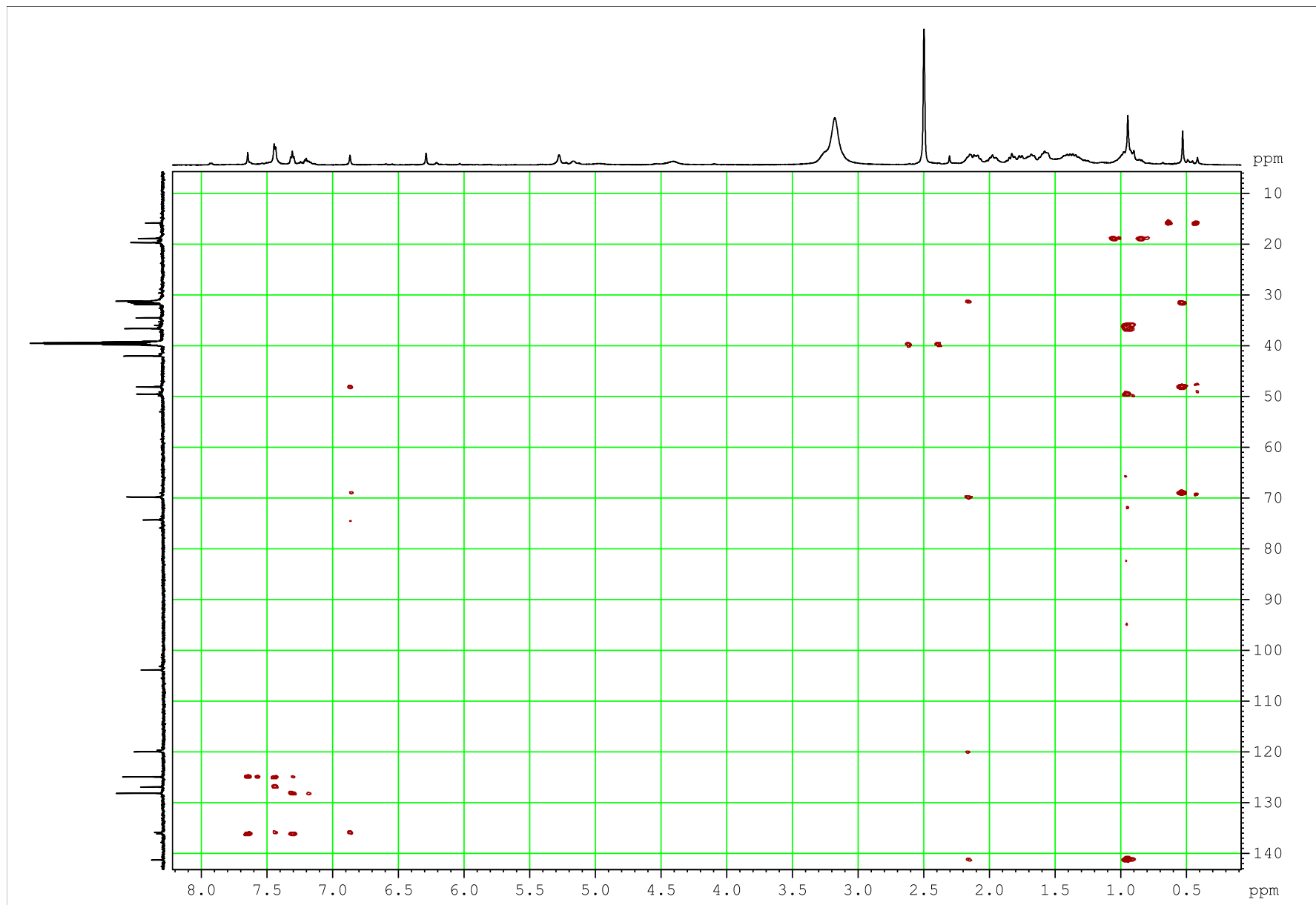
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3h** (DMSO- $d_6$ ).



$^{19}\text{F}$  NMR spectrum of **3h** ( $\text{DMSO-}d_6$ ).

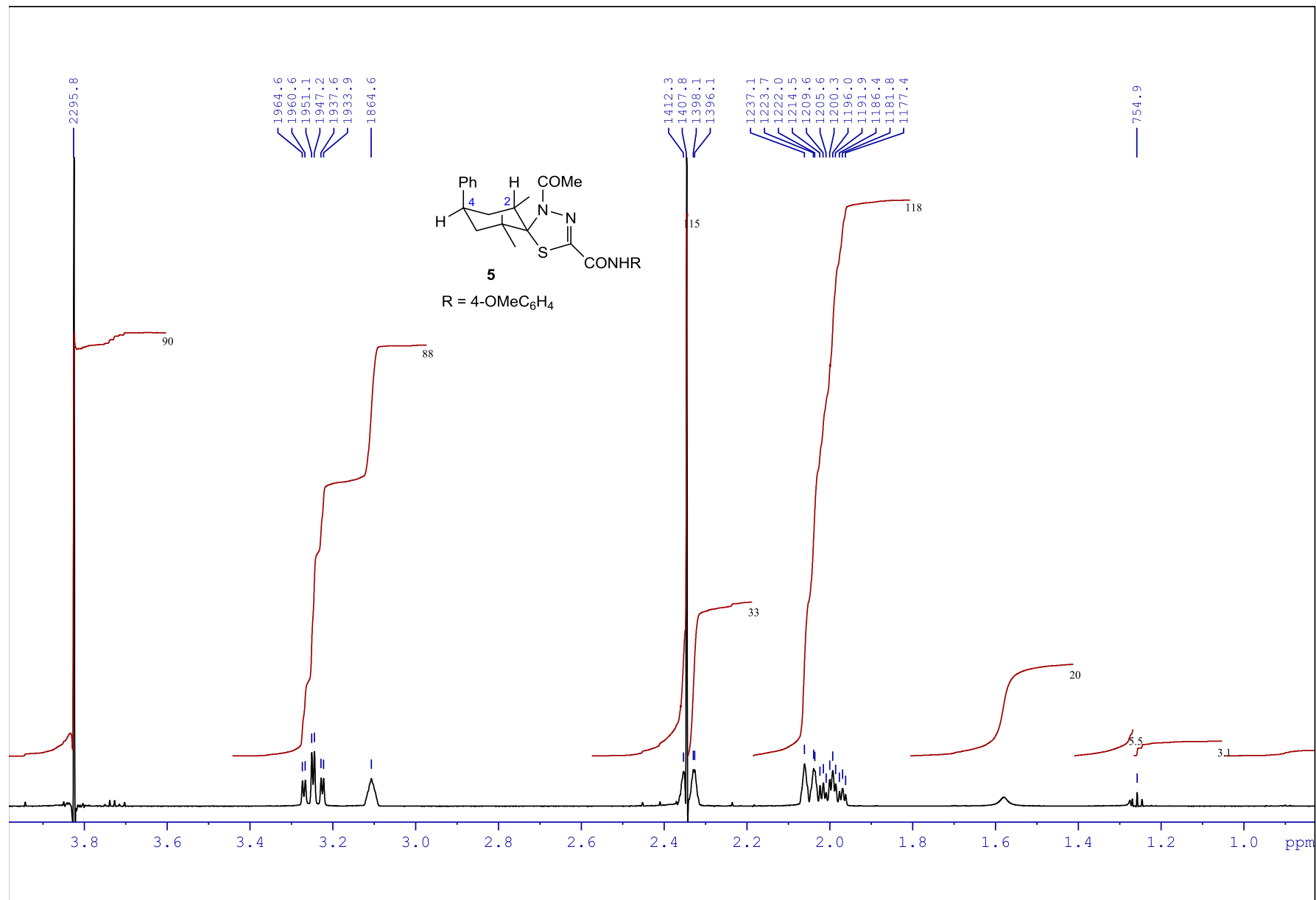


$^1\text{H}$  NMR spectrum of **3i** ( $\text{DMSO-}d_6$ ).

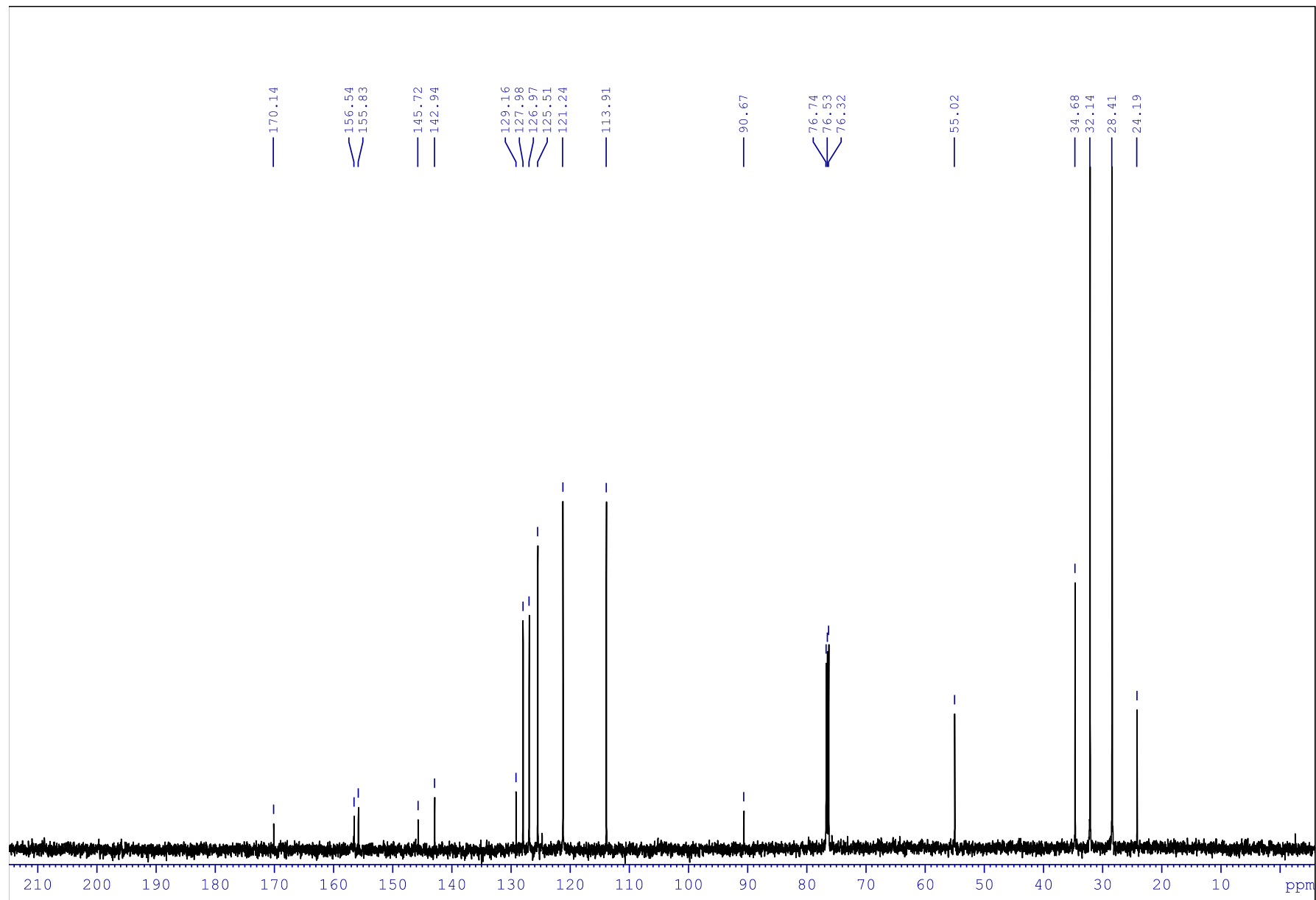


2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3i** ( $\text{DMSO}-d_6$ ).

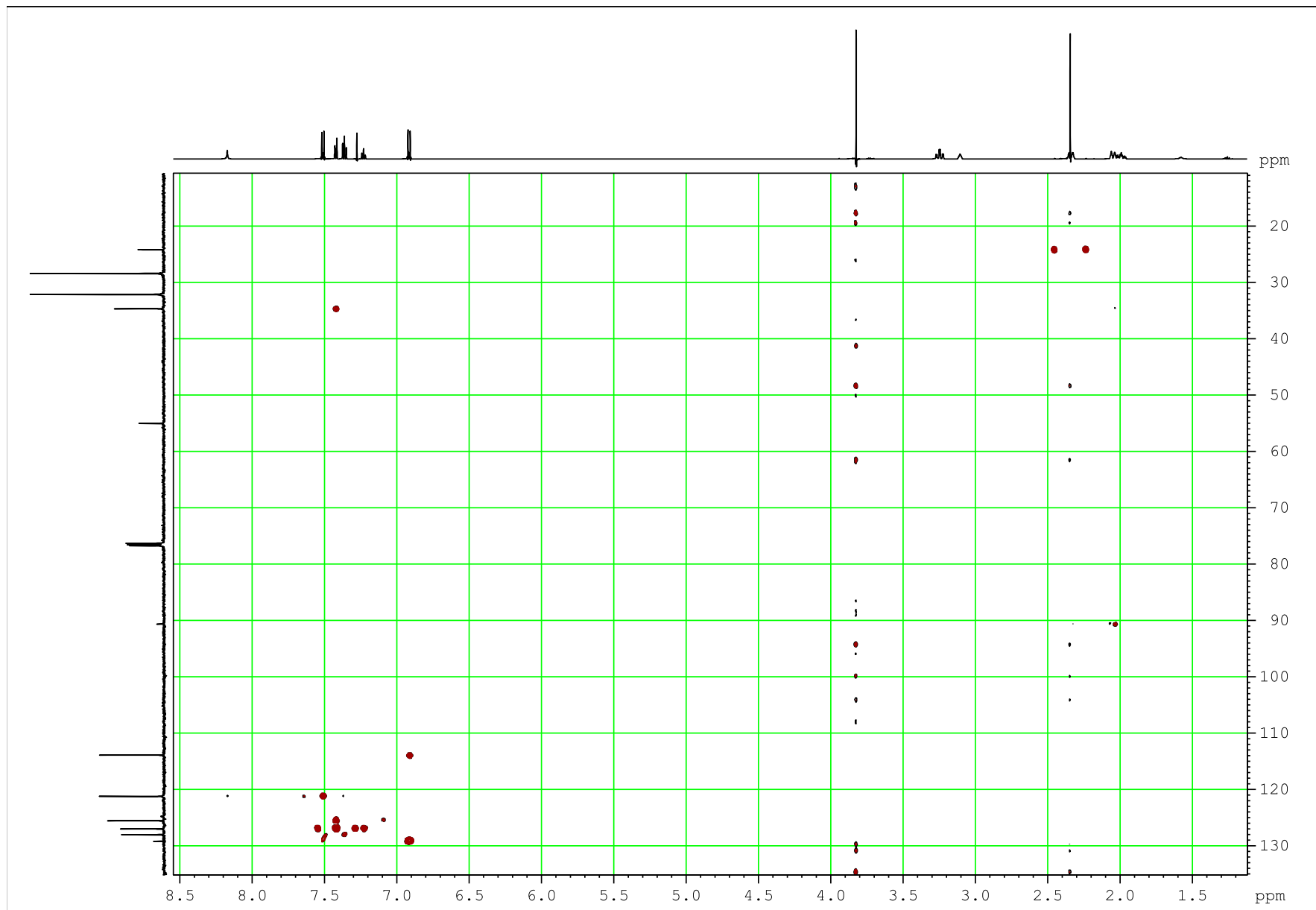




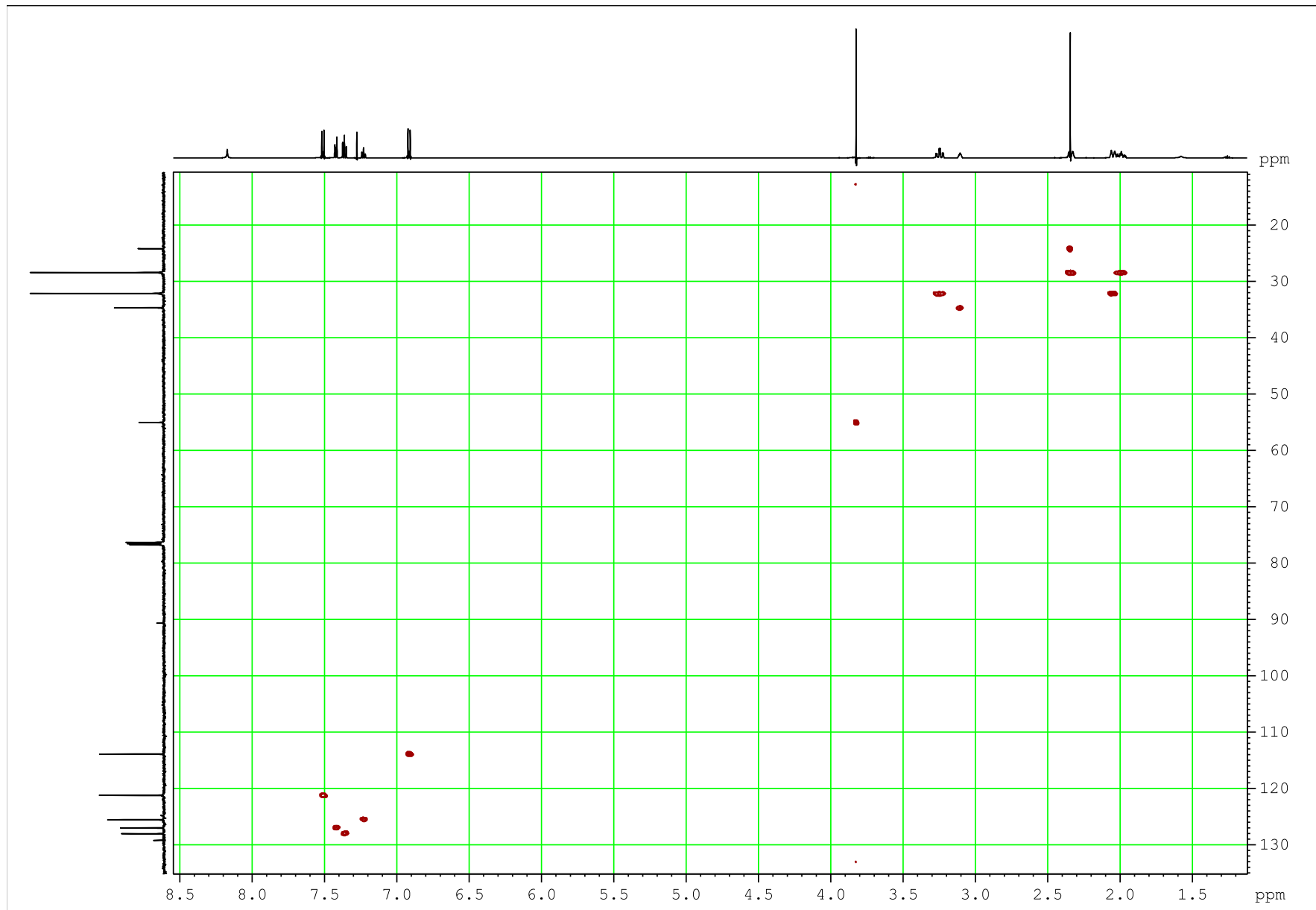
<sup>1</sup>H NMR spectrum of **5** (CDCl<sub>3</sub>).



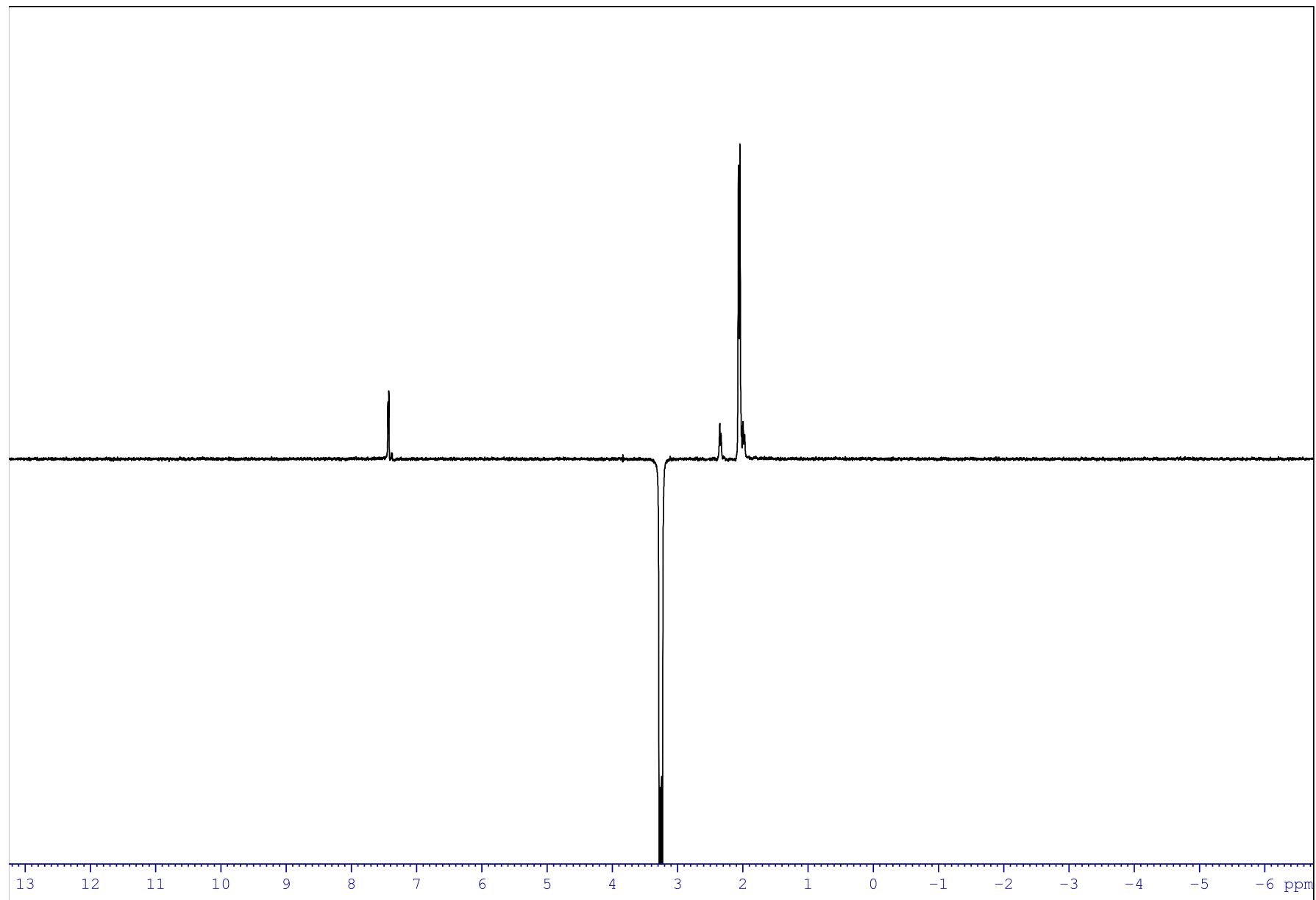
$^{13}\text{C}$  NMR spectrum of **5** ( $\text{CDCl}_3$ ).



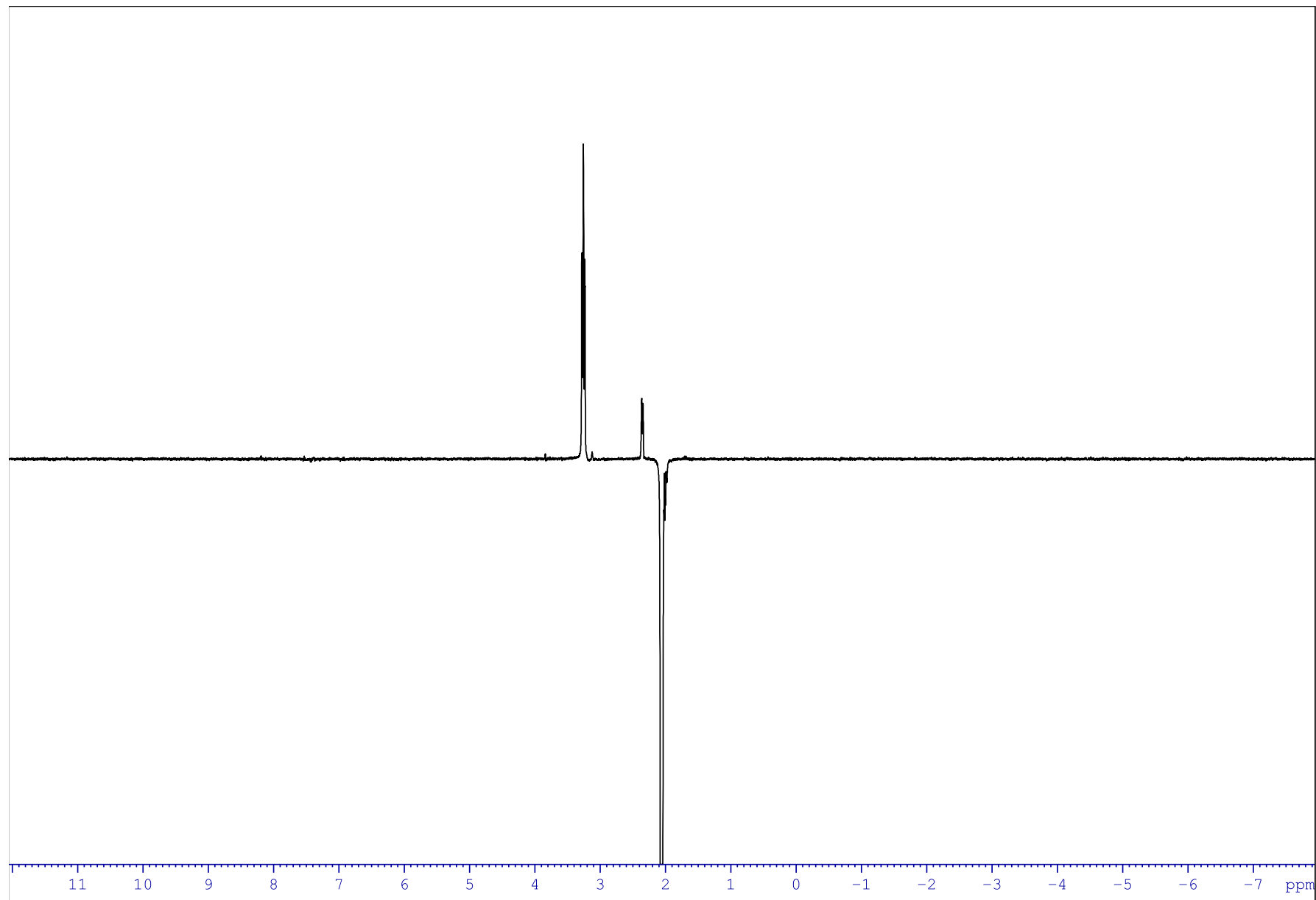
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **5** ( $\text{CDCl}_3$ ).



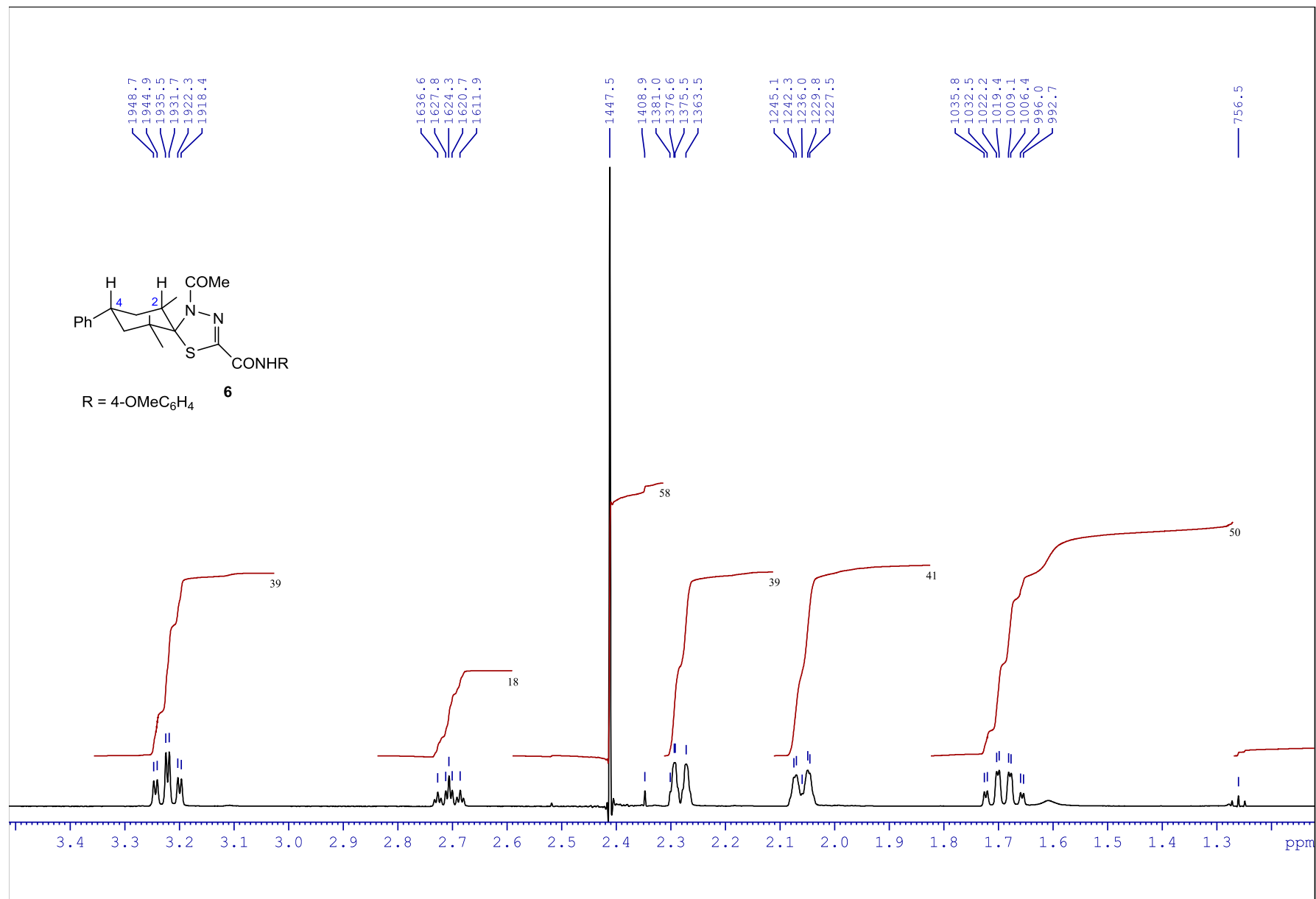
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **5** ( $\text{CDCl}_3$ ).



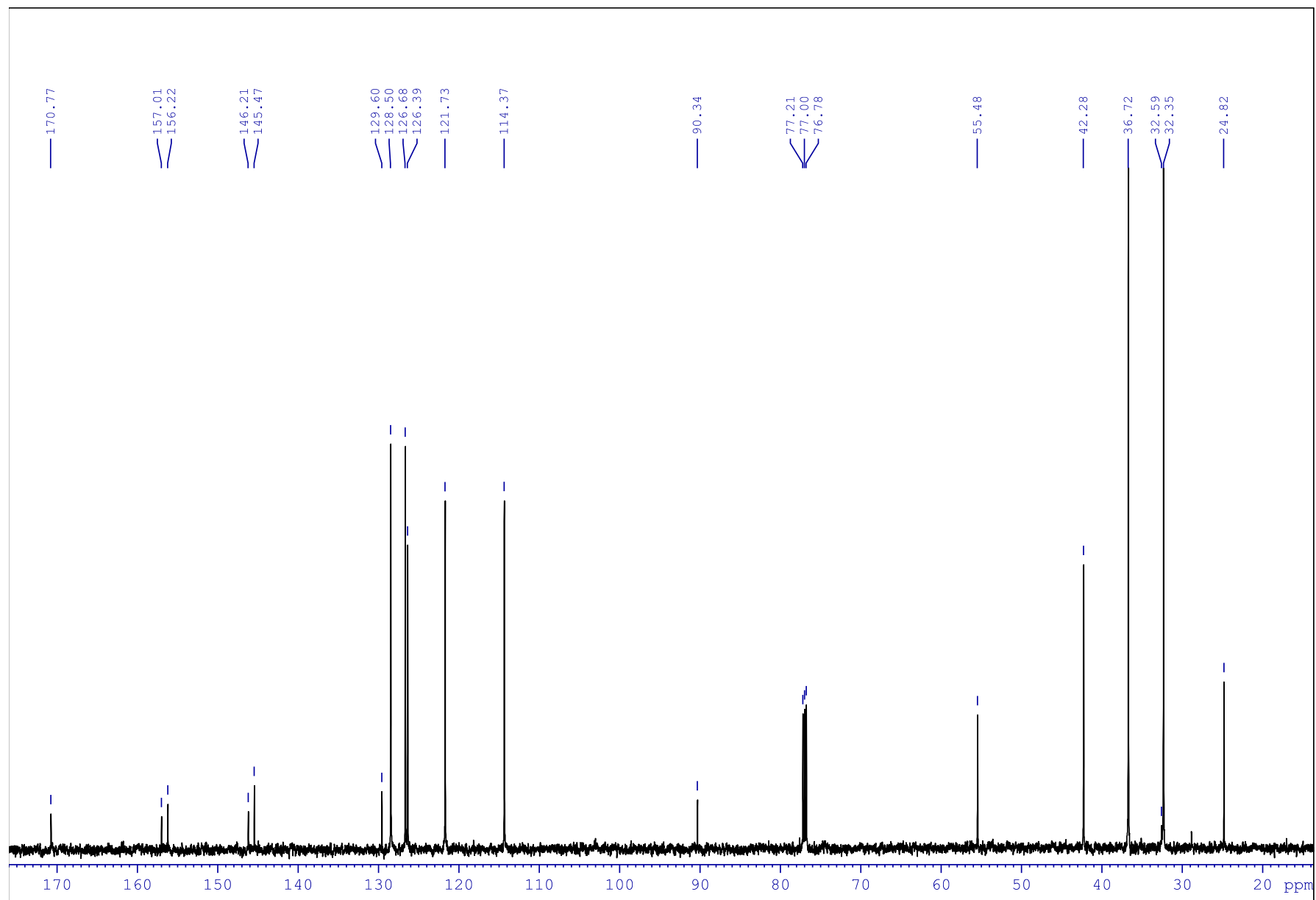
1D  $^1\text{H}$  NOESY (3.25 ppm) NMR spectrum of **5** ( $\text{CDCl}_3$ ).



1D <sup>1</sup>H NOESY (2.04 ppm) NMR spectrum of **5** (CDCl<sub>3</sub>).

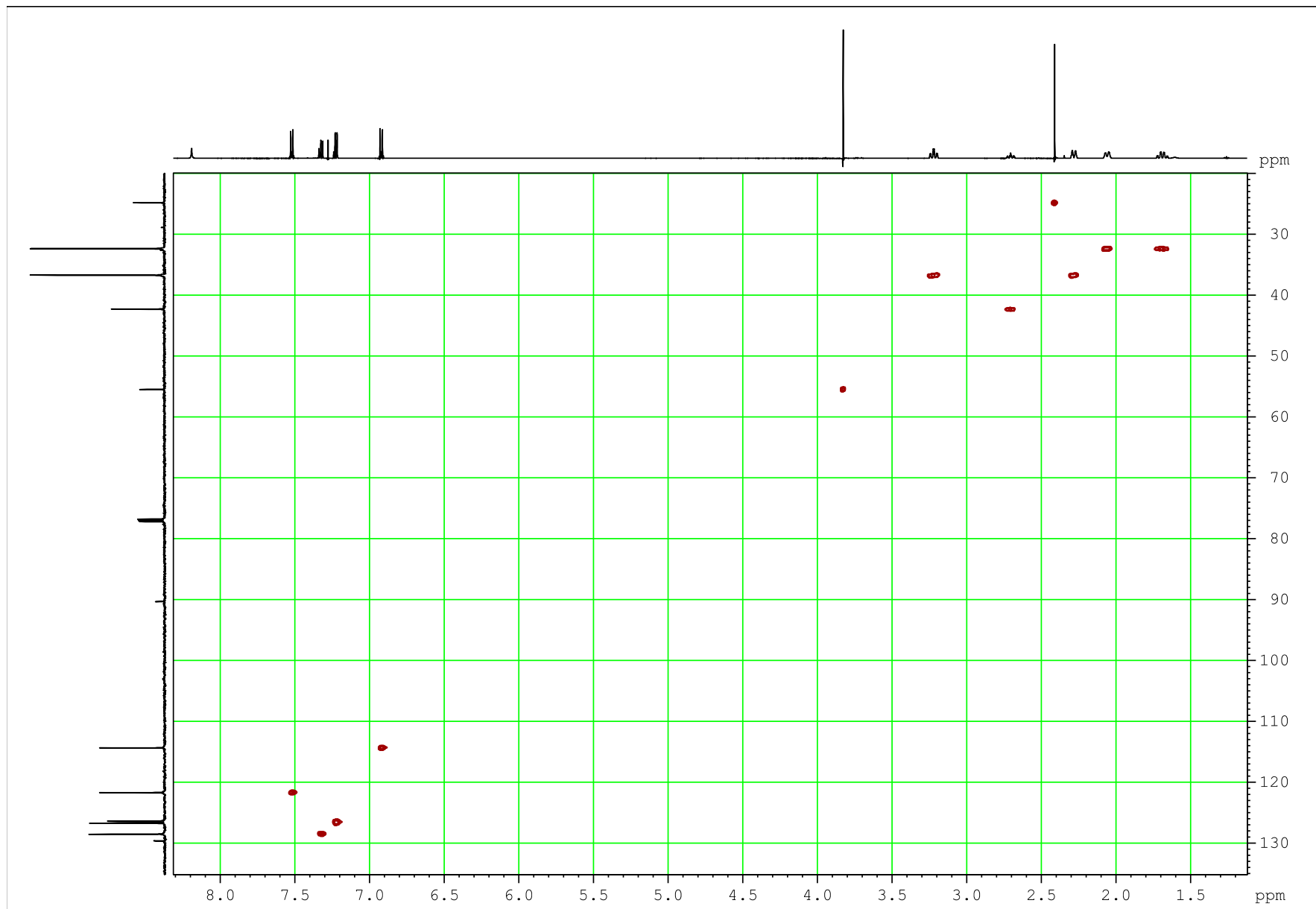


<sup>1</sup>H NMR spectrum of **6** (CDCl<sub>3</sub>).

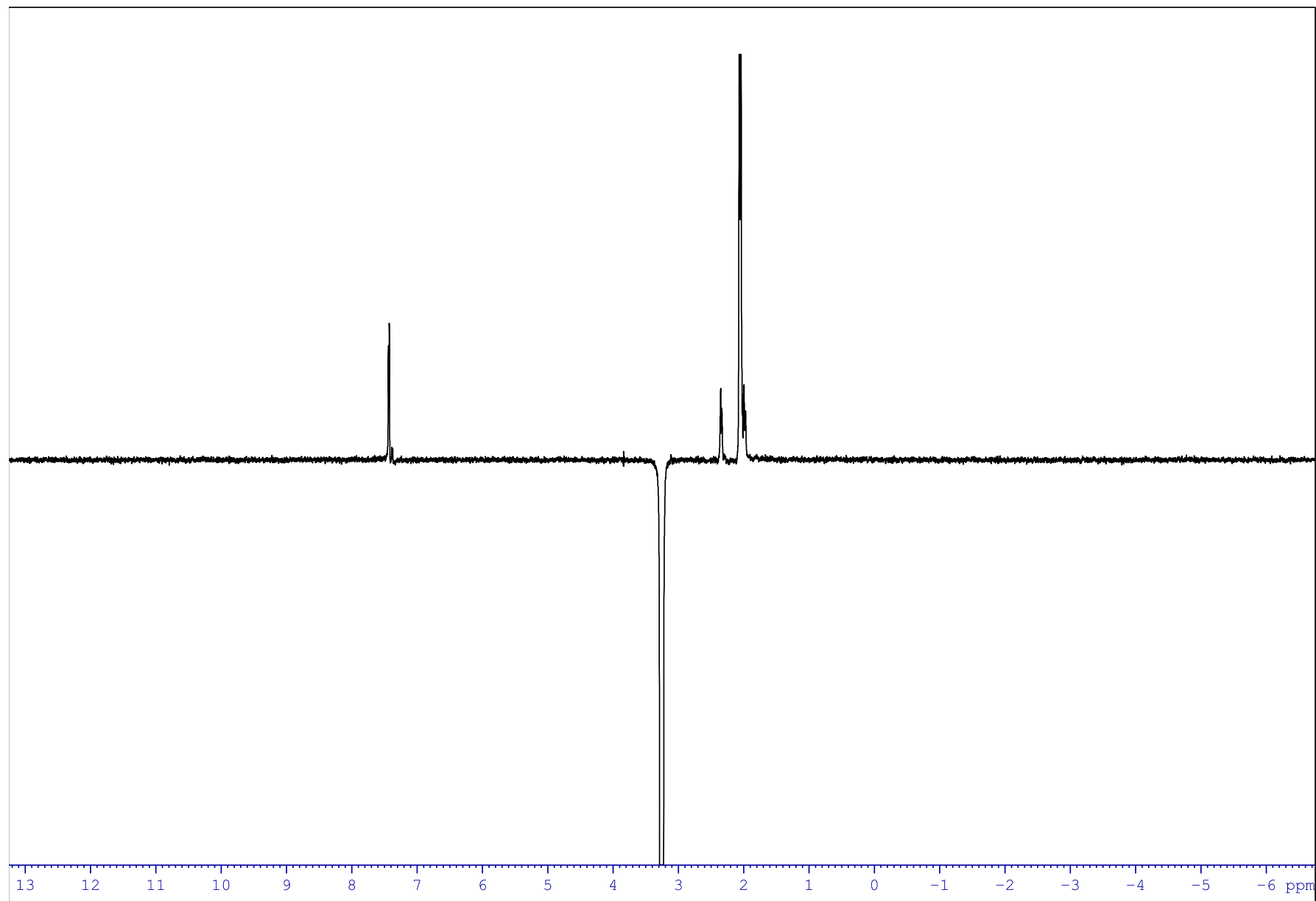


$^{13}\text{C}$  NMR spectrum of **6** ( $\text{CDCl}_3$ ).

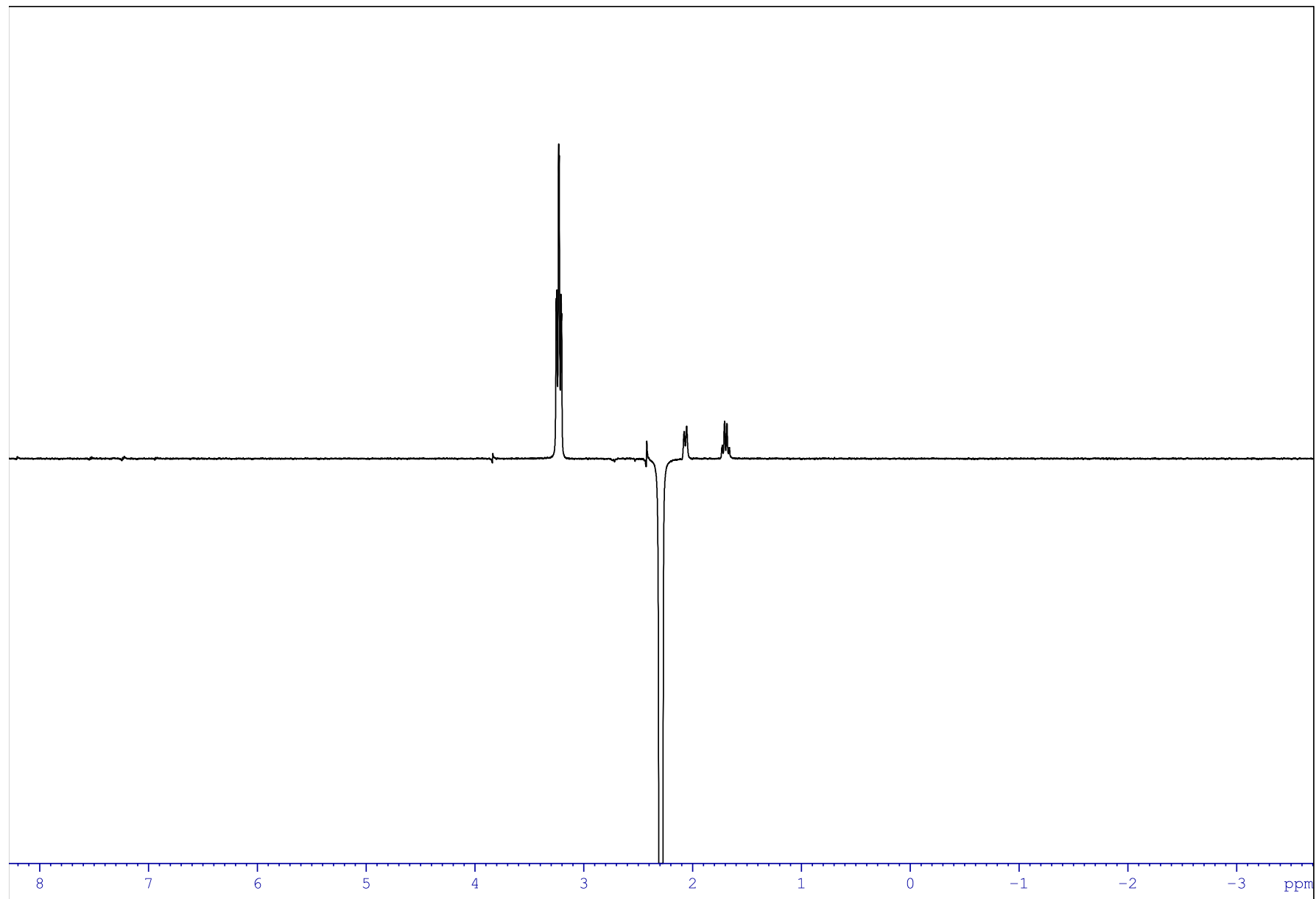




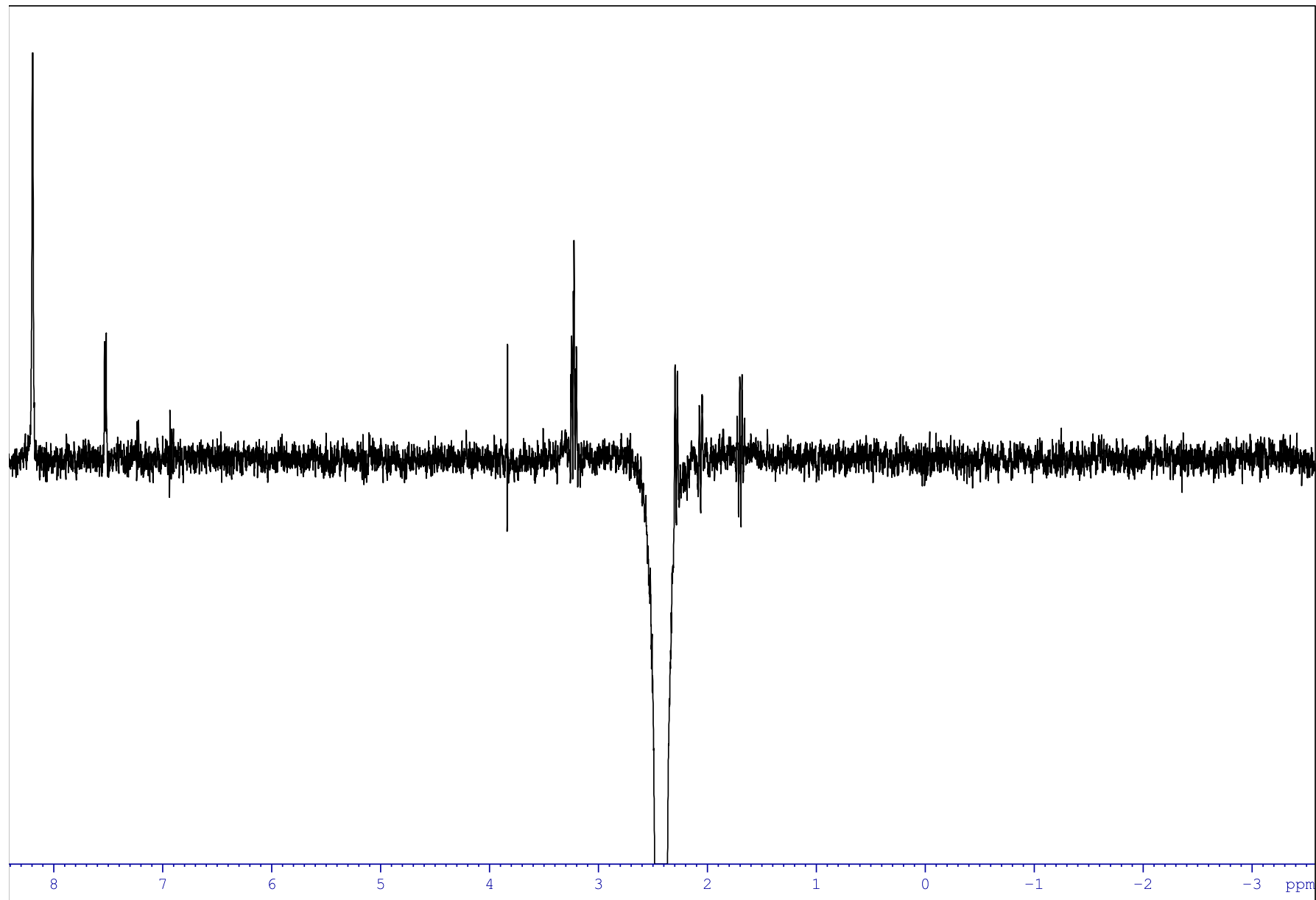
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **6** ( $\text{CDCl}_3$ ).



1D <sup>1</sup>H NOESY (3.25 ppm) NMR spectrum of **6** (CDCl<sub>3</sub>).

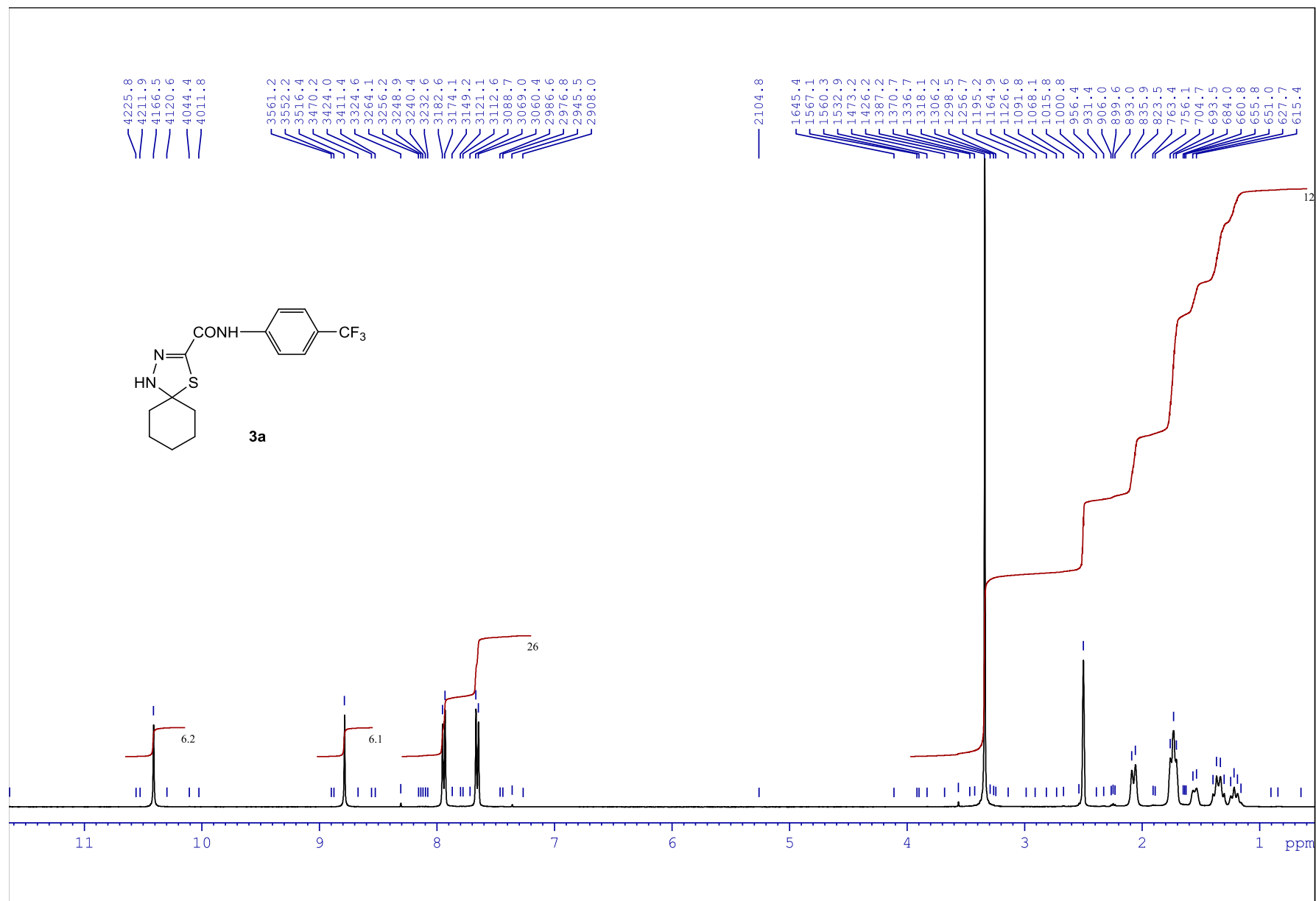


1D <sup>1</sup>H NOESY (2.28 ppm) NMR spectrum of **6** (CDCl<sub>3</sub>).

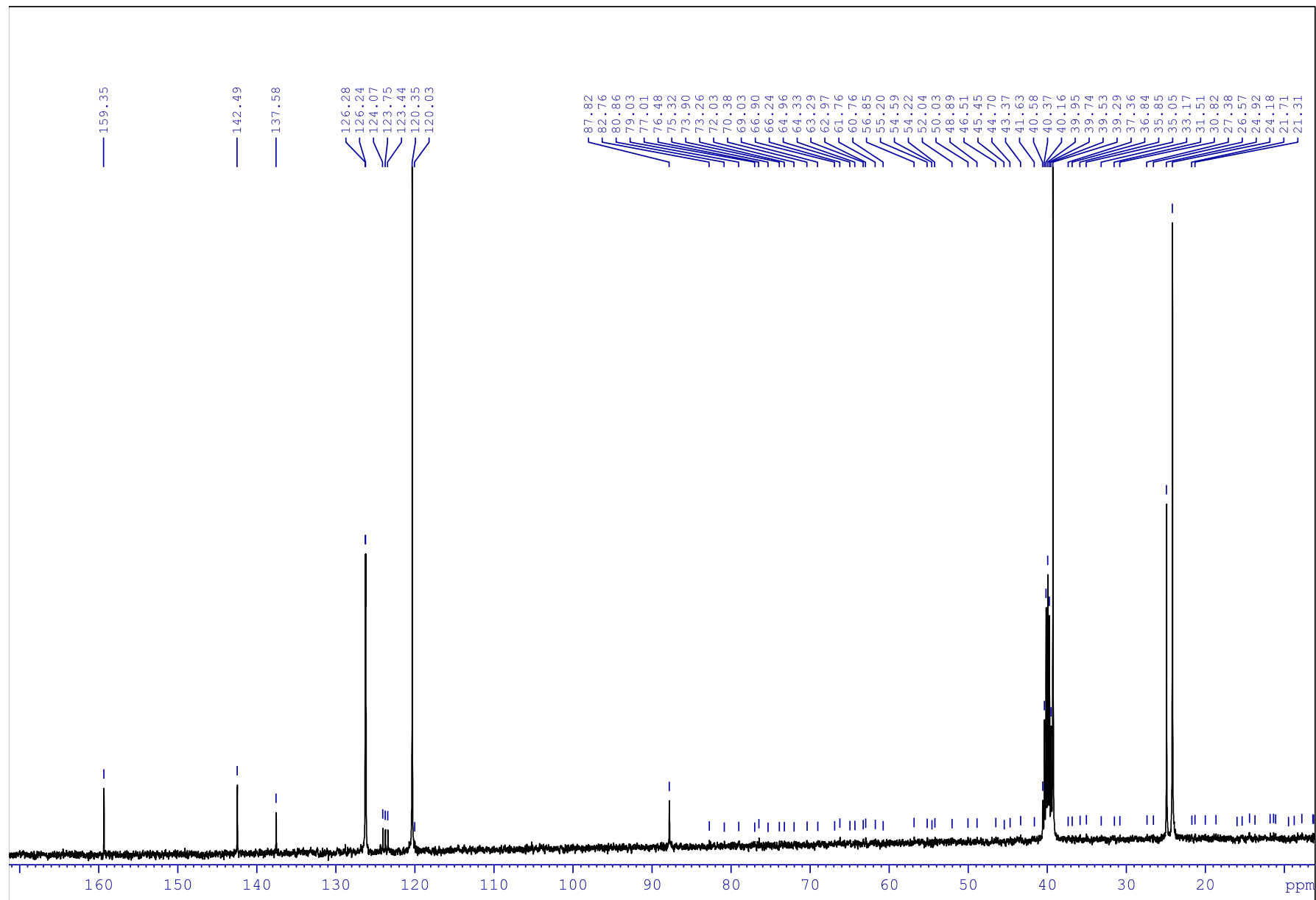


1D  $^1\text{H}$  NOESY (2.41 ppm) NMR spectrum of **6** ( $\text{CDCl}_3$ ).

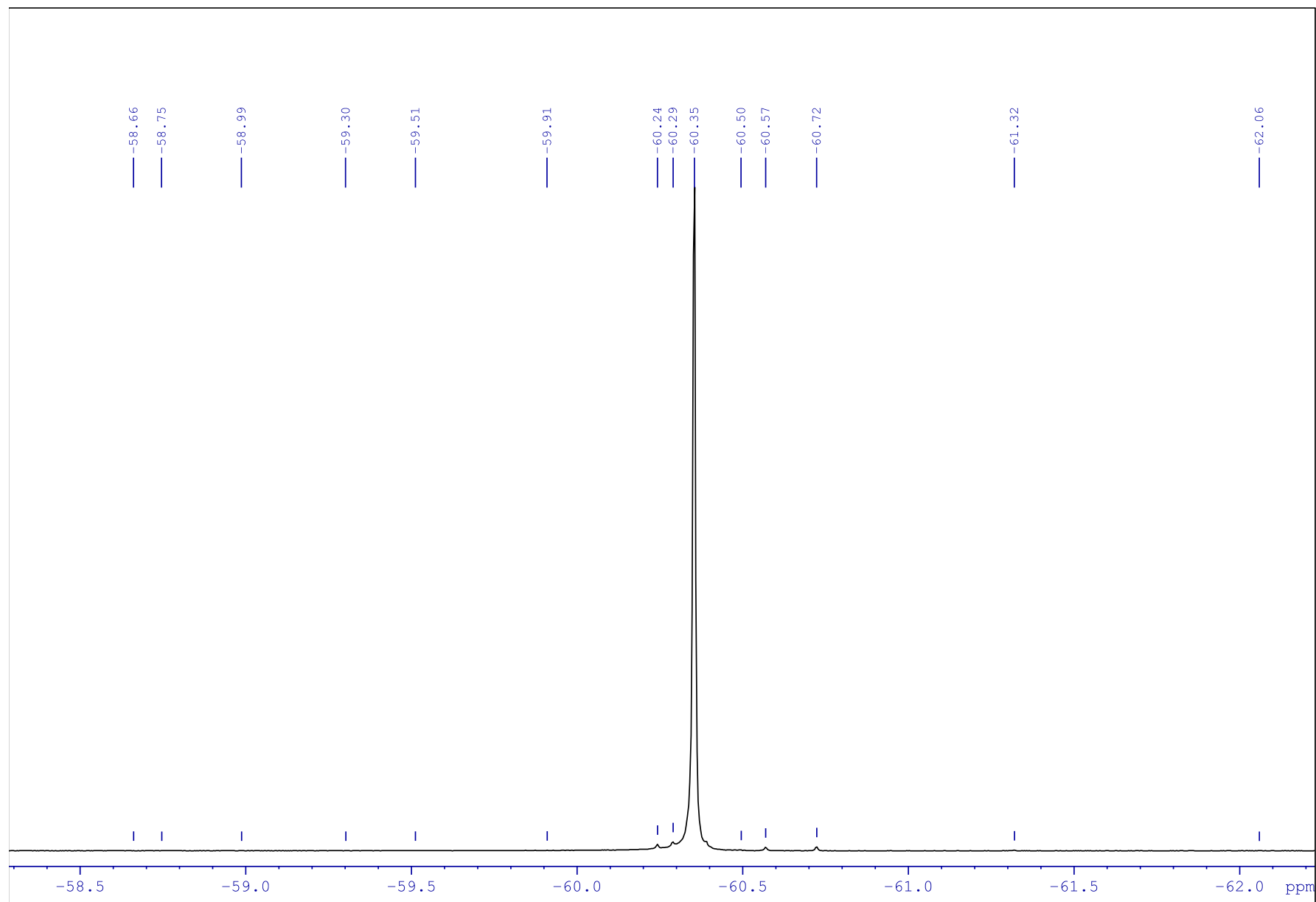
## 2. NMR spectra (Bruker AV-400)



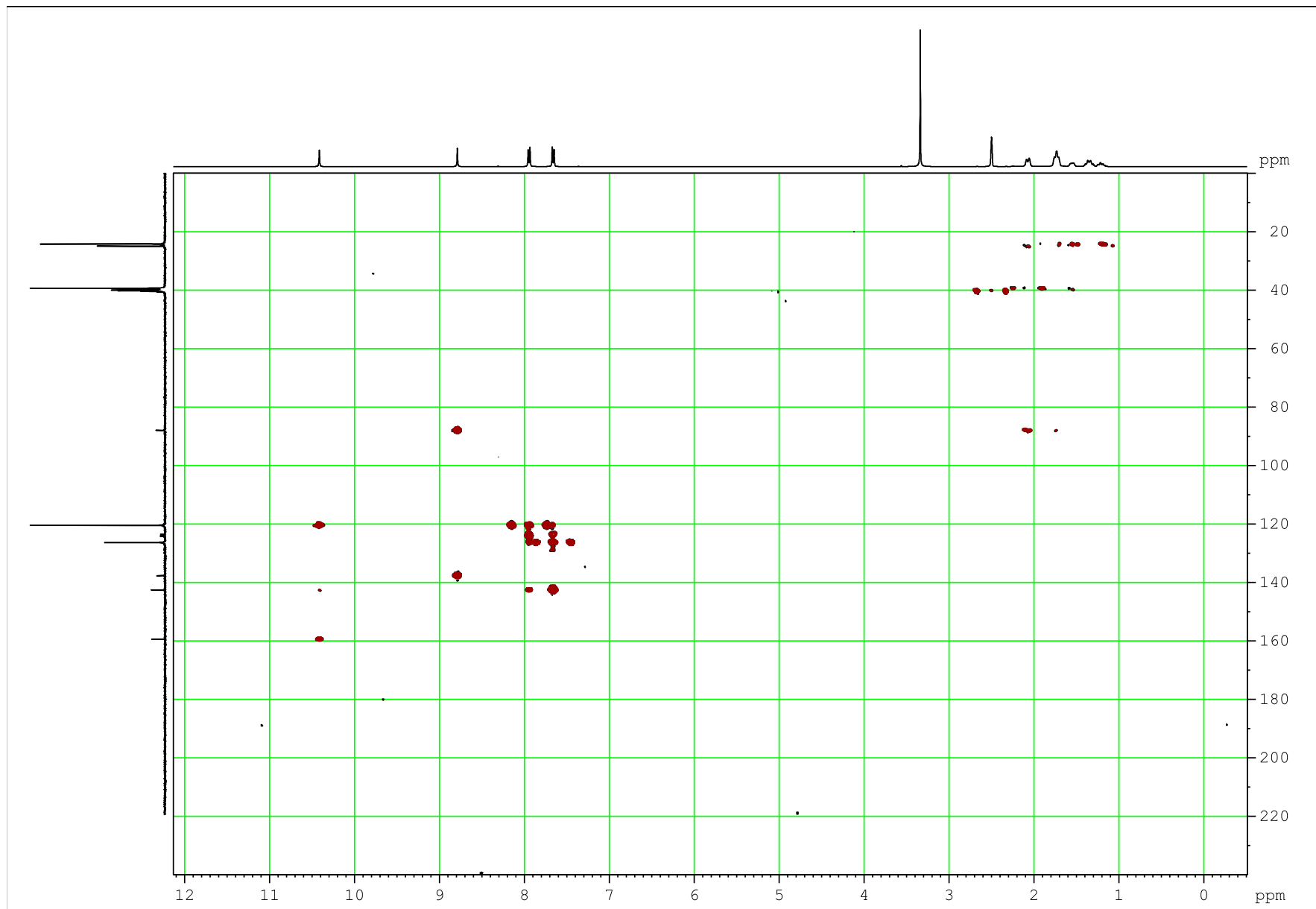
<sup>1</sup>H NMR spectrum of **3a** (DMSO-*d*<sub>6</sub>).



$^{13}\text{C}$  NMR spectrum of **3a** (DMSO- $d_6$ ).

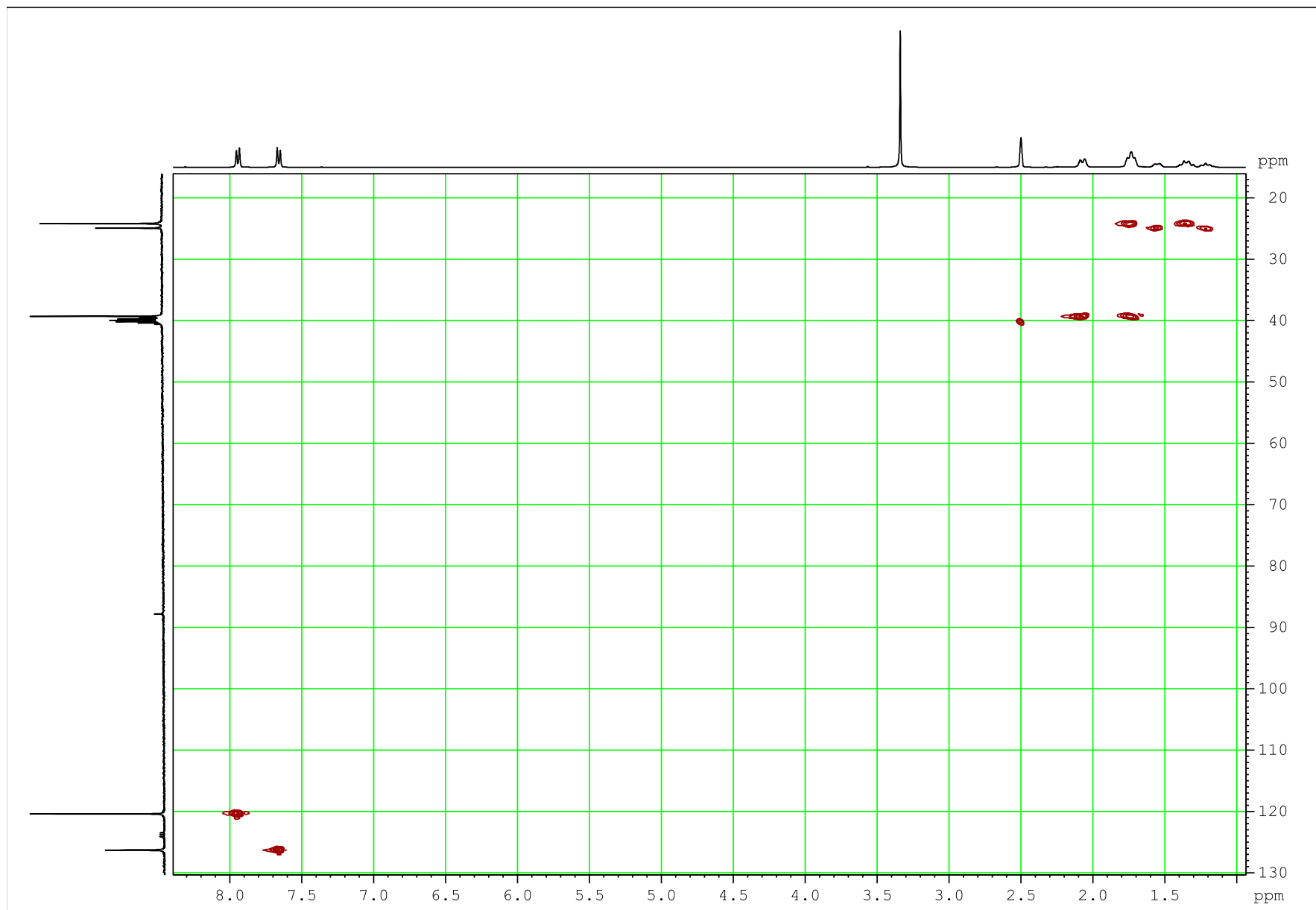


$^{19}\text{F}$  NMR spectrum of **3a** ( $\text{DMSO-}d_6$ ).

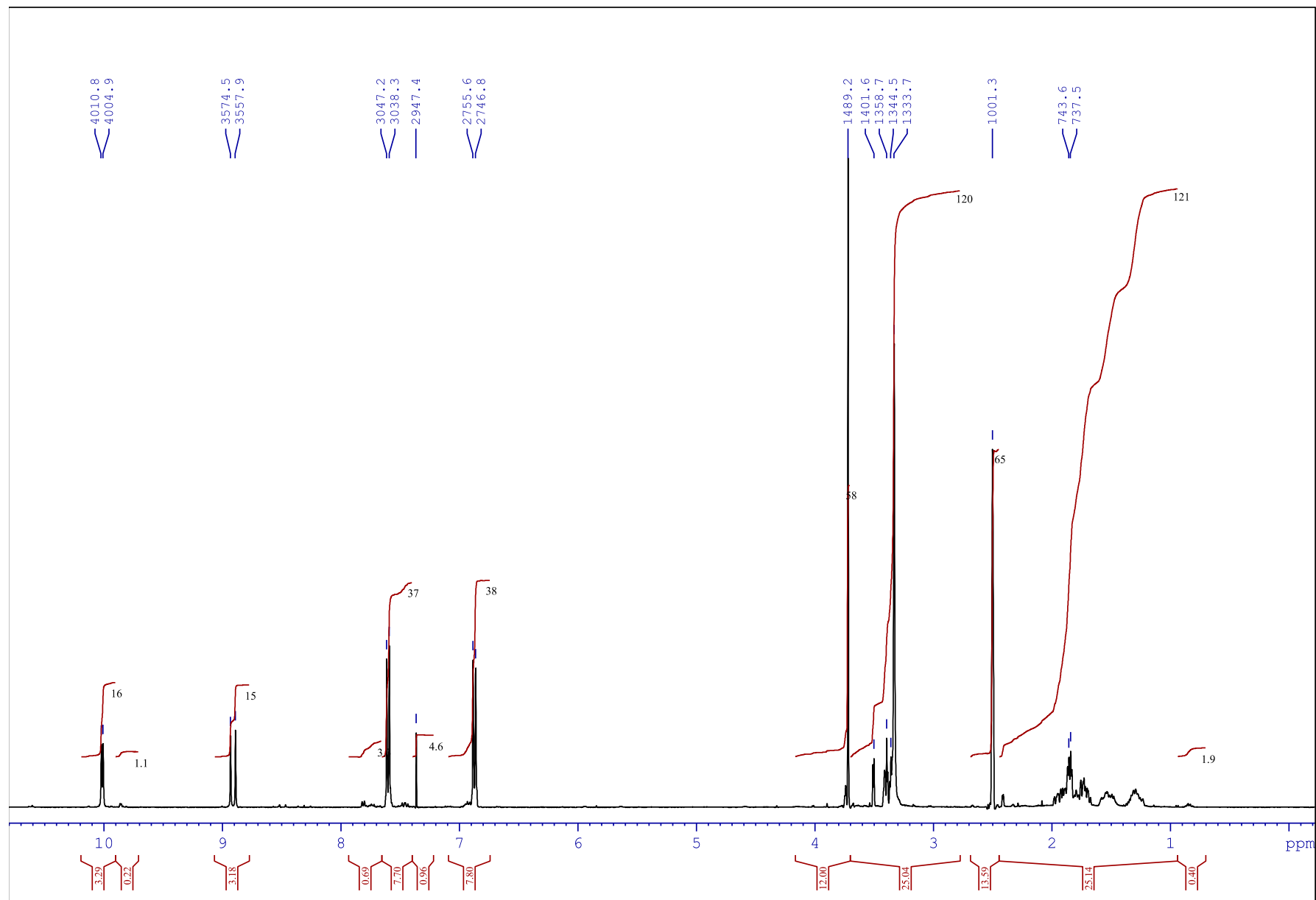


2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3a** ( $\text{DMSO-}d_6$ ).

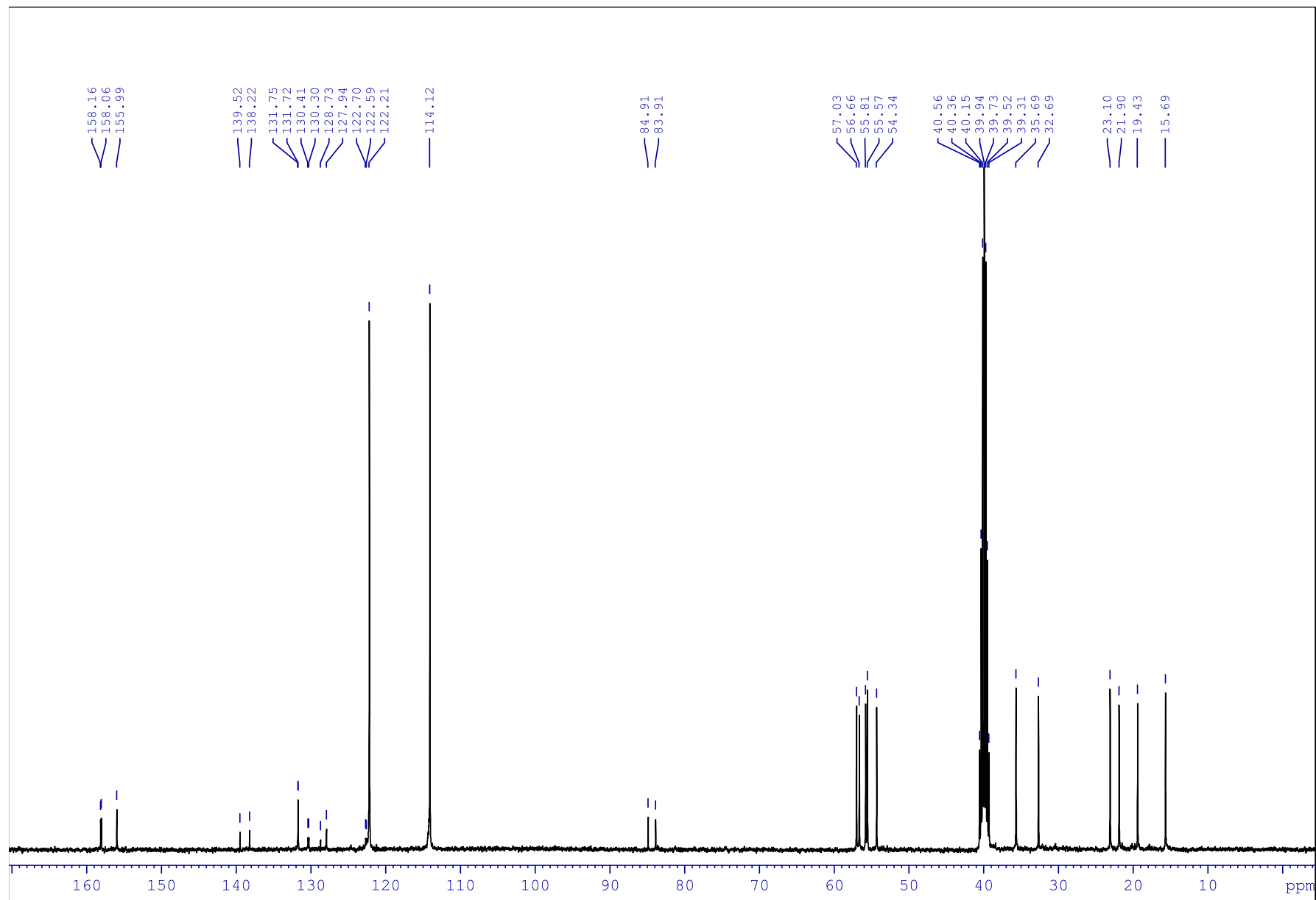




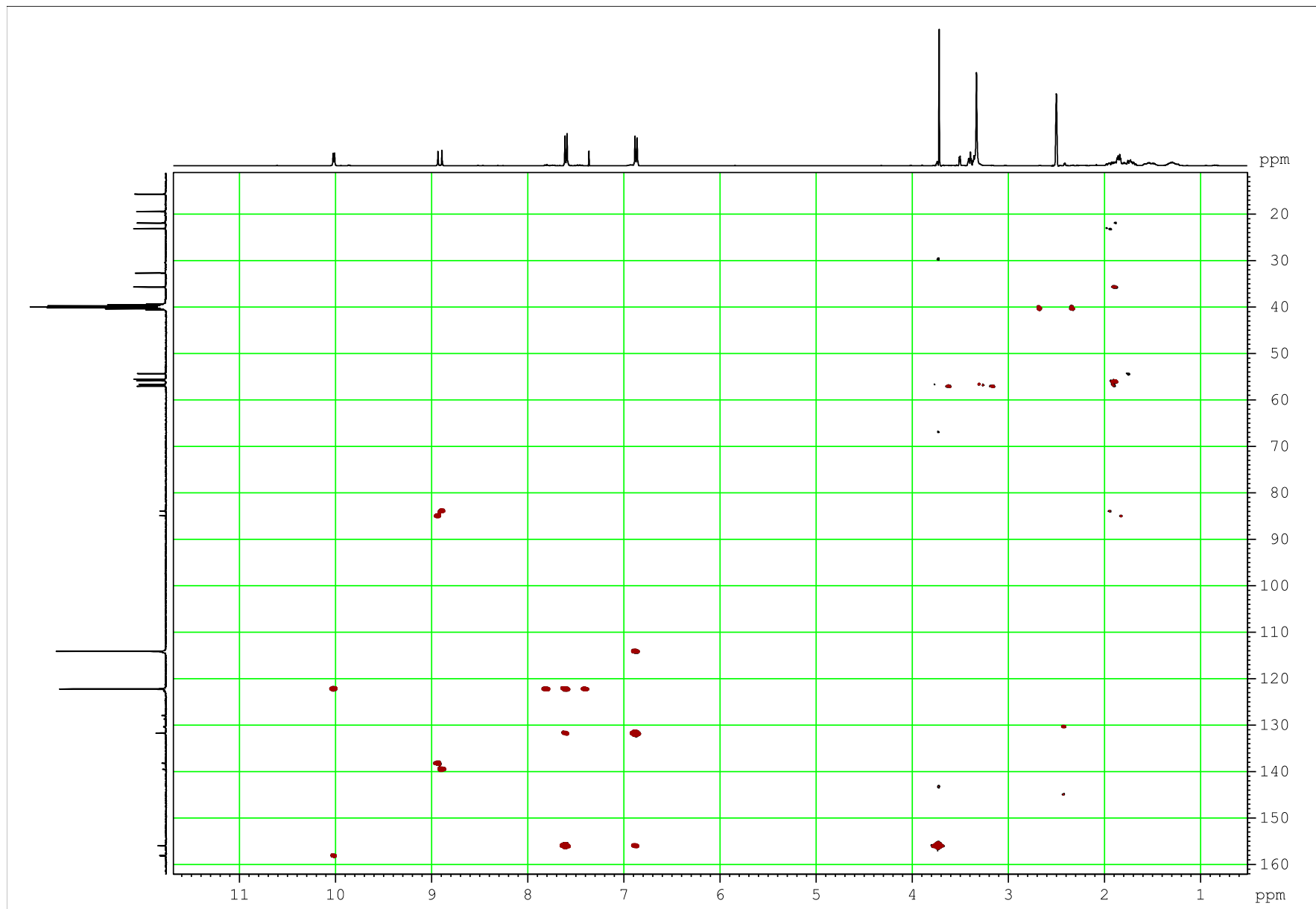
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3a** (DMSO- $d_6$ ).



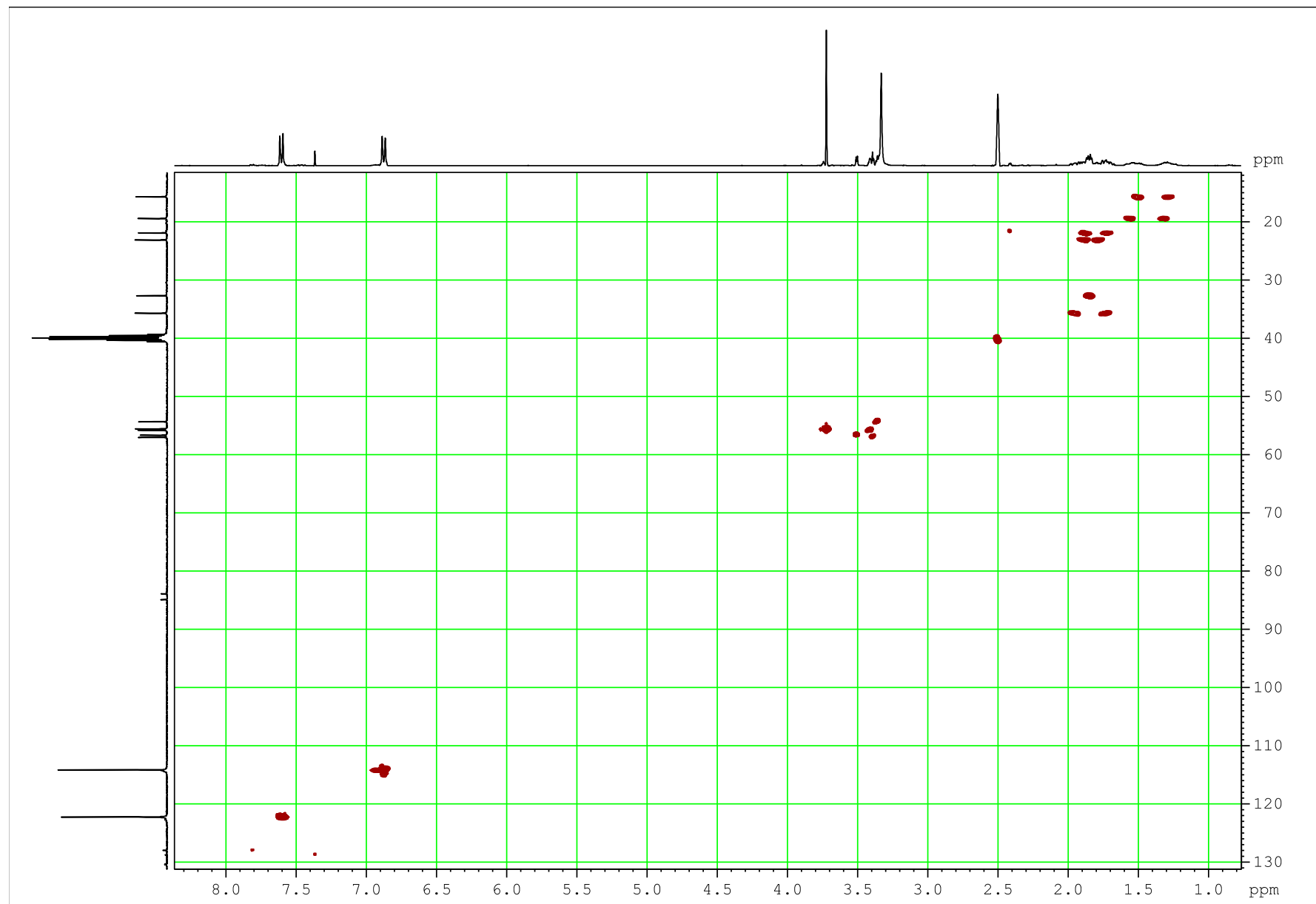
$^1\text{H}$  NMR spectrum of **3e** ( $\text{DMSO-}d_6$ ).



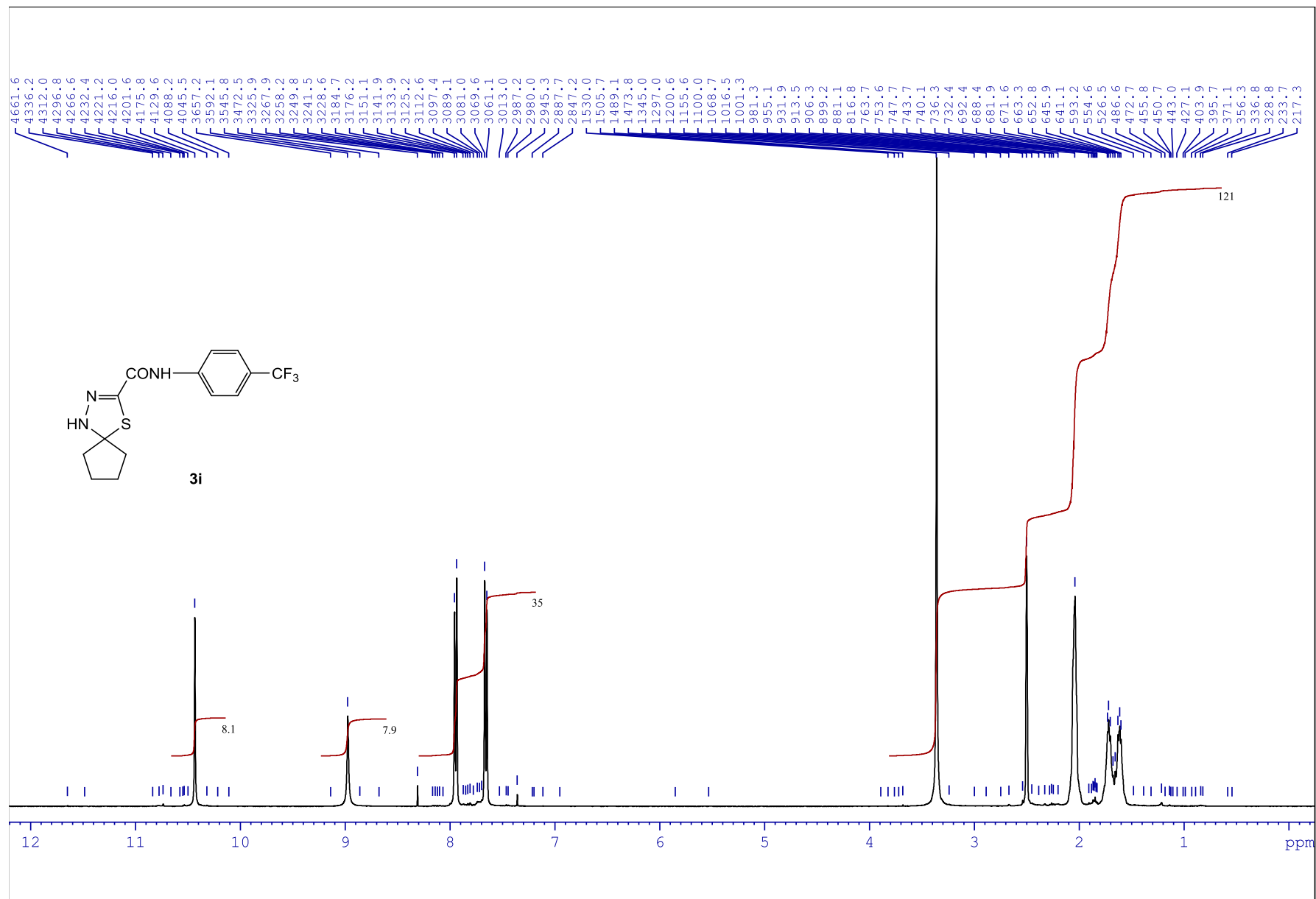
<sup>13</sup>C NMR spectrum of **3e** (DMSO-*d*<sub>6</sub>).



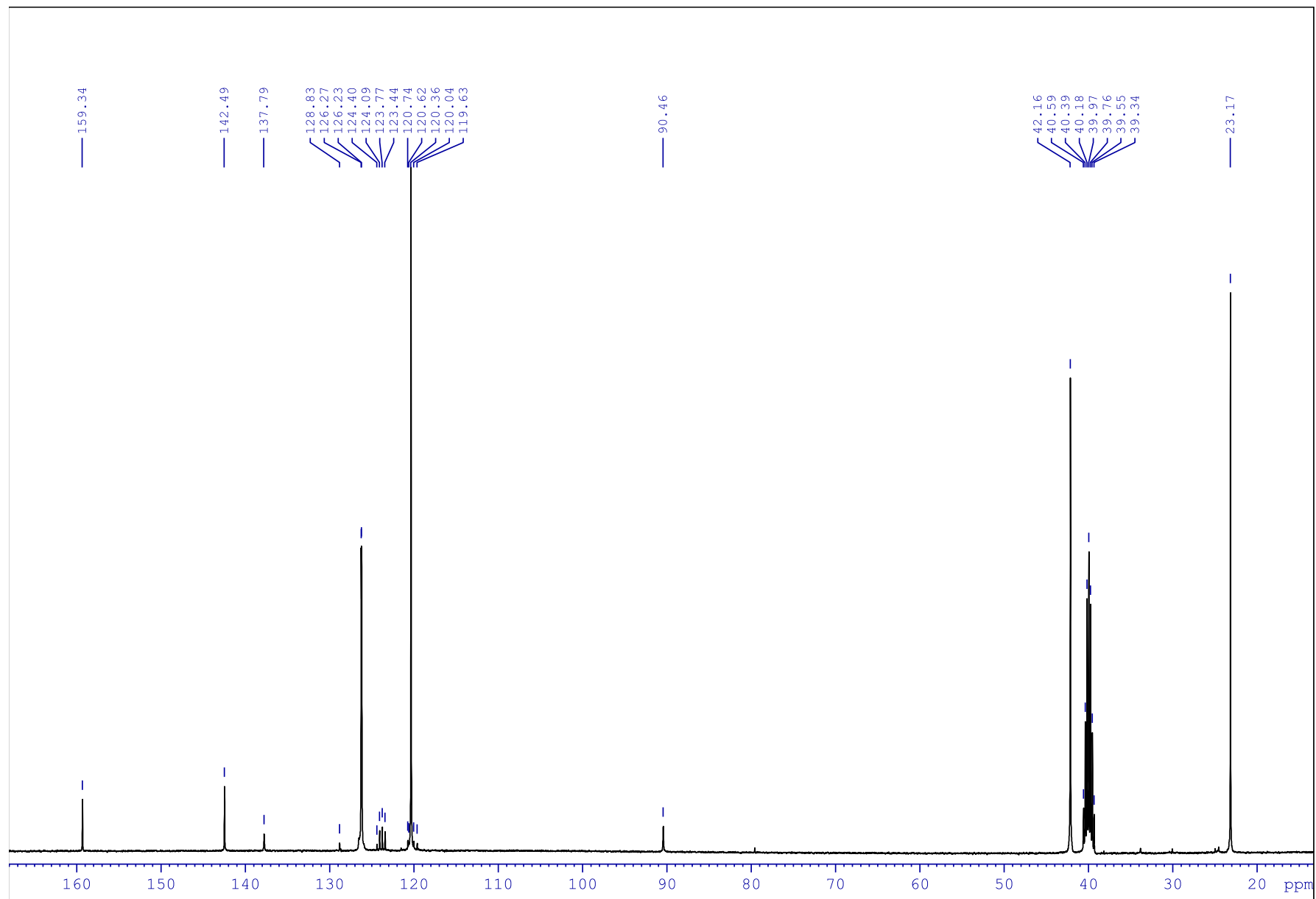
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3e** ( $\text{DMSO-}d_6$ ).



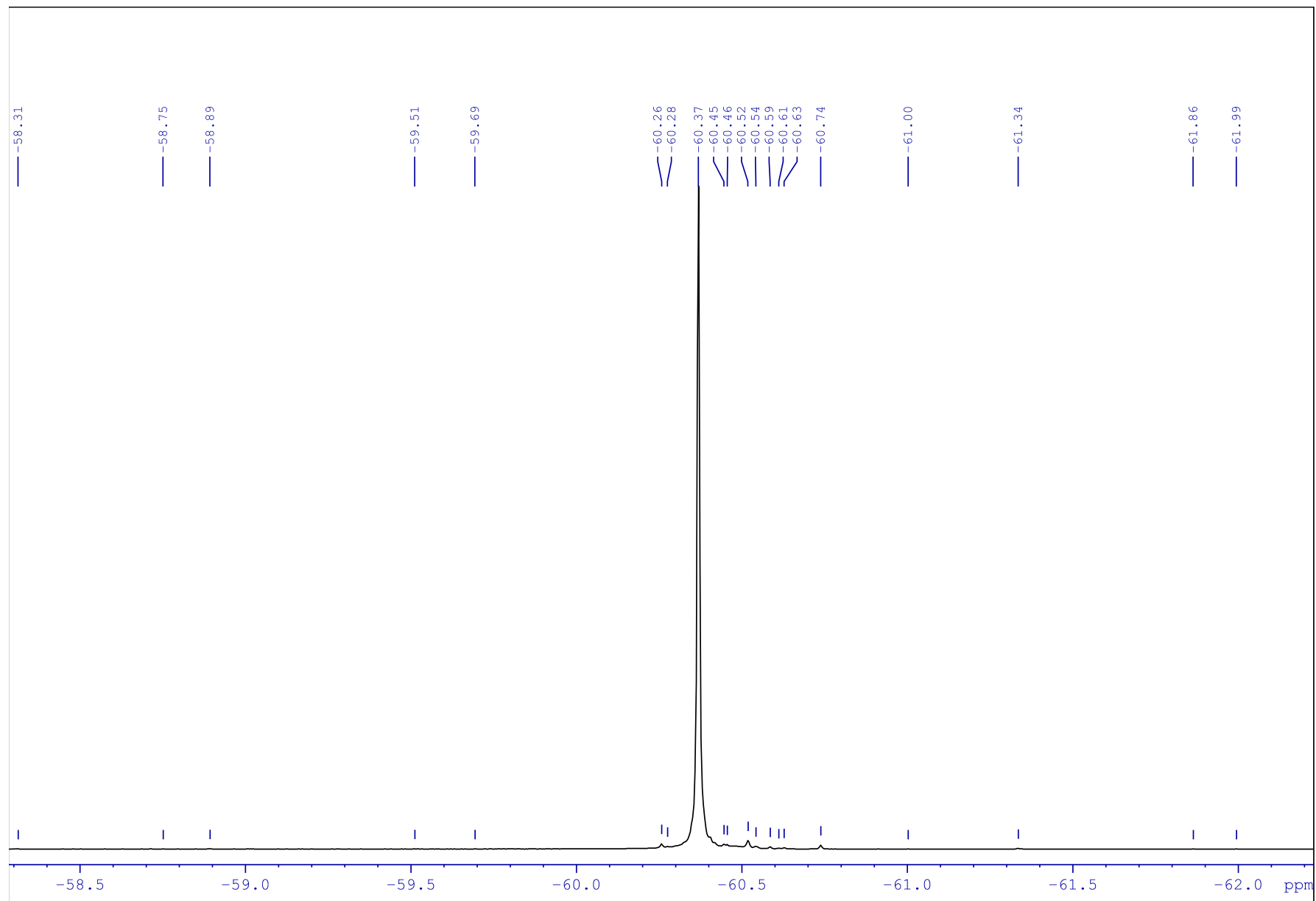
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3e** ( $\text{DMSO-}d_6$ ).



<sup>1</sup>H NMR spectrum of **3i** (DMSO-*d*<sub>6</sub>).



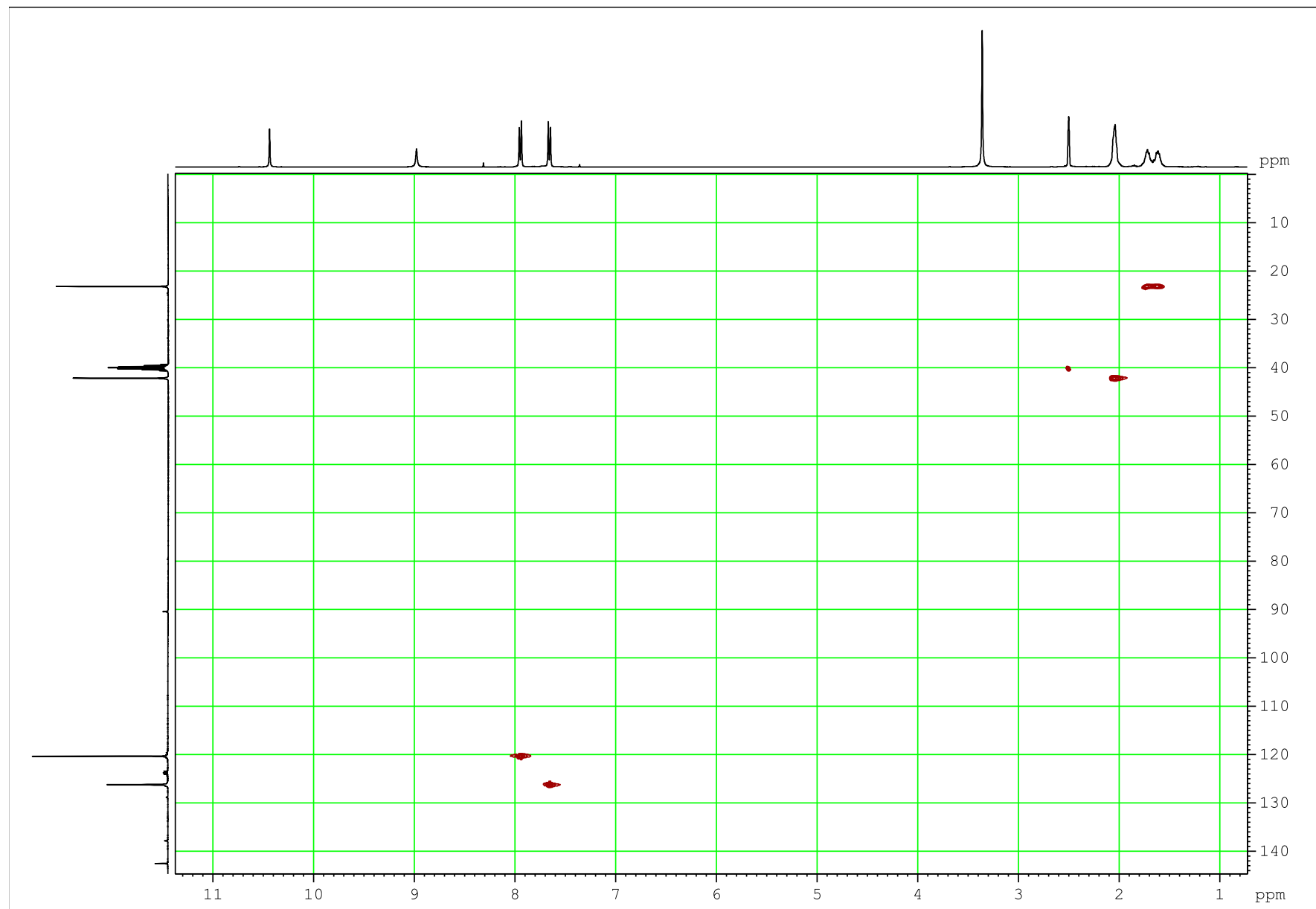
$^{13}\text{C}$  NMR spectrum of **3i** ( $\text{DMSO-}d_6$ ).



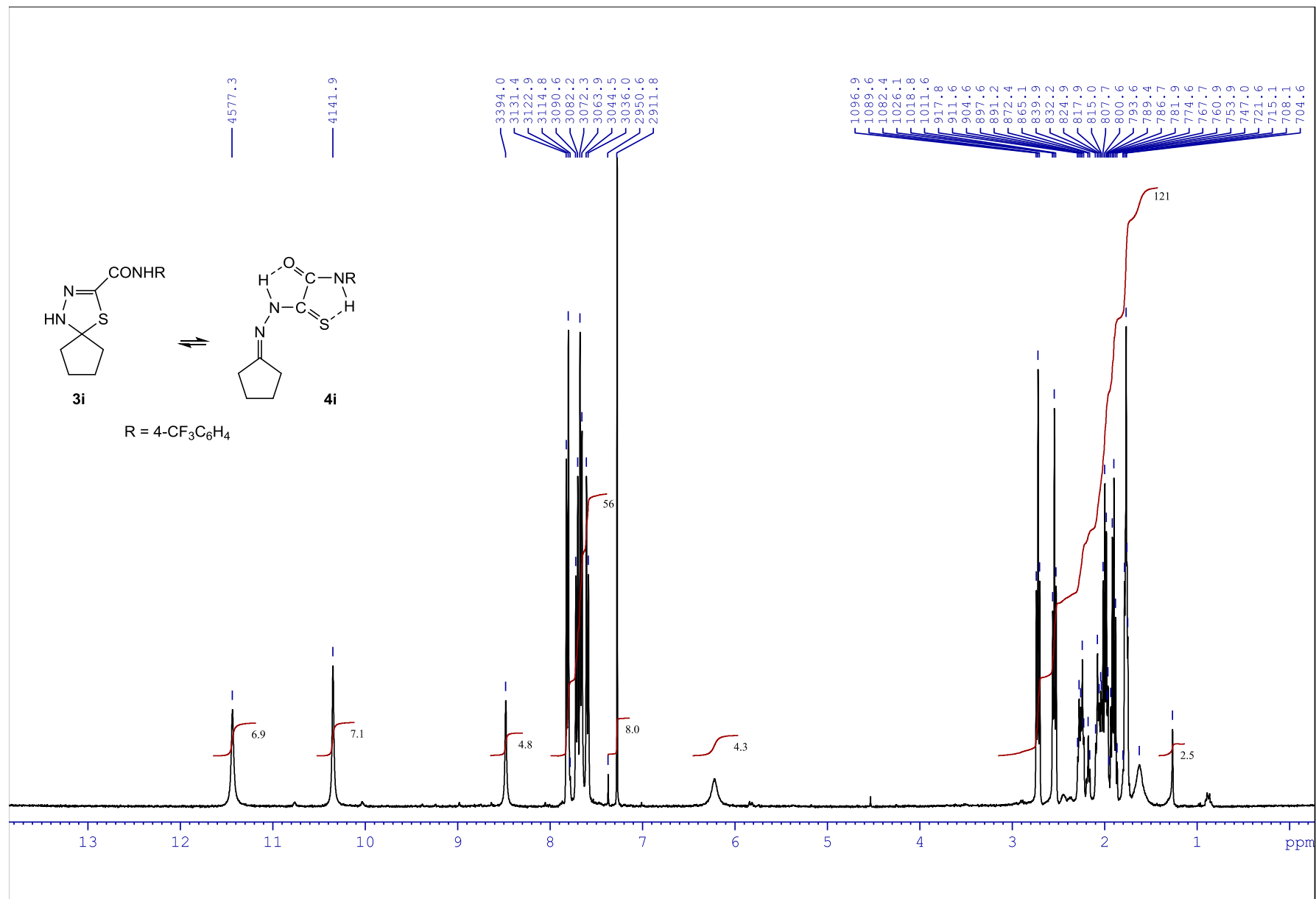
$^{19}\text{F}$  NMR spectrum of **3i** ( $\text{DMSO-}d_6$ ).



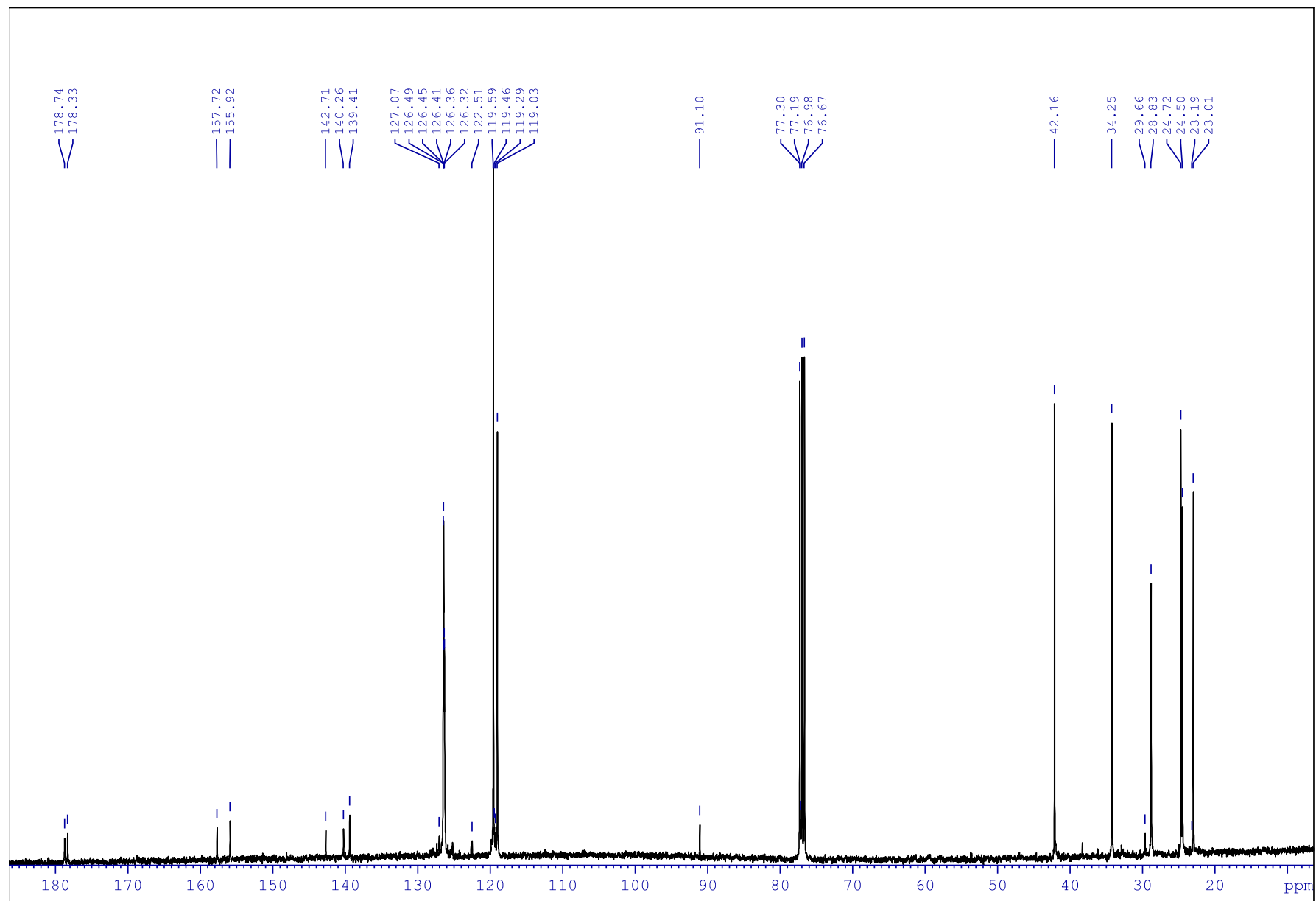




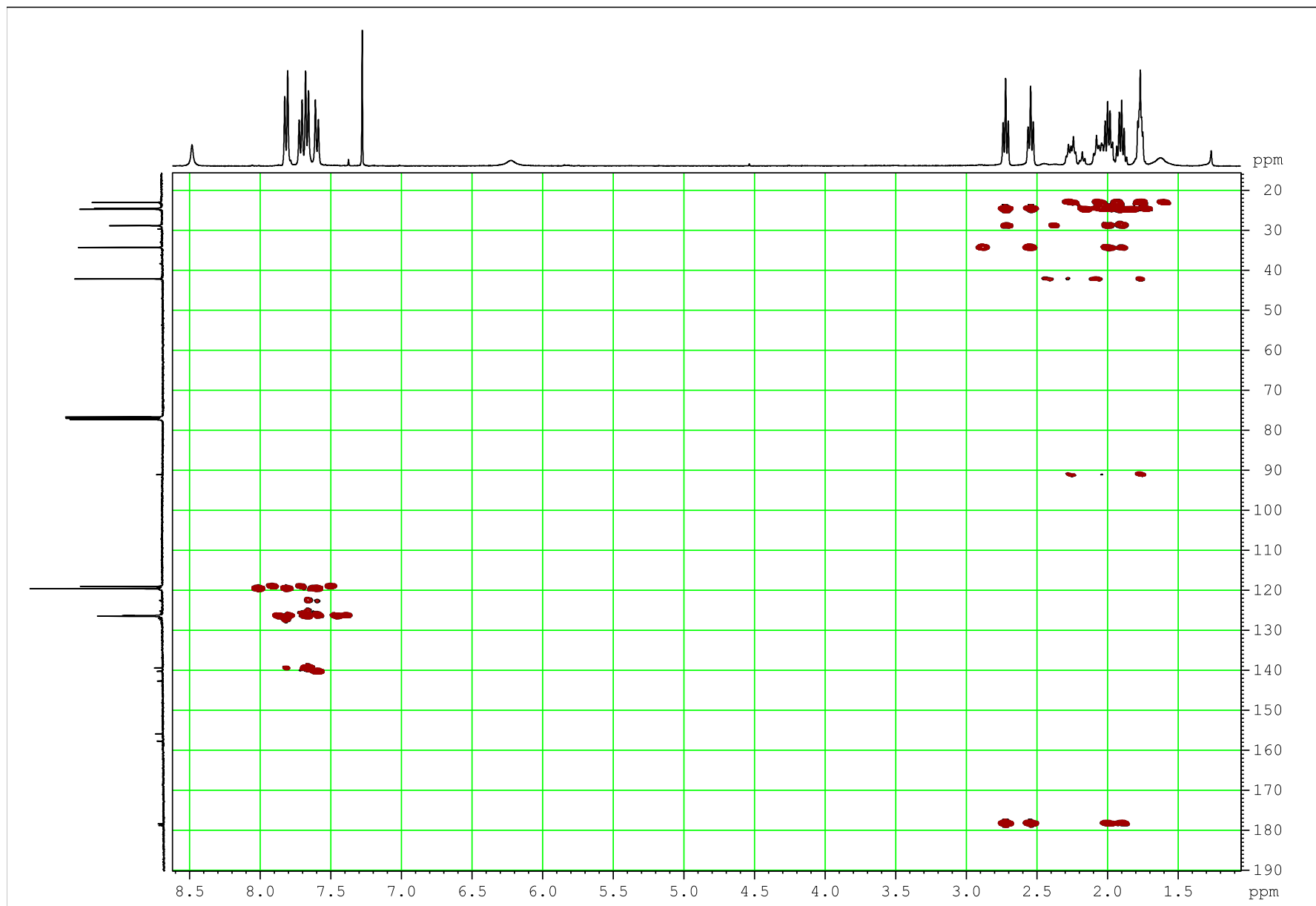
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3i** ( $\text{DMSO}-d_6$ ).



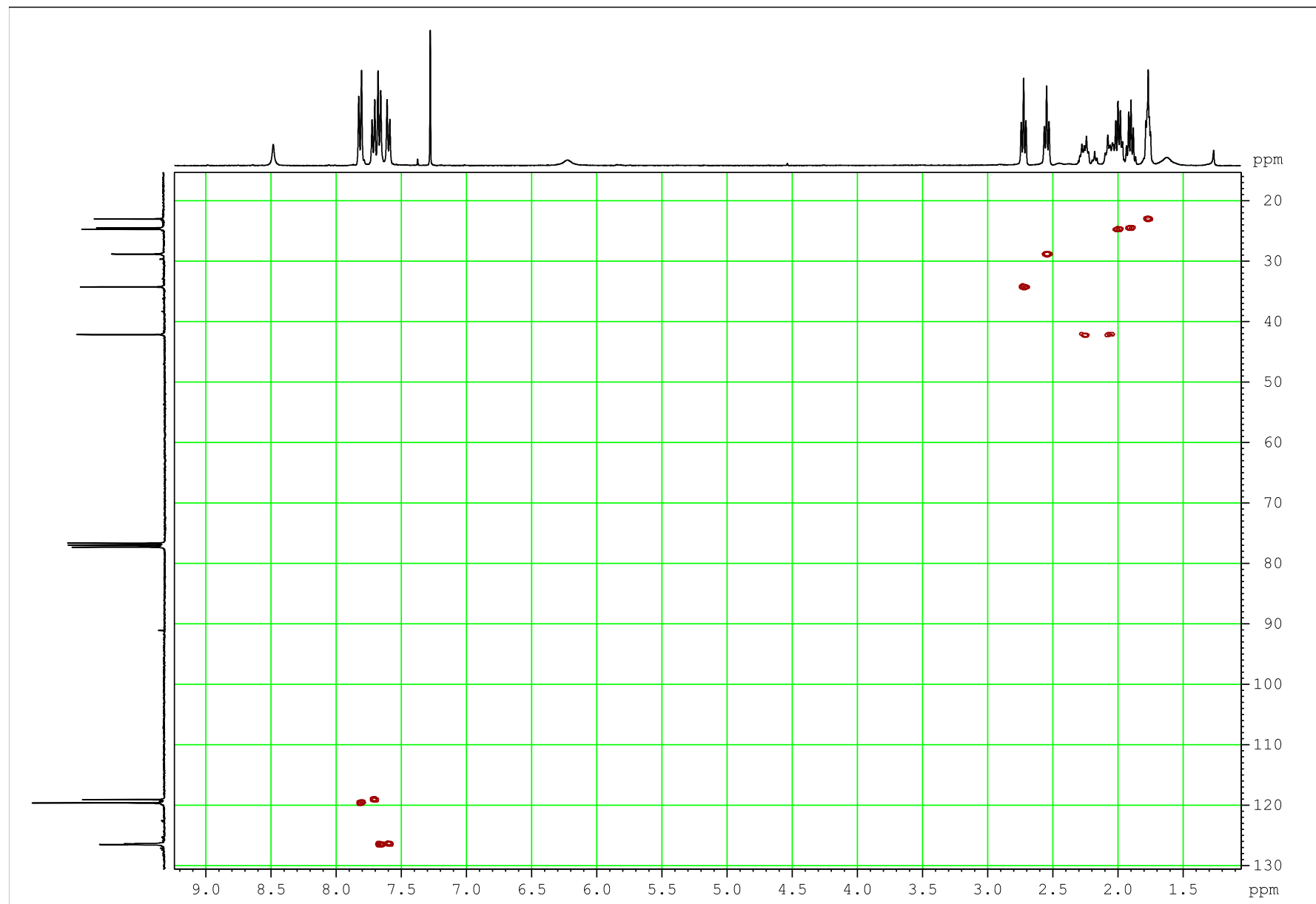
$^1\text{H}$  NMR spectrum of **3i/4i** (CDCl<sub>3</sub>).



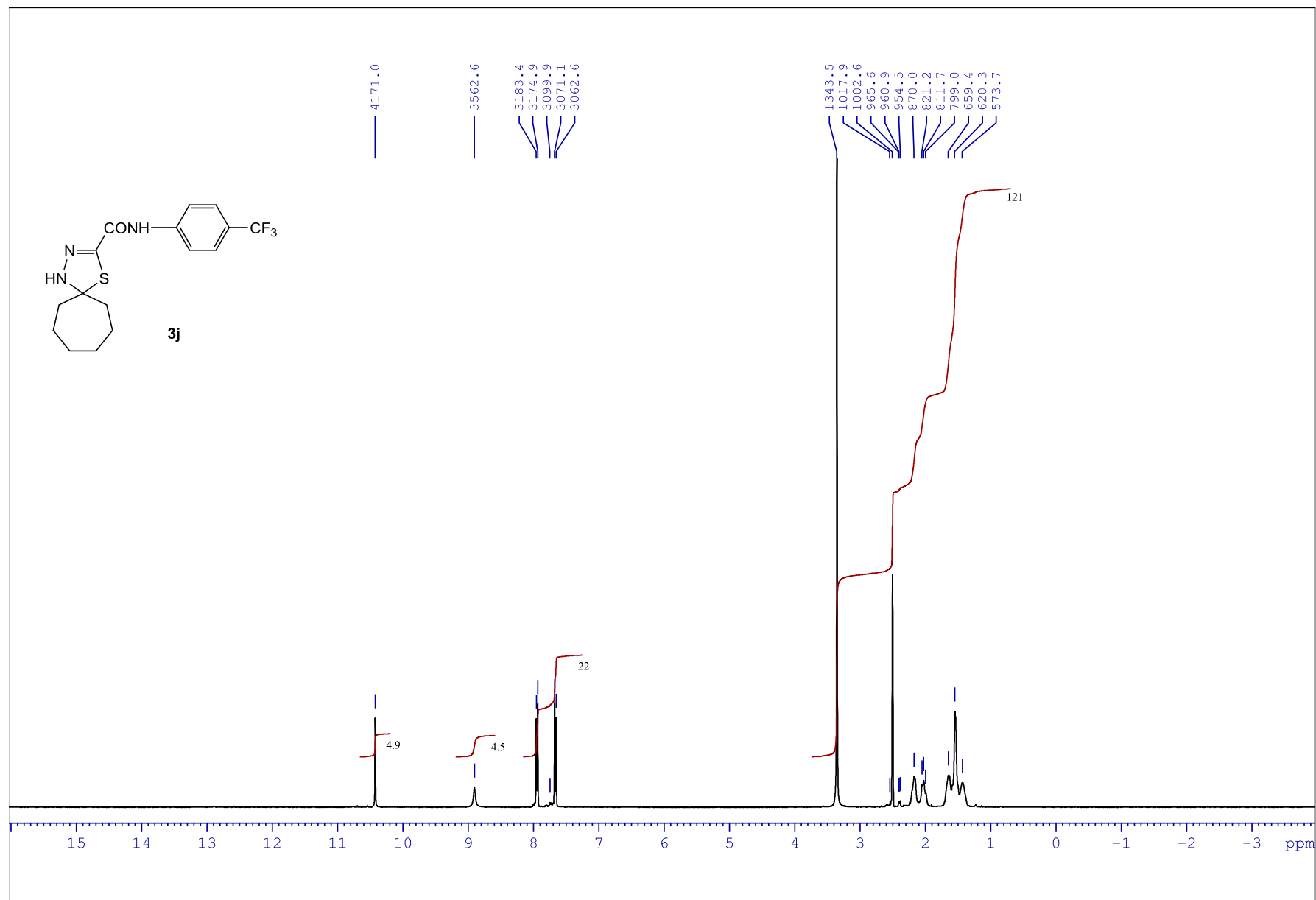
<sup>13</sup>C NMR spectrum of **3i/4i** (CDCl<sub>3</sub>).



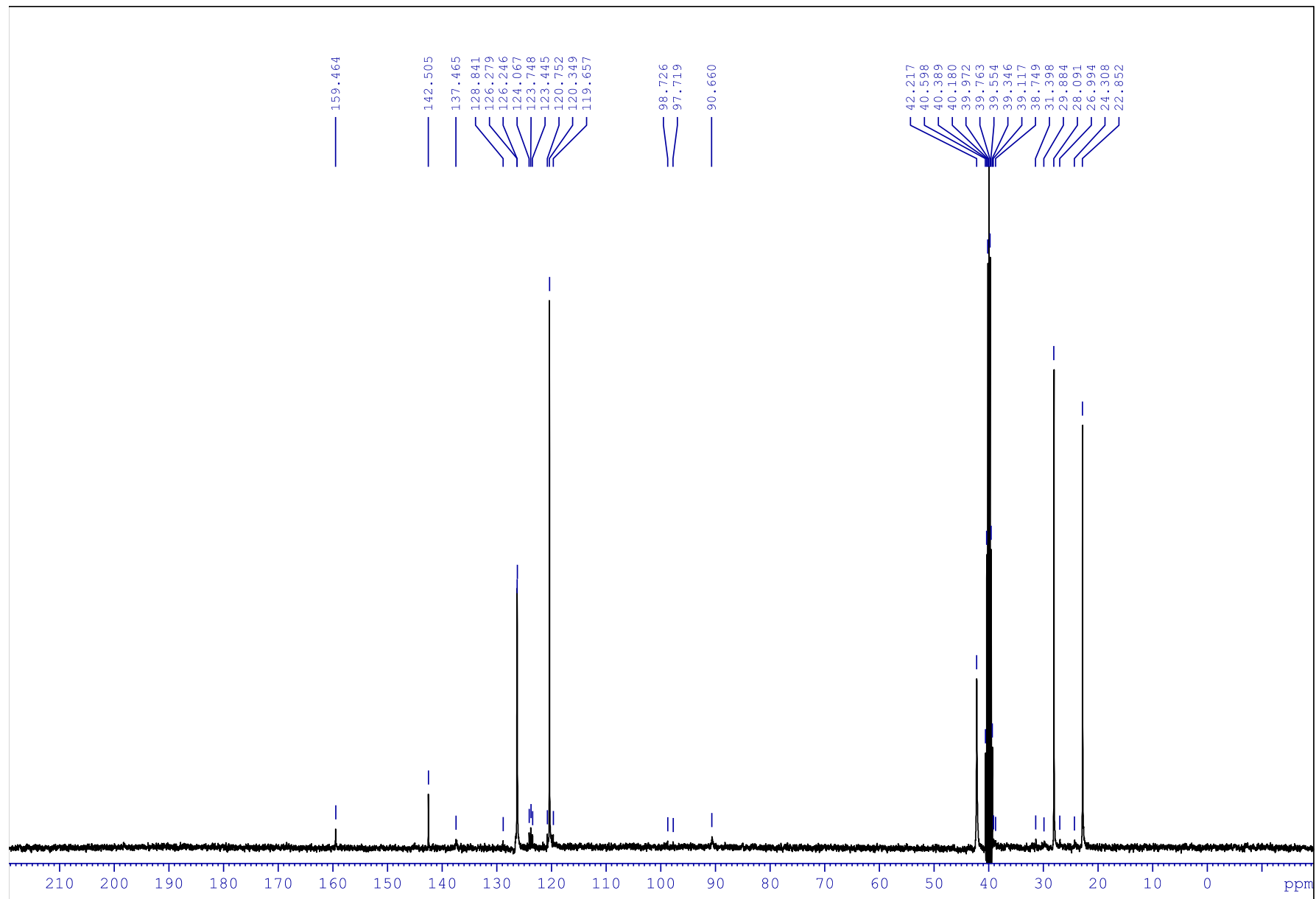
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3i/4i** ( $\text{CDCl}_3$ ).



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3i/4i** ( $\text{CDCl}_3$ ).

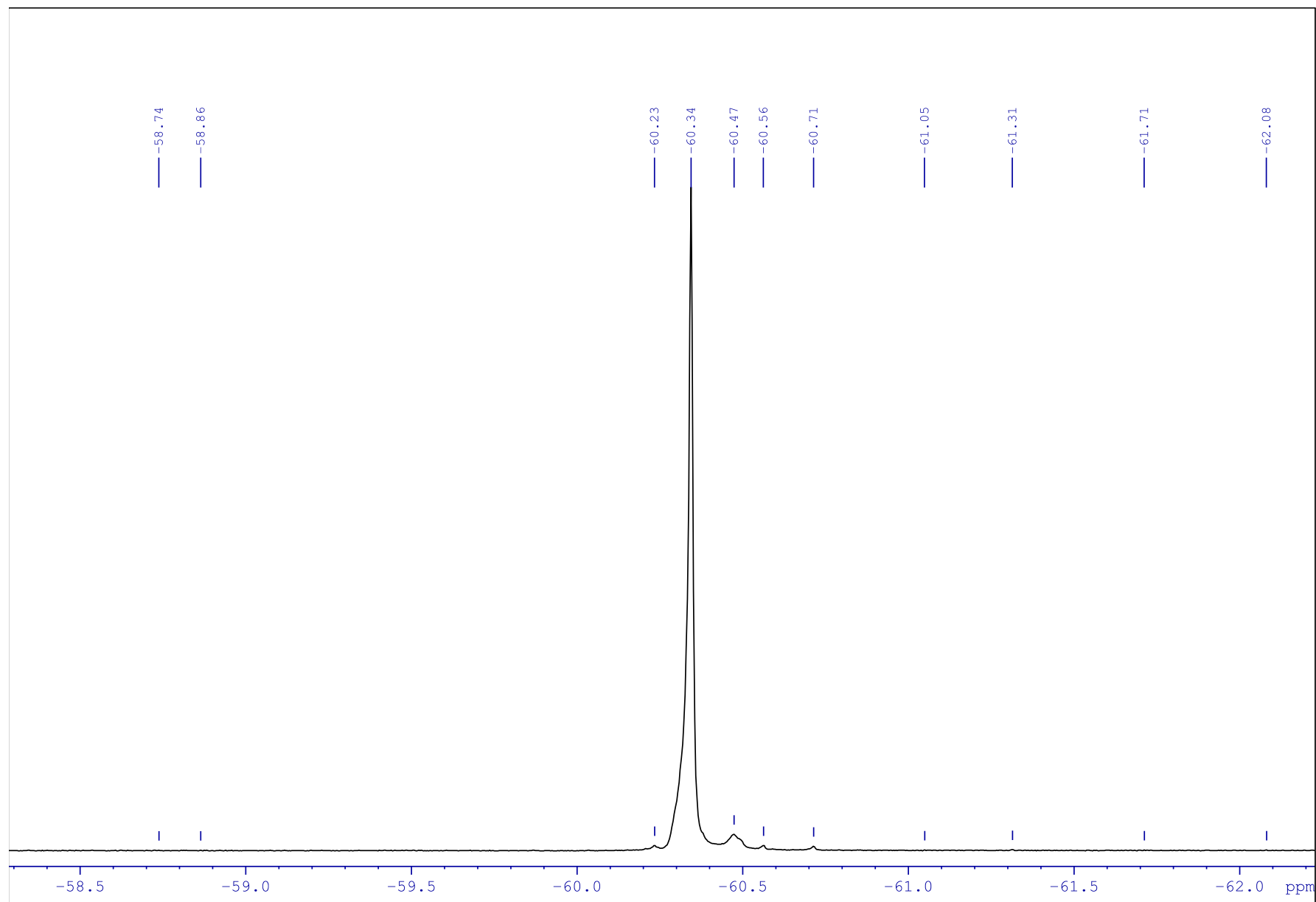


<sup>1</sup>H NMR spectrum of **3j** (DMSO-*d*<sub>6</sub>).

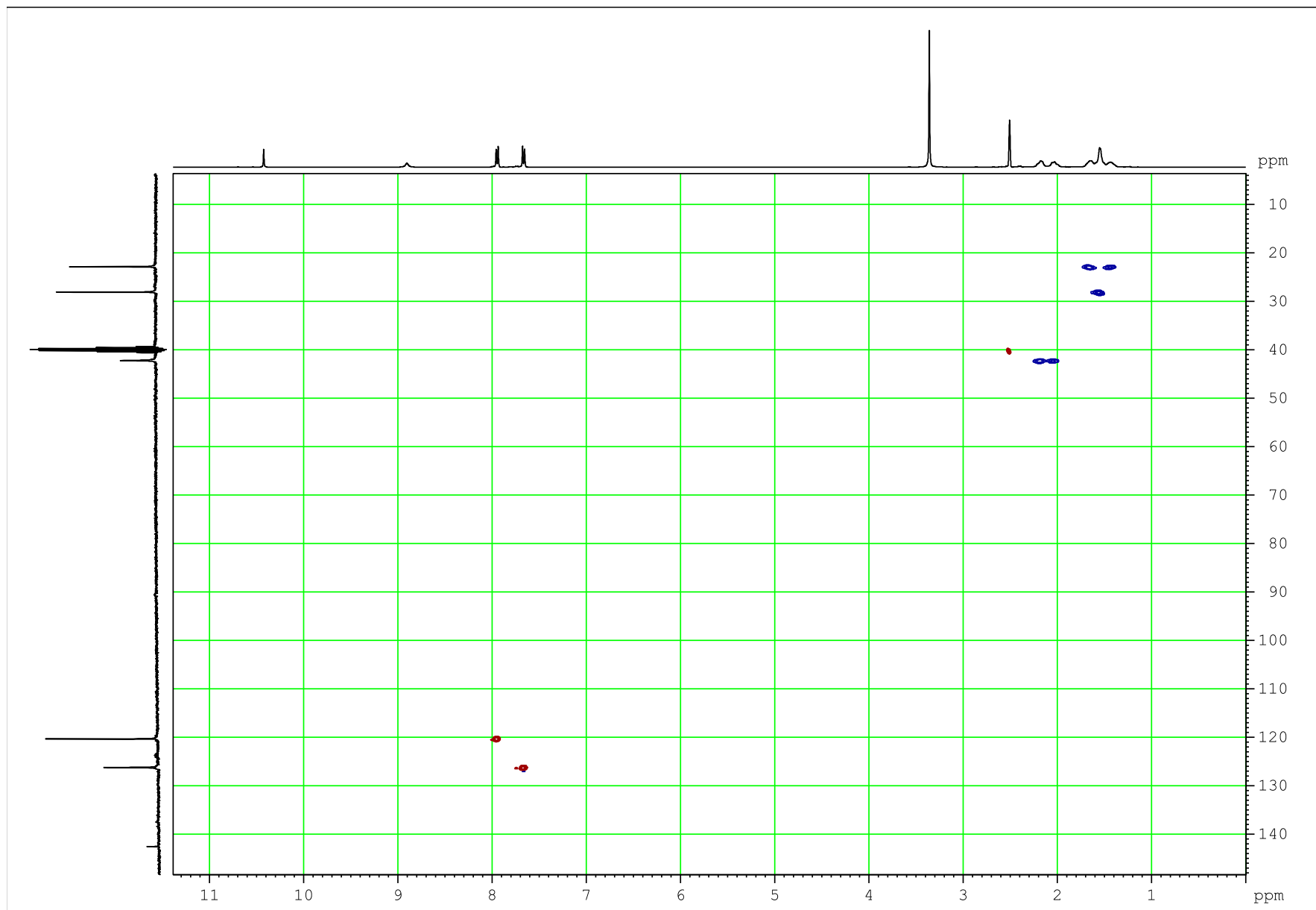


$^{13}\text{C}$  NMR spectrum of **3j** (DMSO- $d_6$ ).

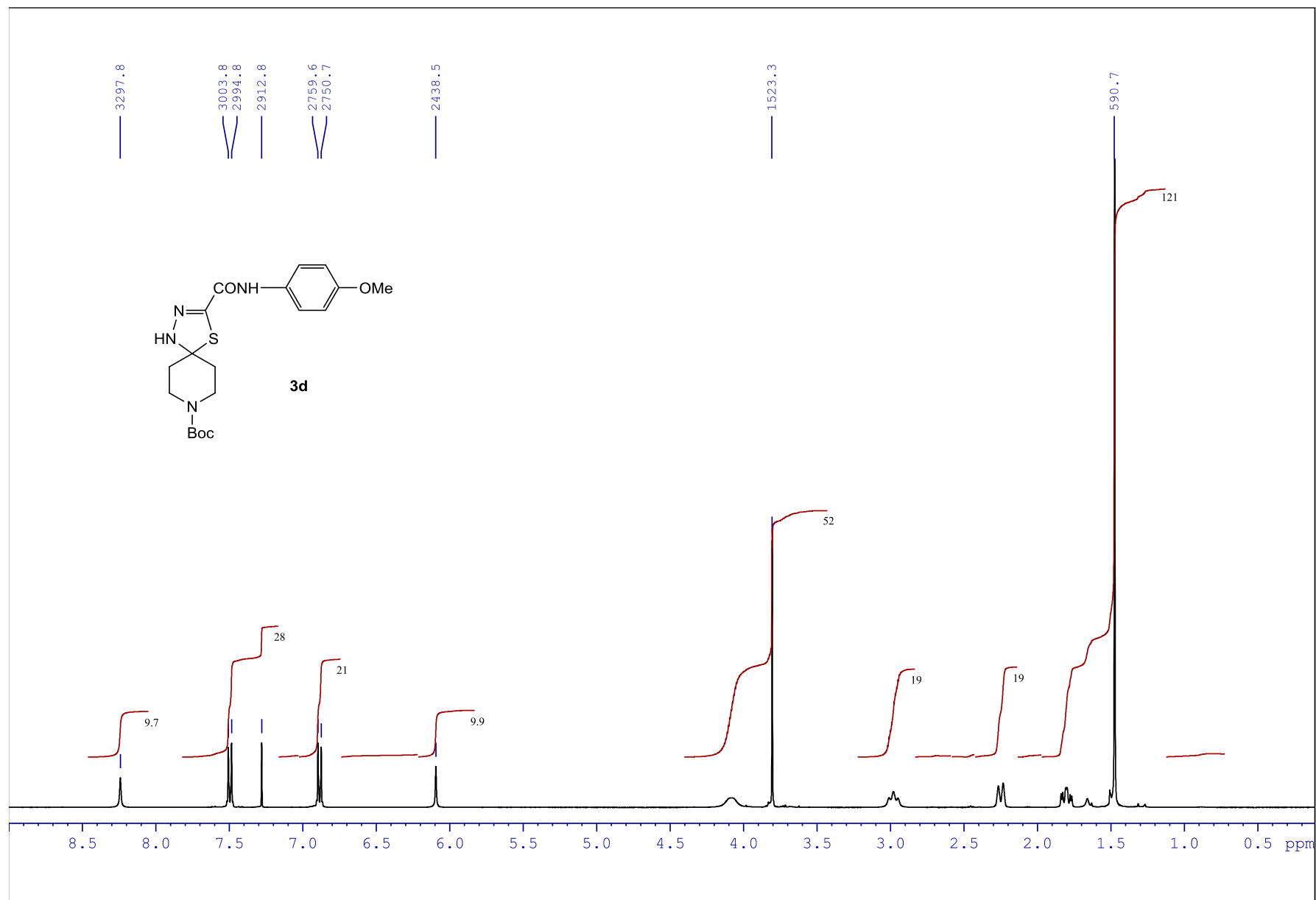




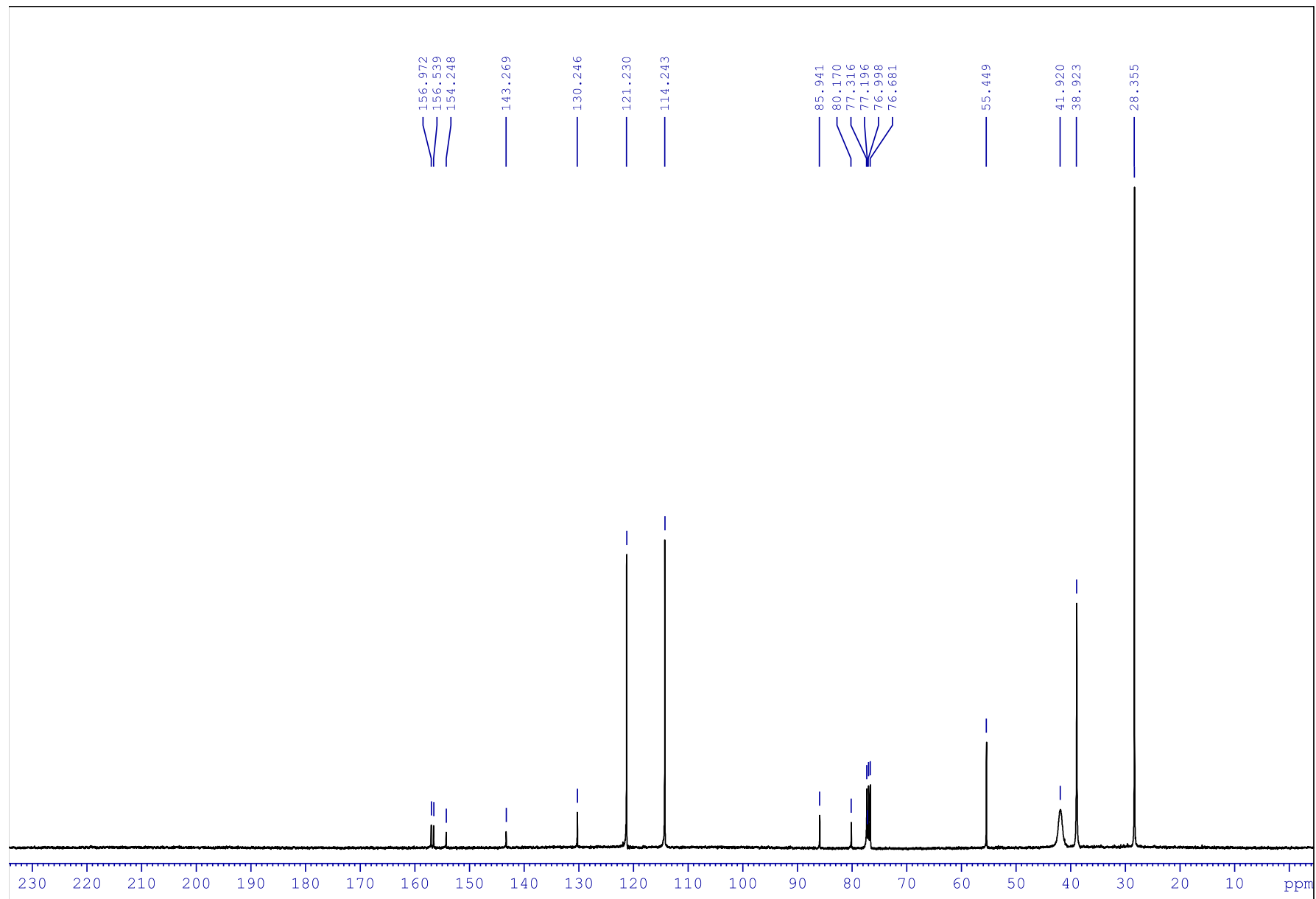
$^{19}\text{F}$  NMR spectrum of **3j** ( $\text{DMSO-}d_6$ ).



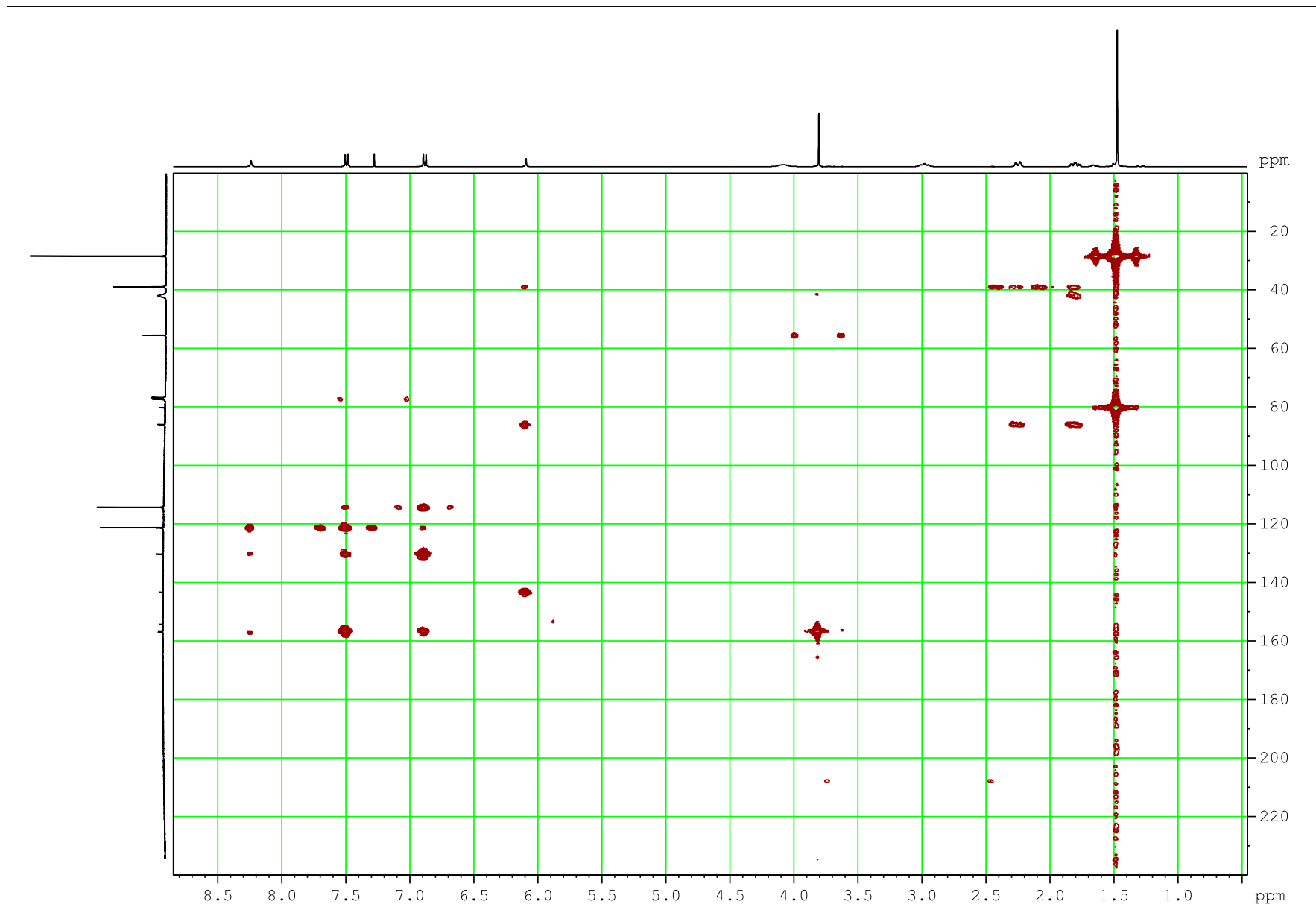
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3j** ( $\text{DMSO-}d_6$ ).



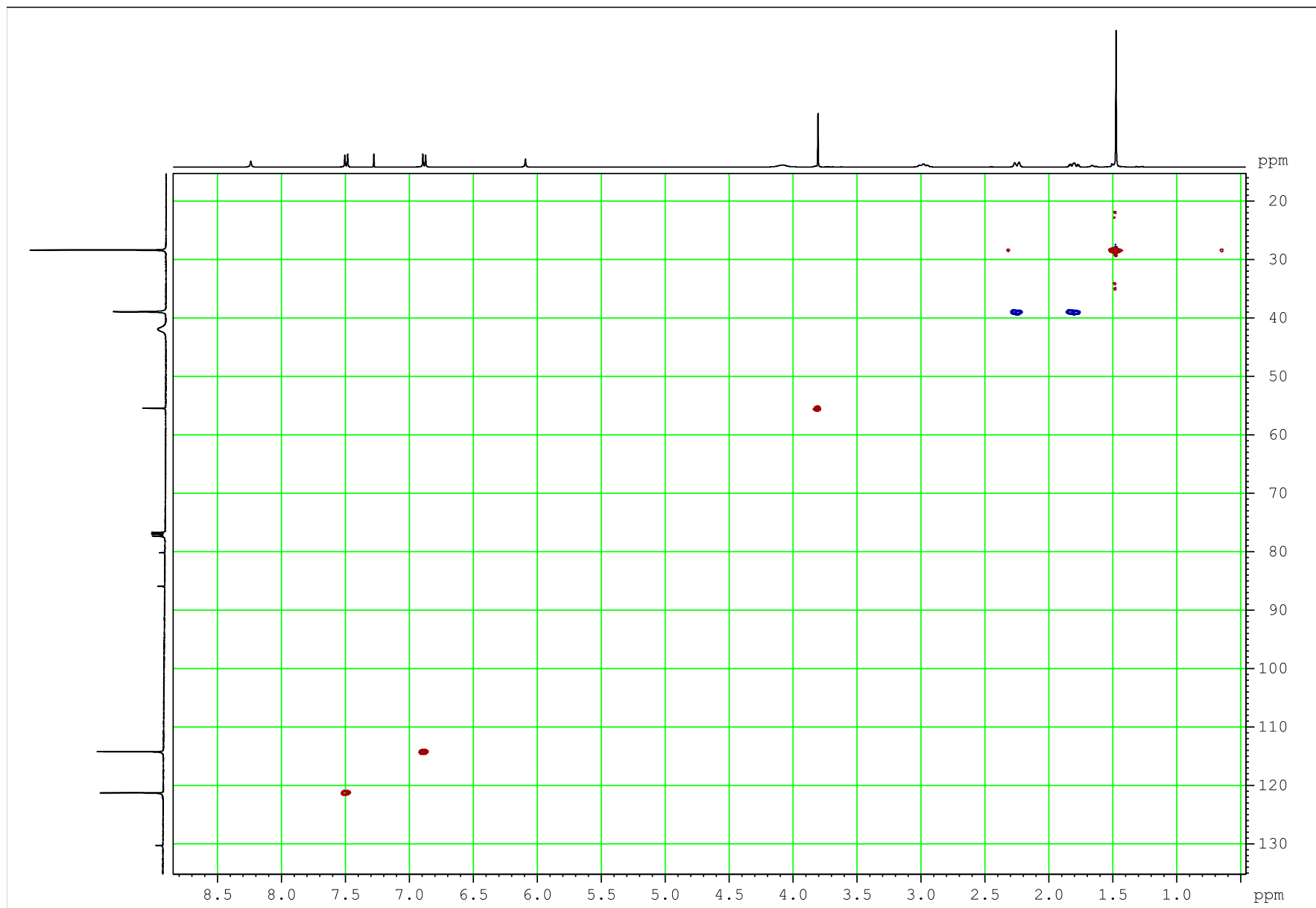
<sup>1</sup>H NMR spectrum of **3d** (DMSO-*d*<sub>6</sub>).



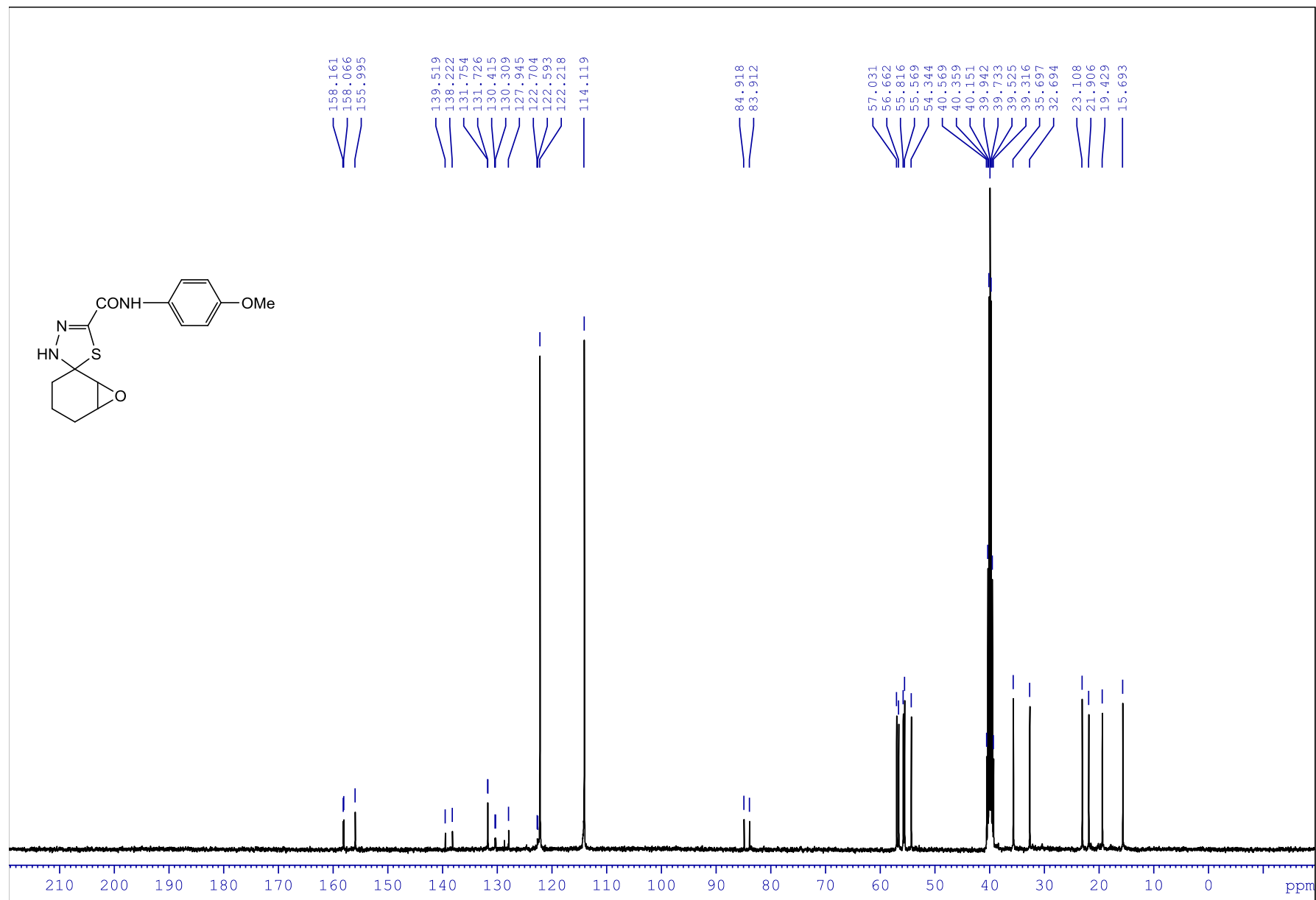
$^{13}\text{C}$  NMR spectrum of **3d** ( $\text{DMSO-}d_6$ ).



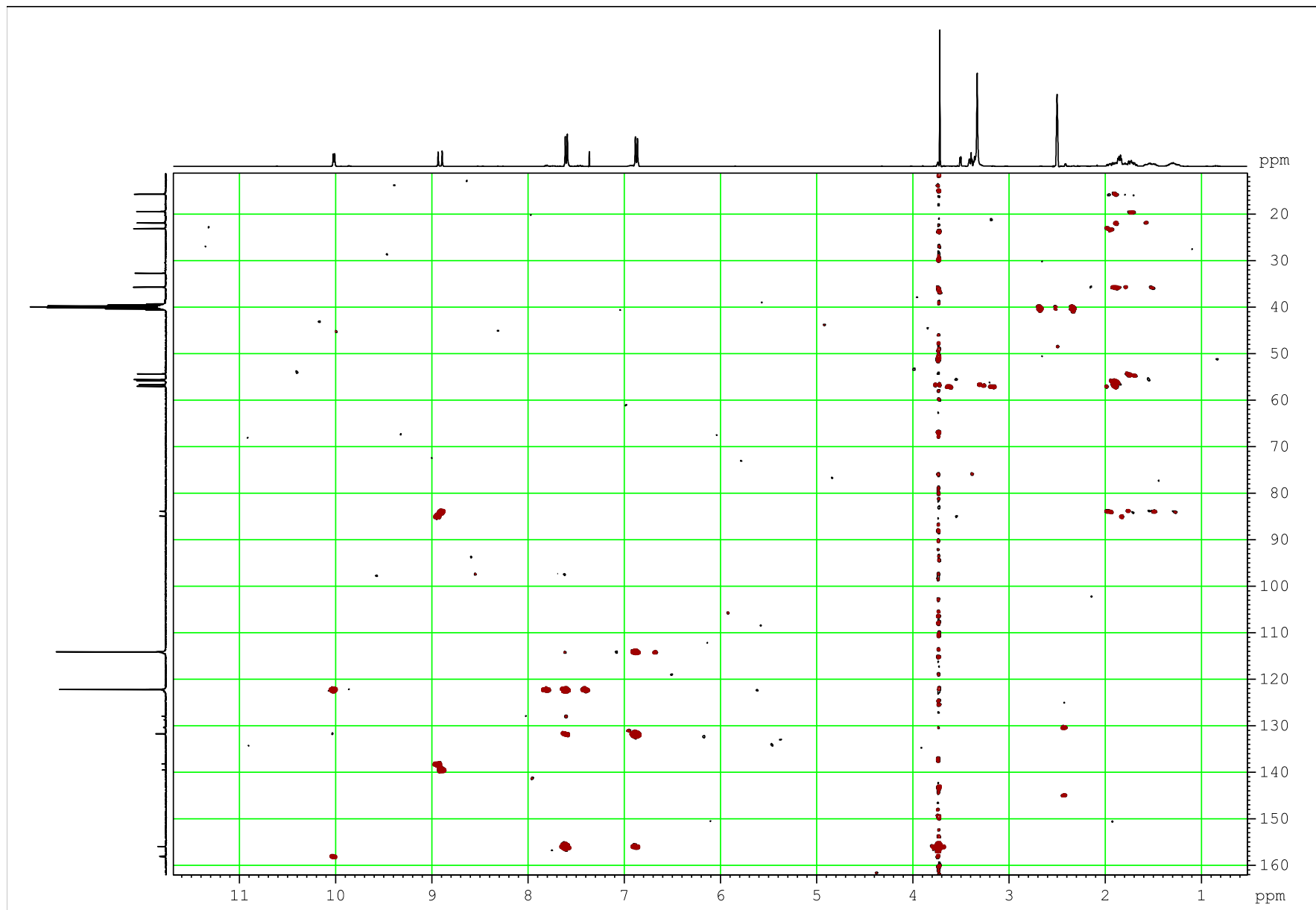
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3d** ( $\text{DMSO-}d_6$ ).



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3d** ( $\text{DMSO-}d_6$ ).

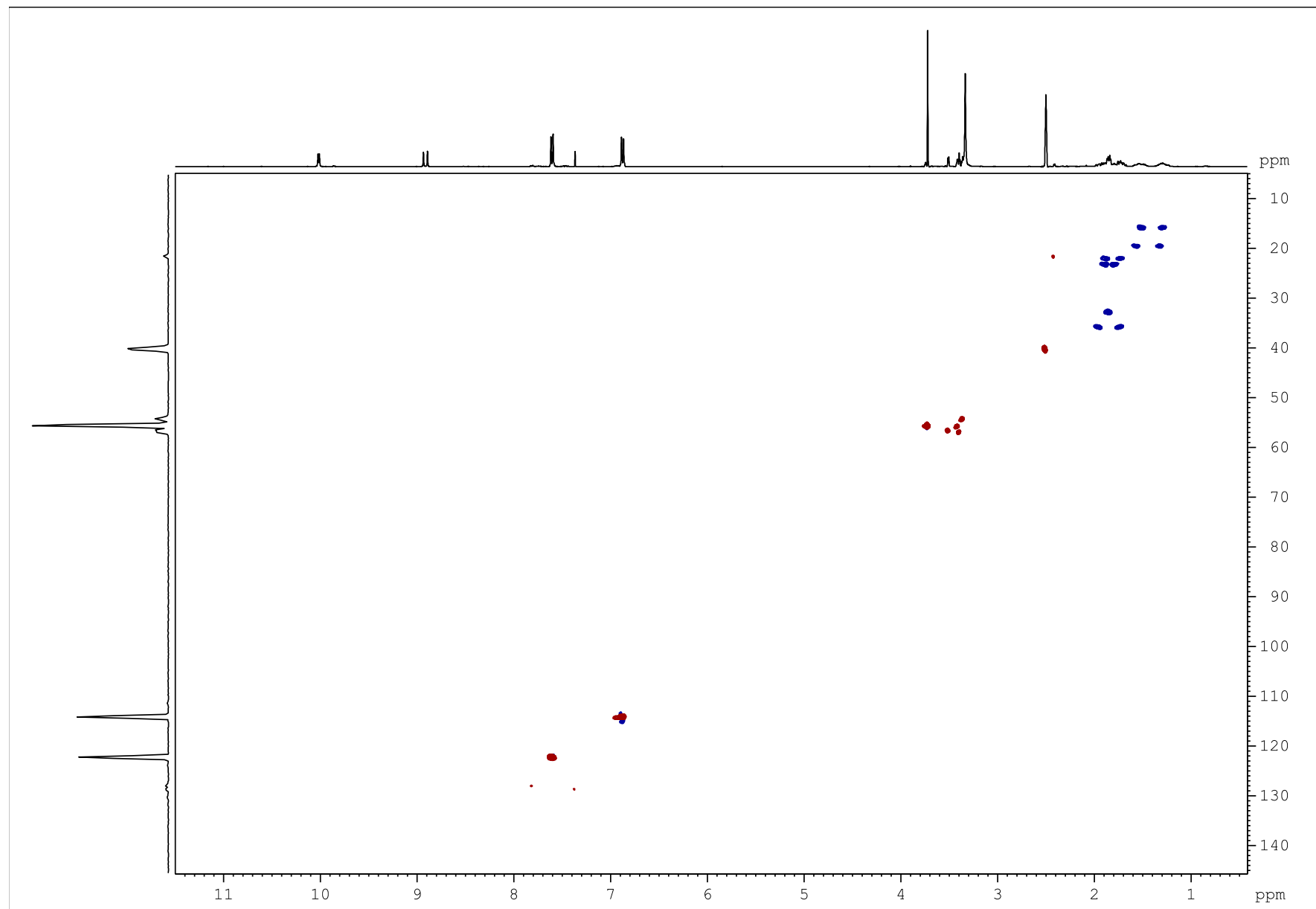


$^{13}\text{C}$  NMR spectrum of **3e** ( $\text{DMSO-}d_6$ ).

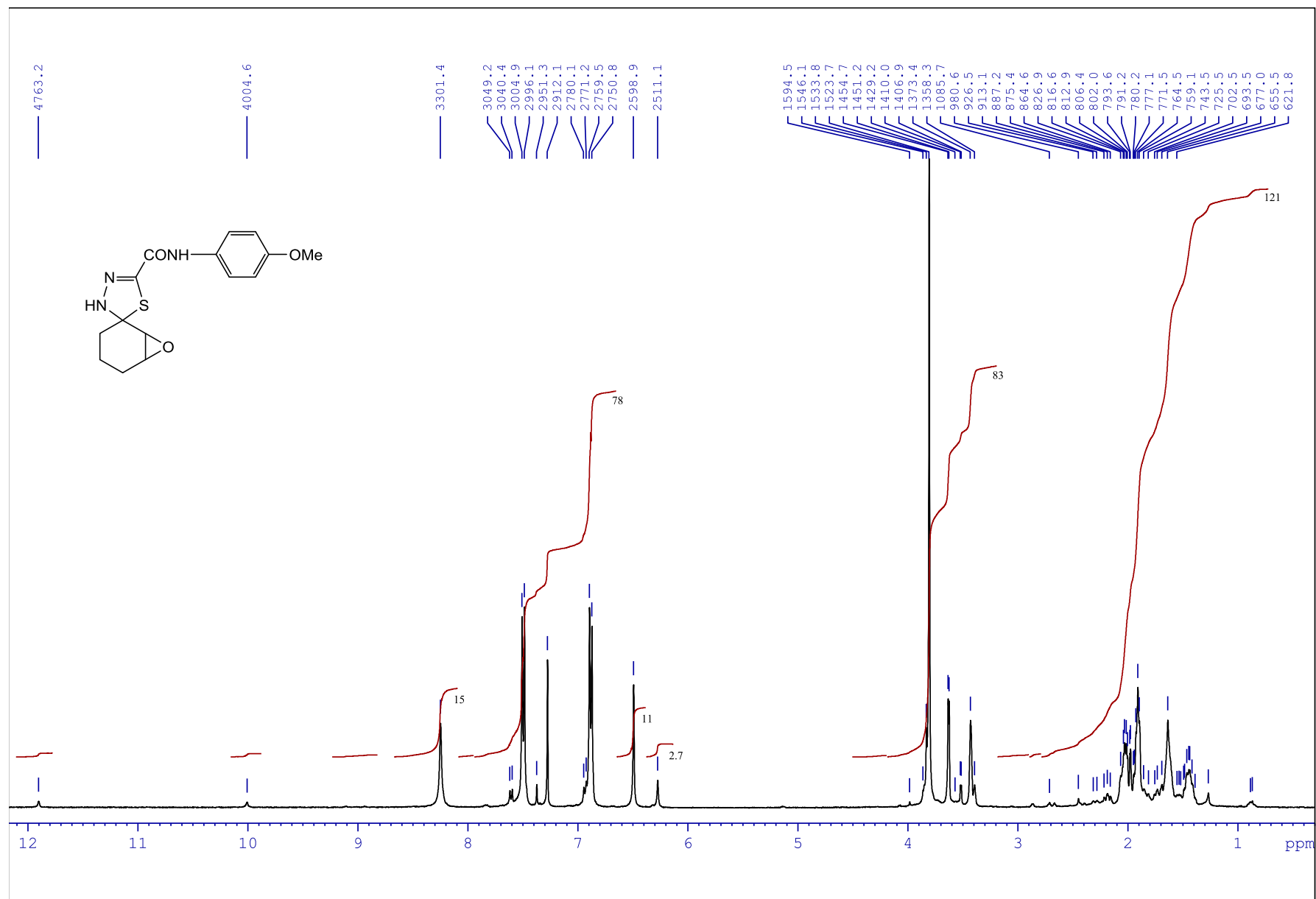


2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum of **3e** ( $\text{DMSO-}d_6$ ).

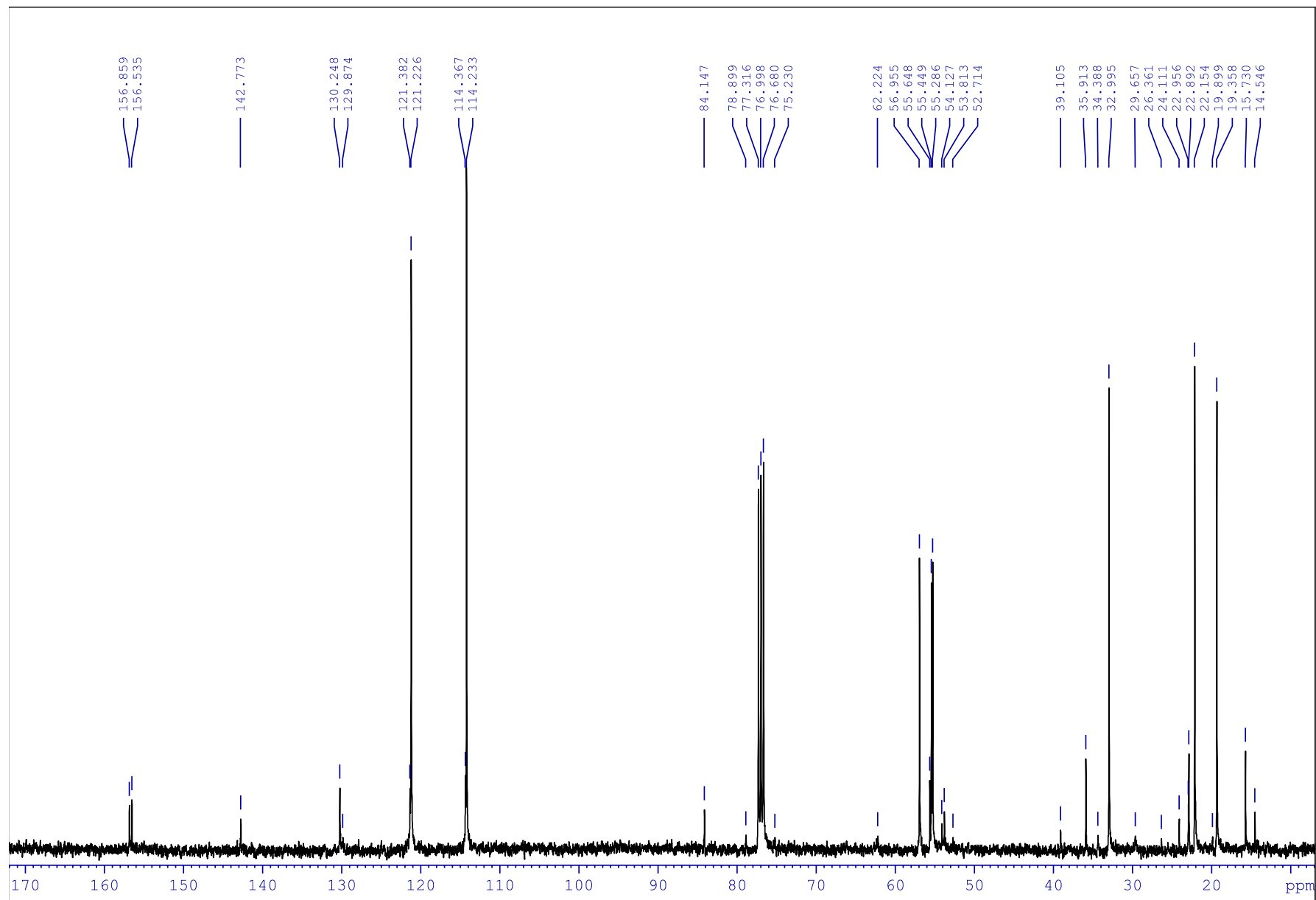




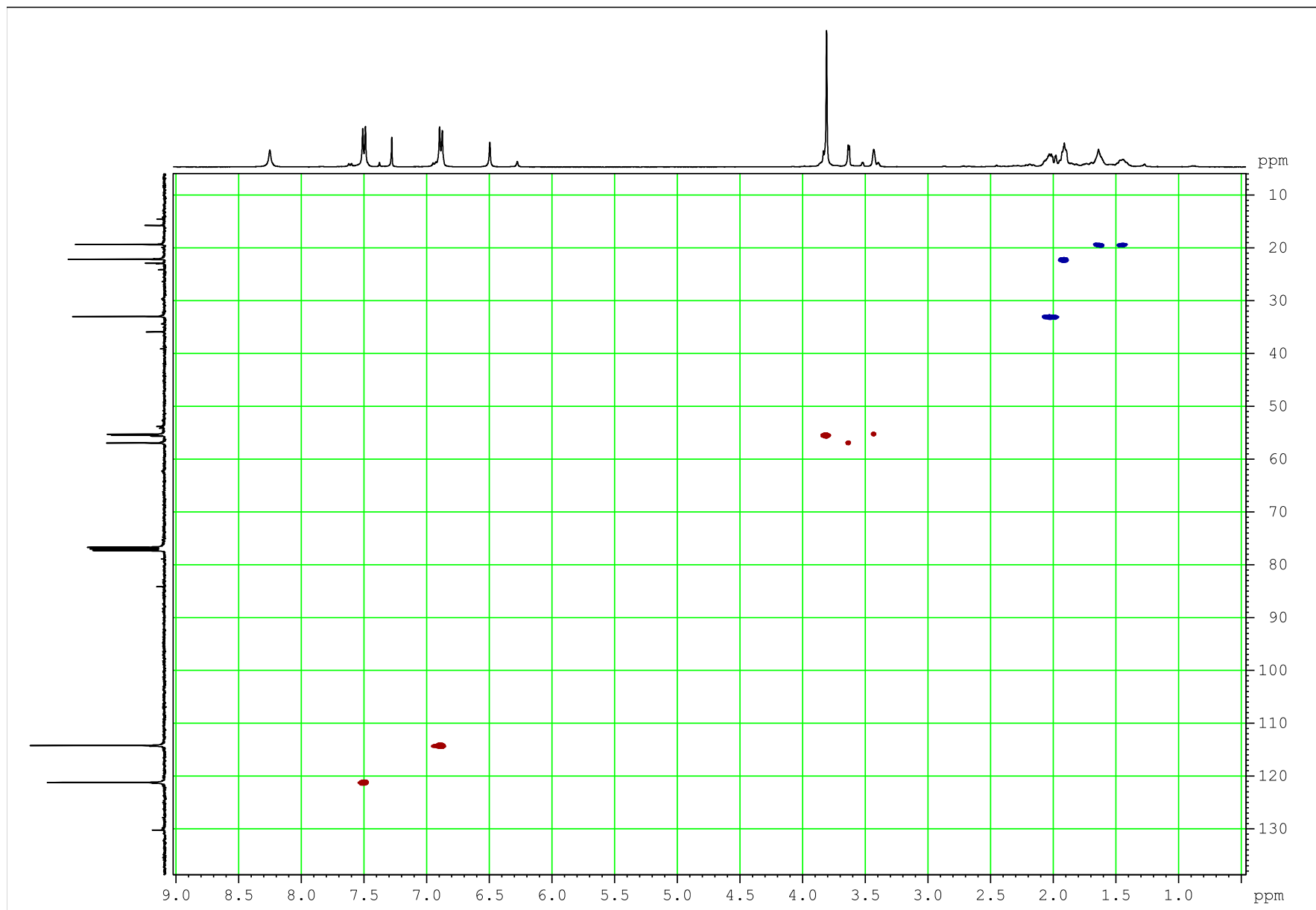
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3e** ( $\text{DMSO-}d_6$ ).



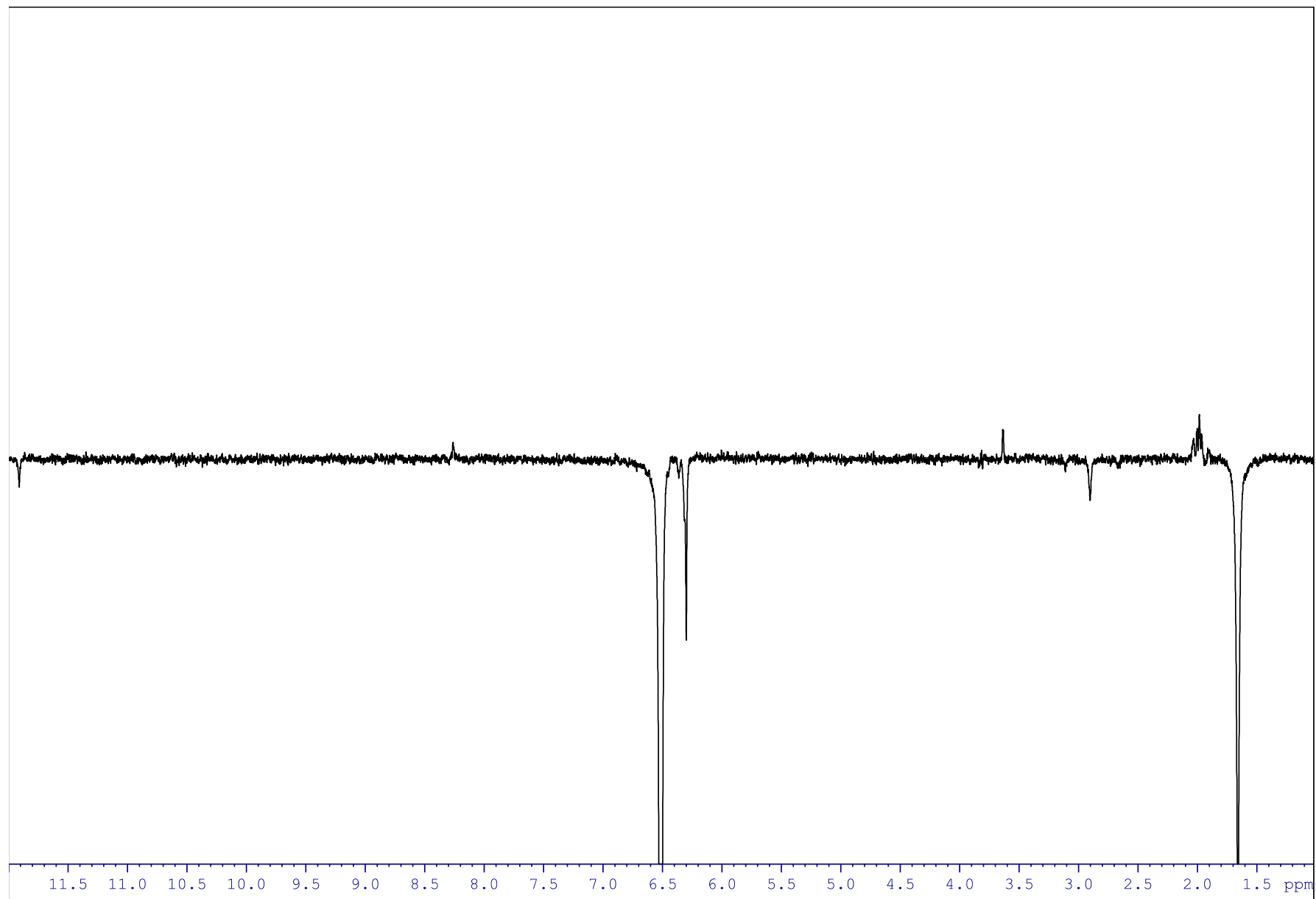
<sup>1</sup>H NMR spectrum of **3e** (CDCl<sub>3</sub>).



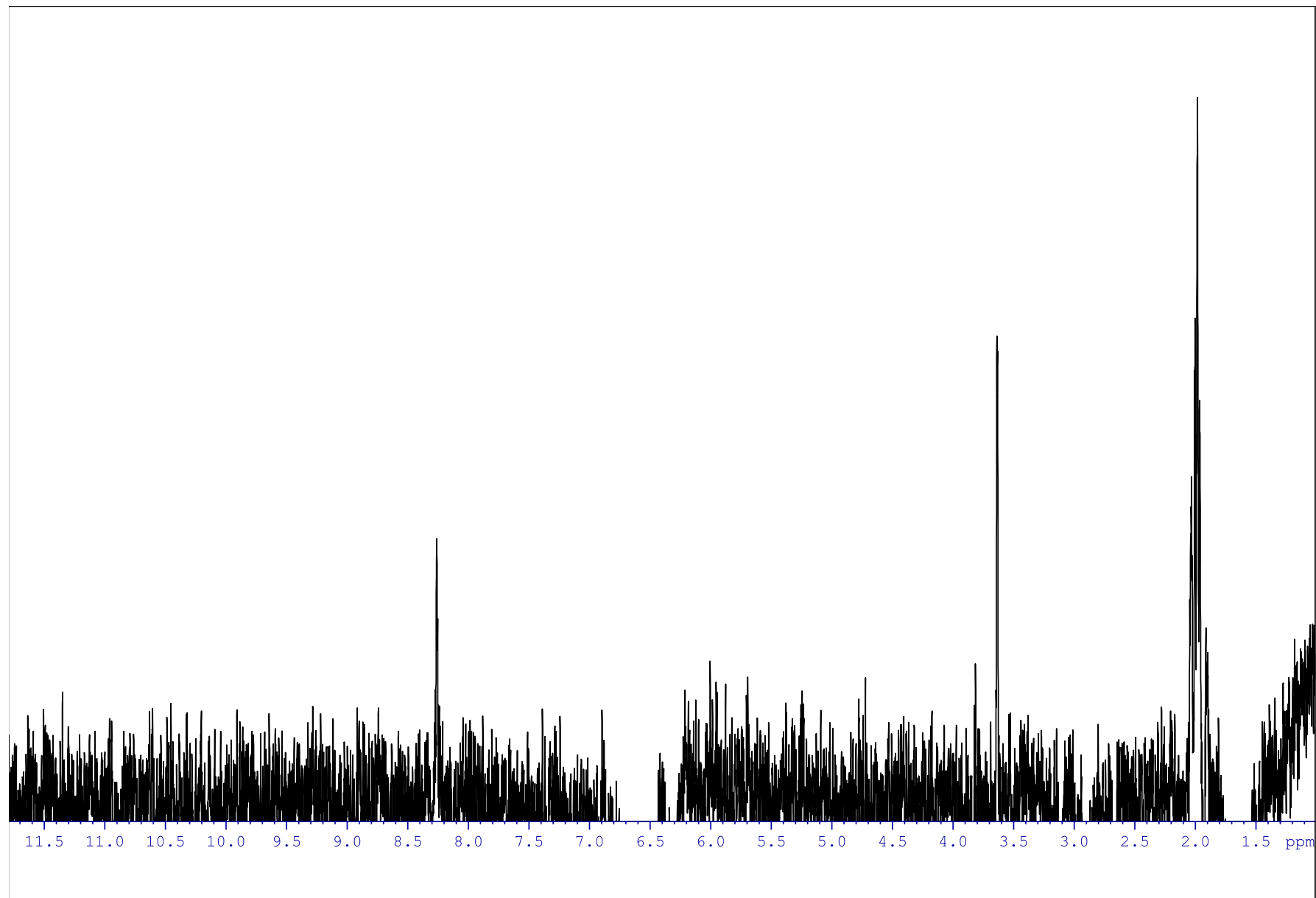
$^{13}\text{C}$  NMR spectrum of **3e** ( $\text{CDCl}_3$ ).



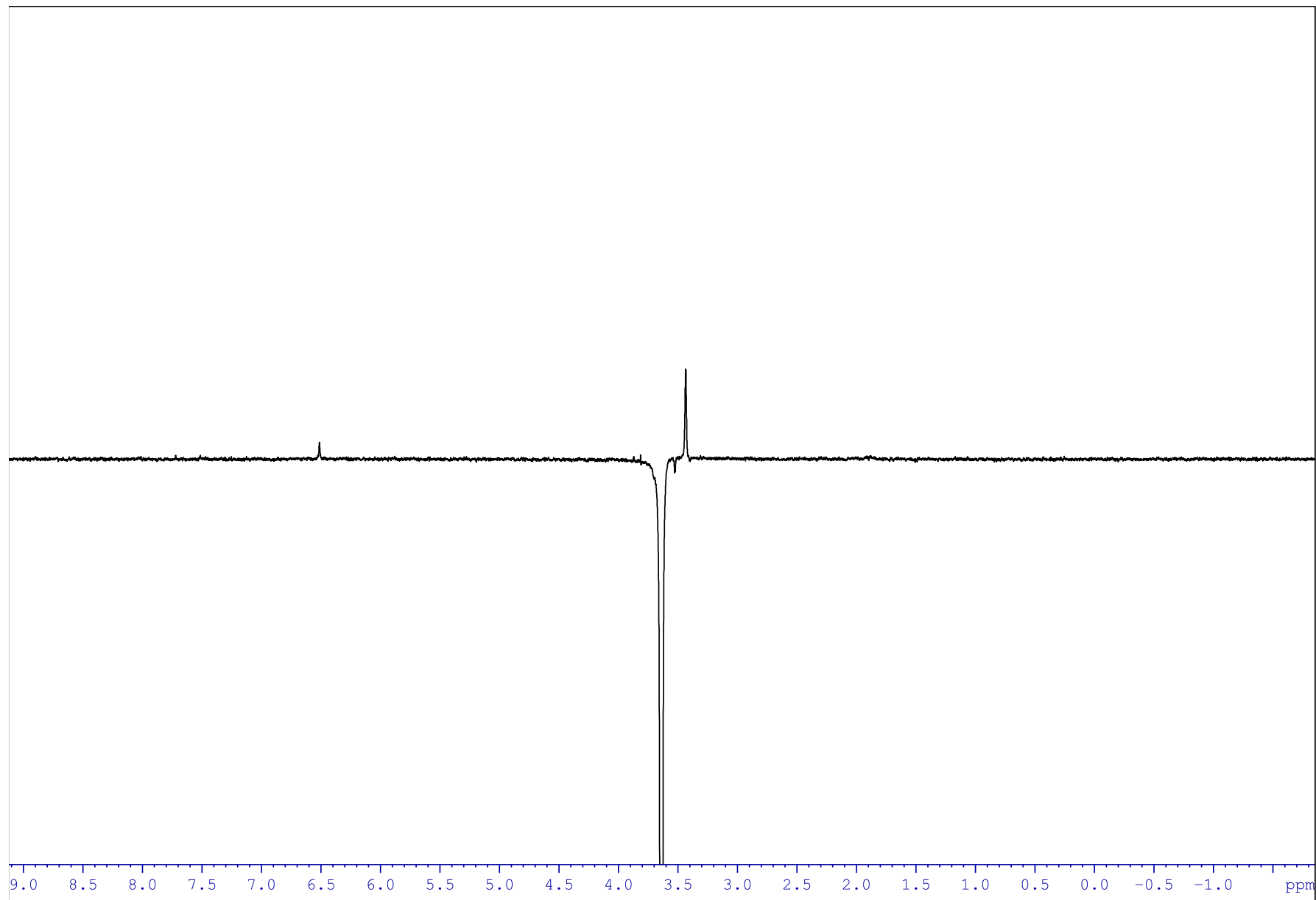
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum of **3e** ( $\text{CDCl}_3$ ).



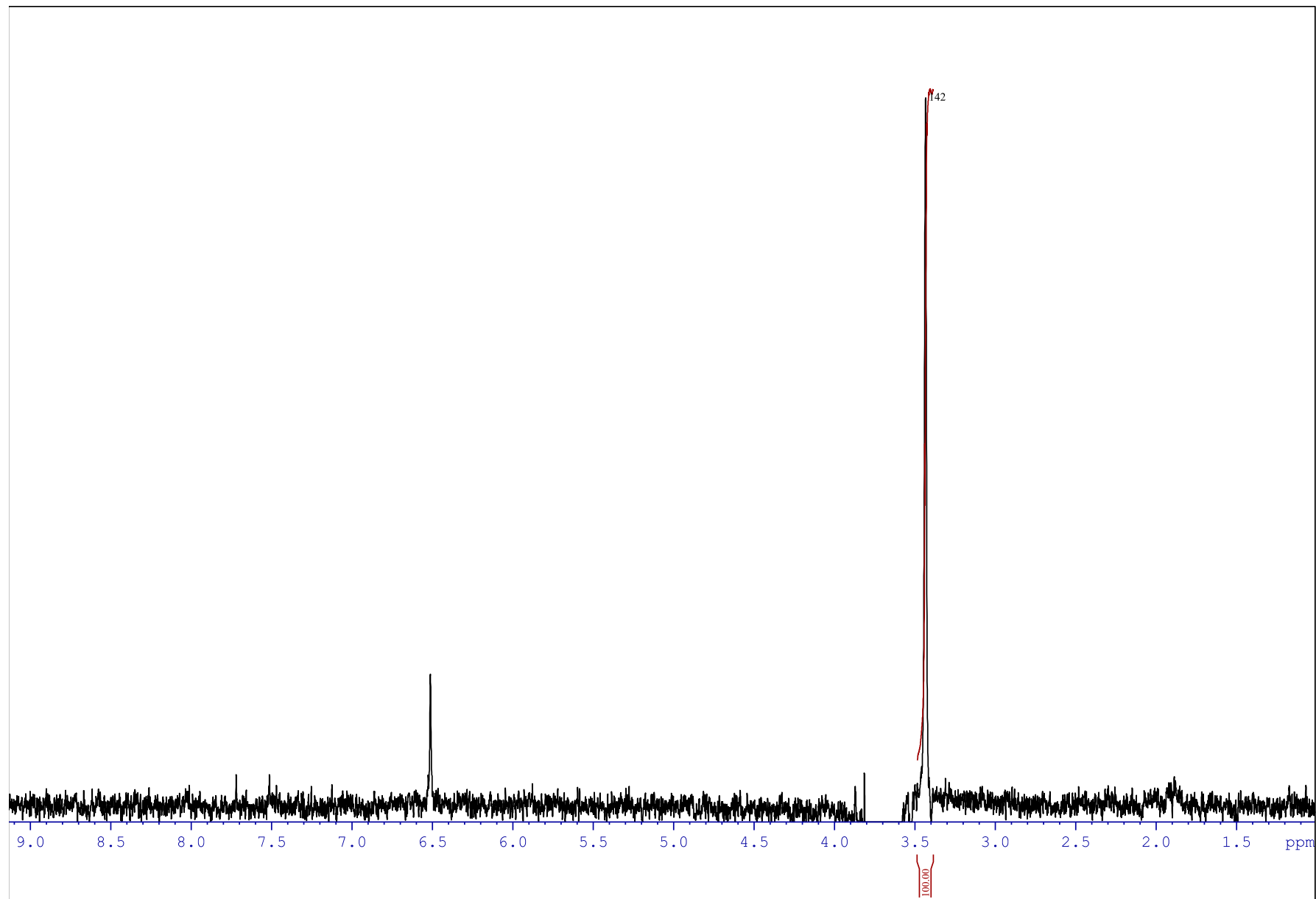
1D  $^1\text{H}$  NOESY (6.51 ppm) NMR spectrum of **3e** ( $\text{CDCl}_3$ ).



1D  $^1\text{H}$  NOESY (6.51 ppm) NMR spectrum of **3e** ( $\text{CDCl}_3$ ).

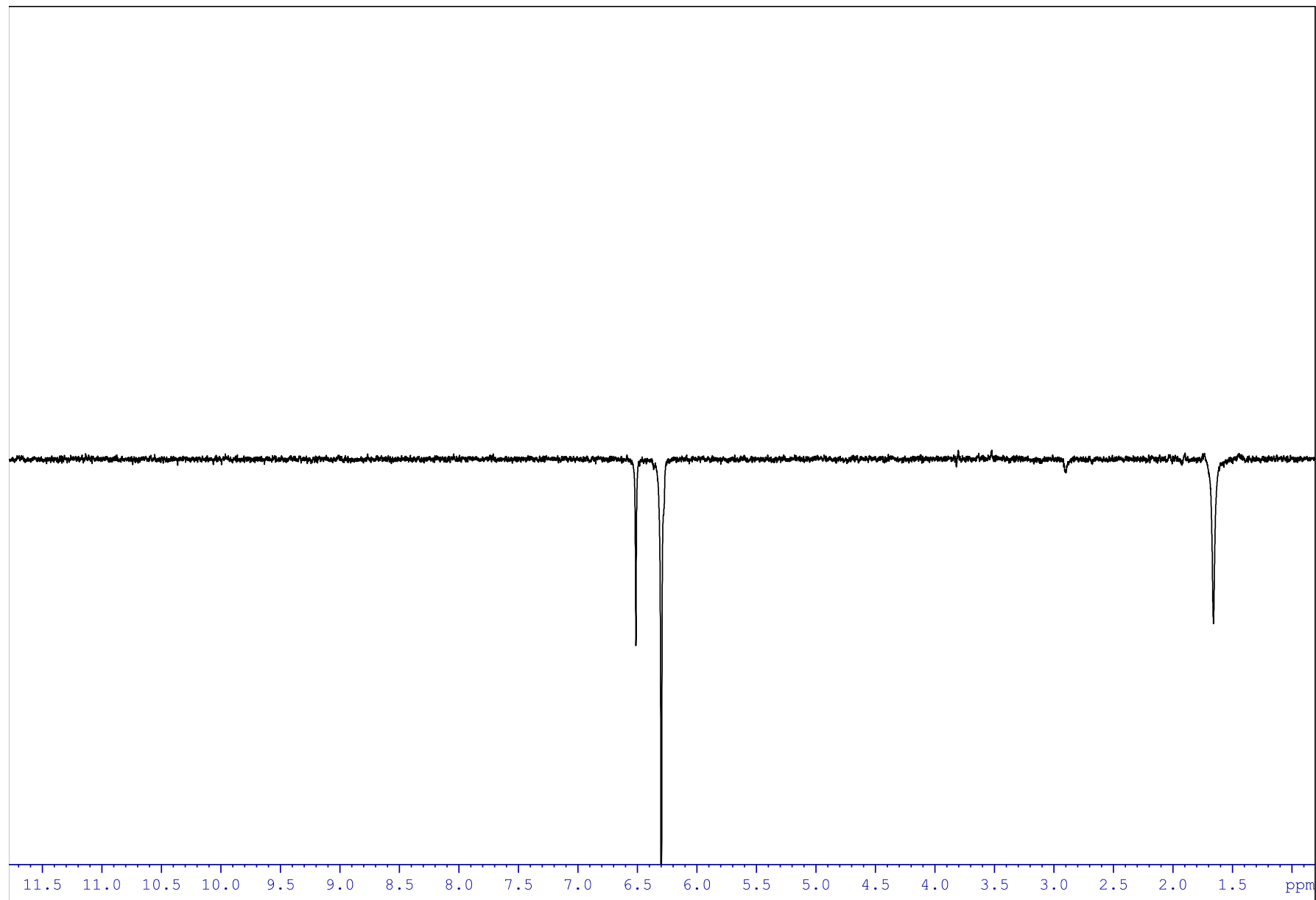


1D <sup>1</sup>H NOESY (3.63 ppm) NMR spectrum of **3e** (CDCl<sub>3</sub>).



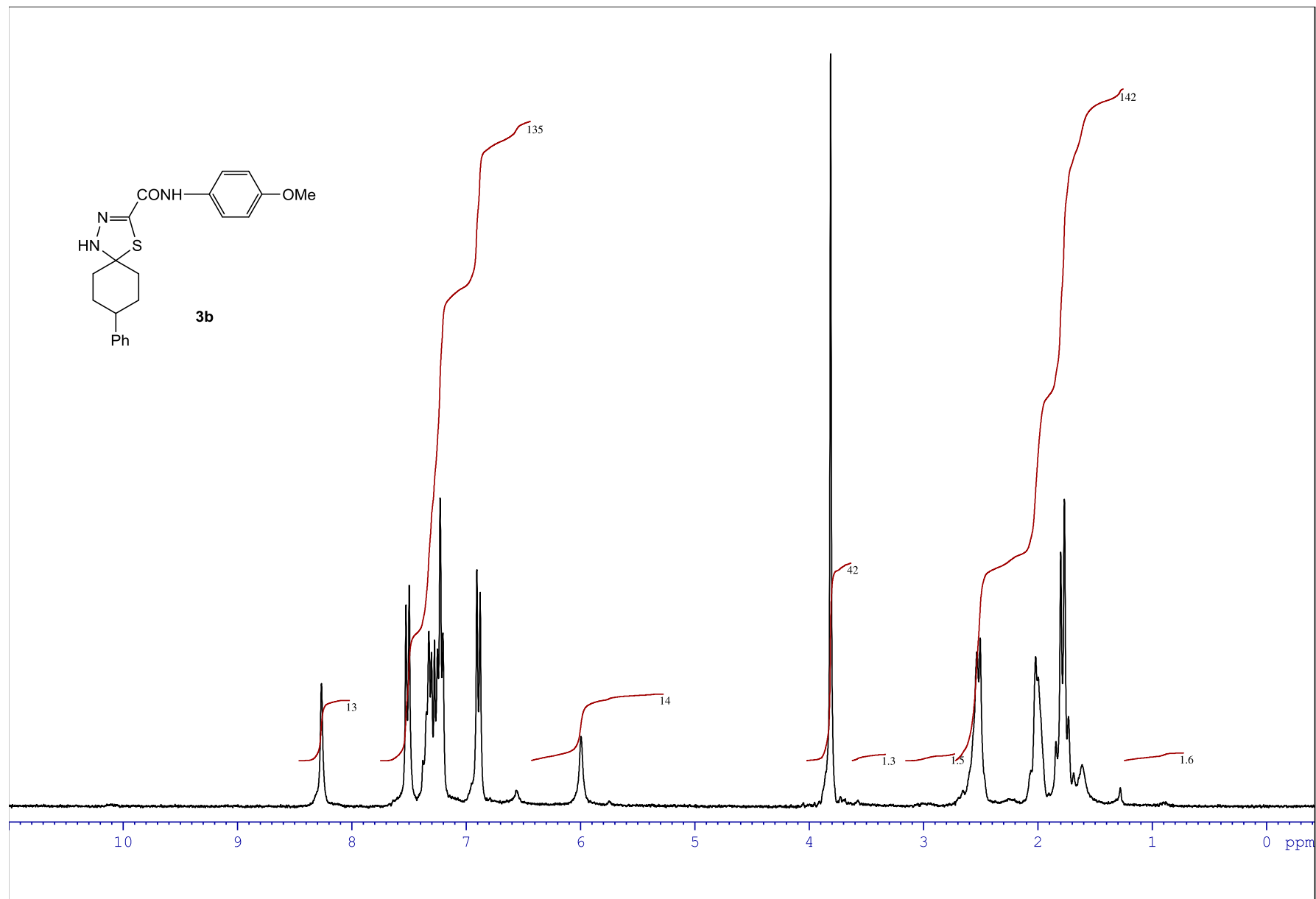
1D <sup>1</sup>H NOESY (3.63 ppm) NMR spectrum of **3e** (CDCl<sub>3</sub>).



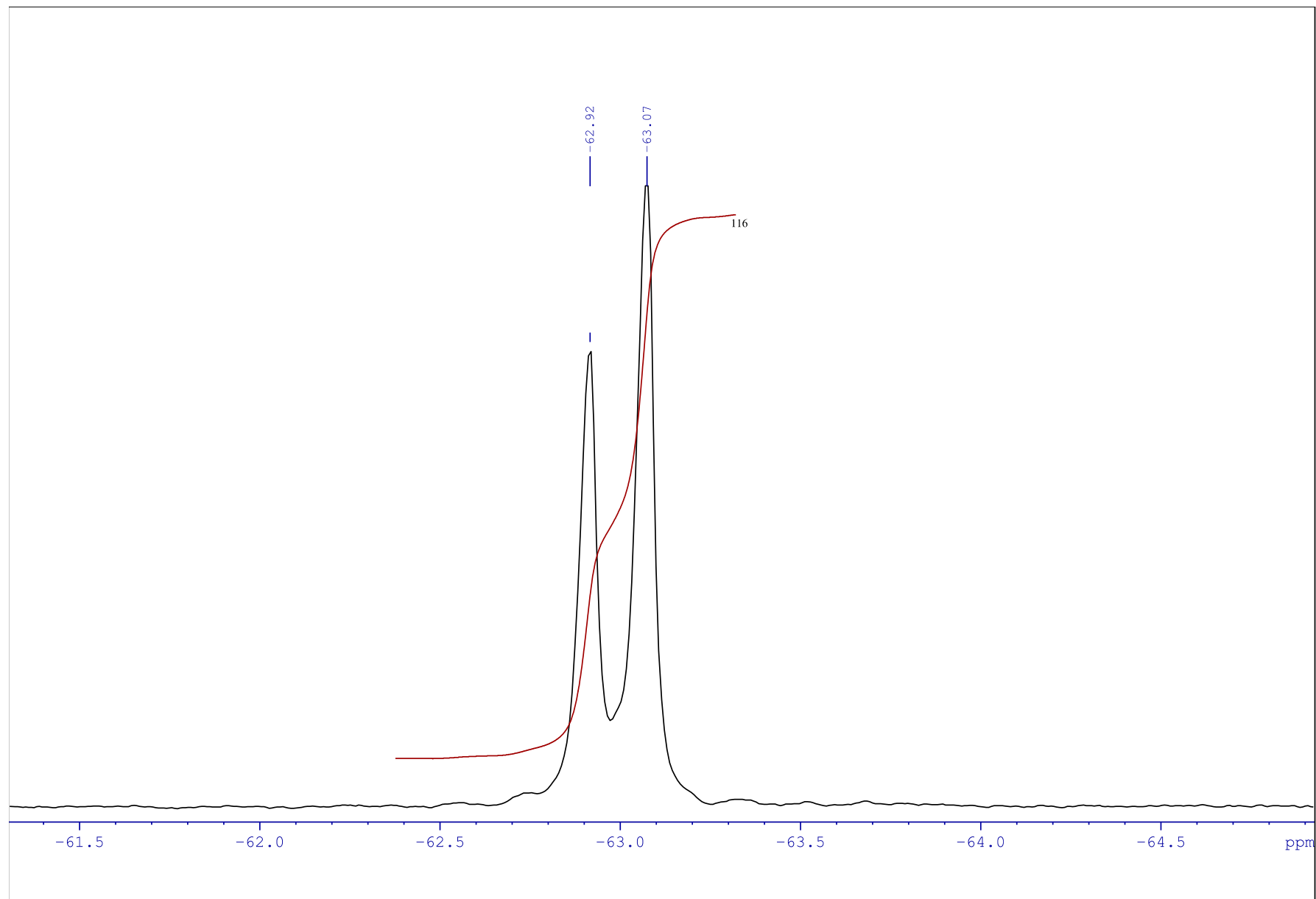


1D  $^1\text{H}$  NOESY (6.31 ppm) NMR spectrum of **3e** ( $\text{CDCl}_3$ ).

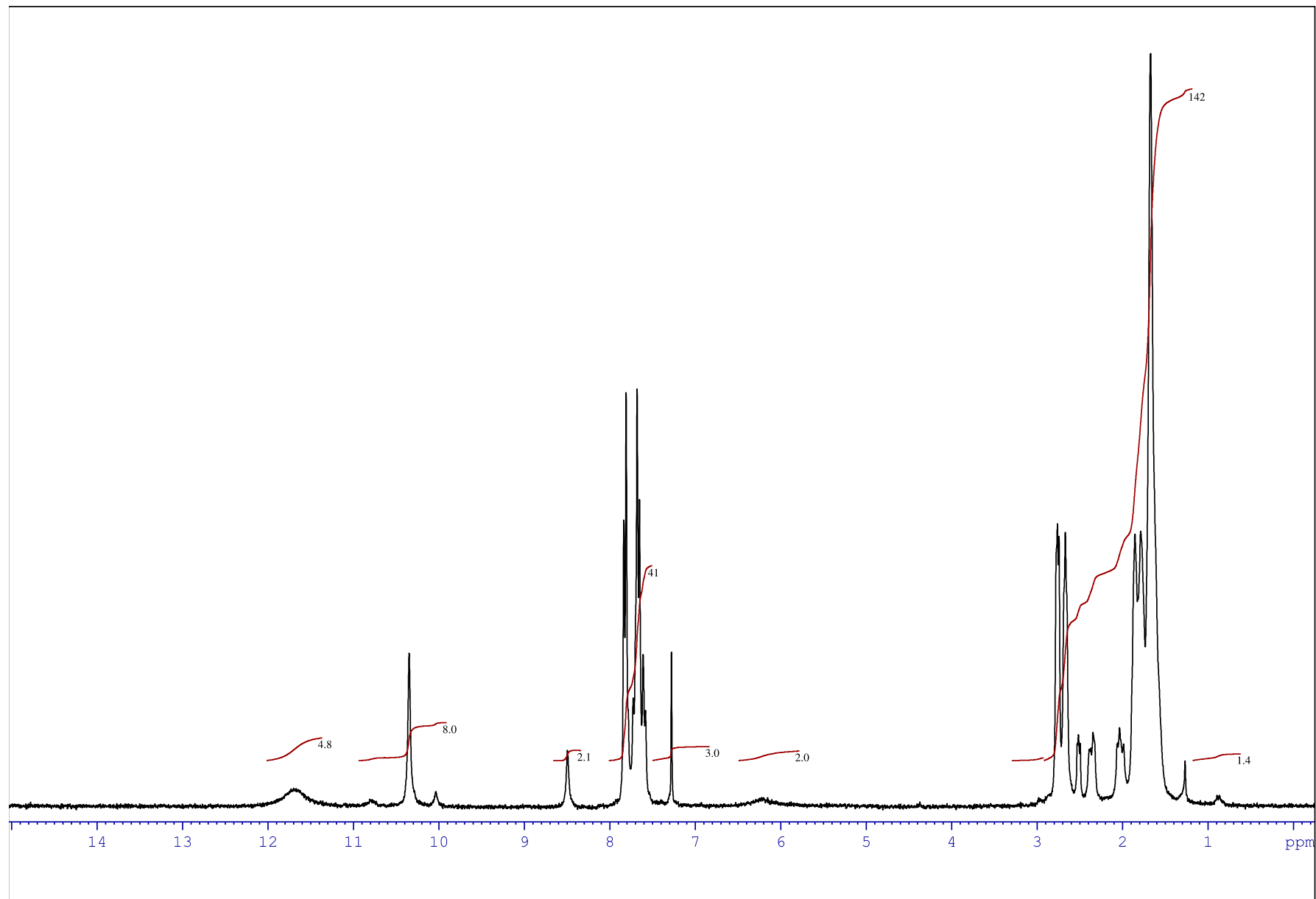
## 2. NMR spectra (Bruker AM-300)



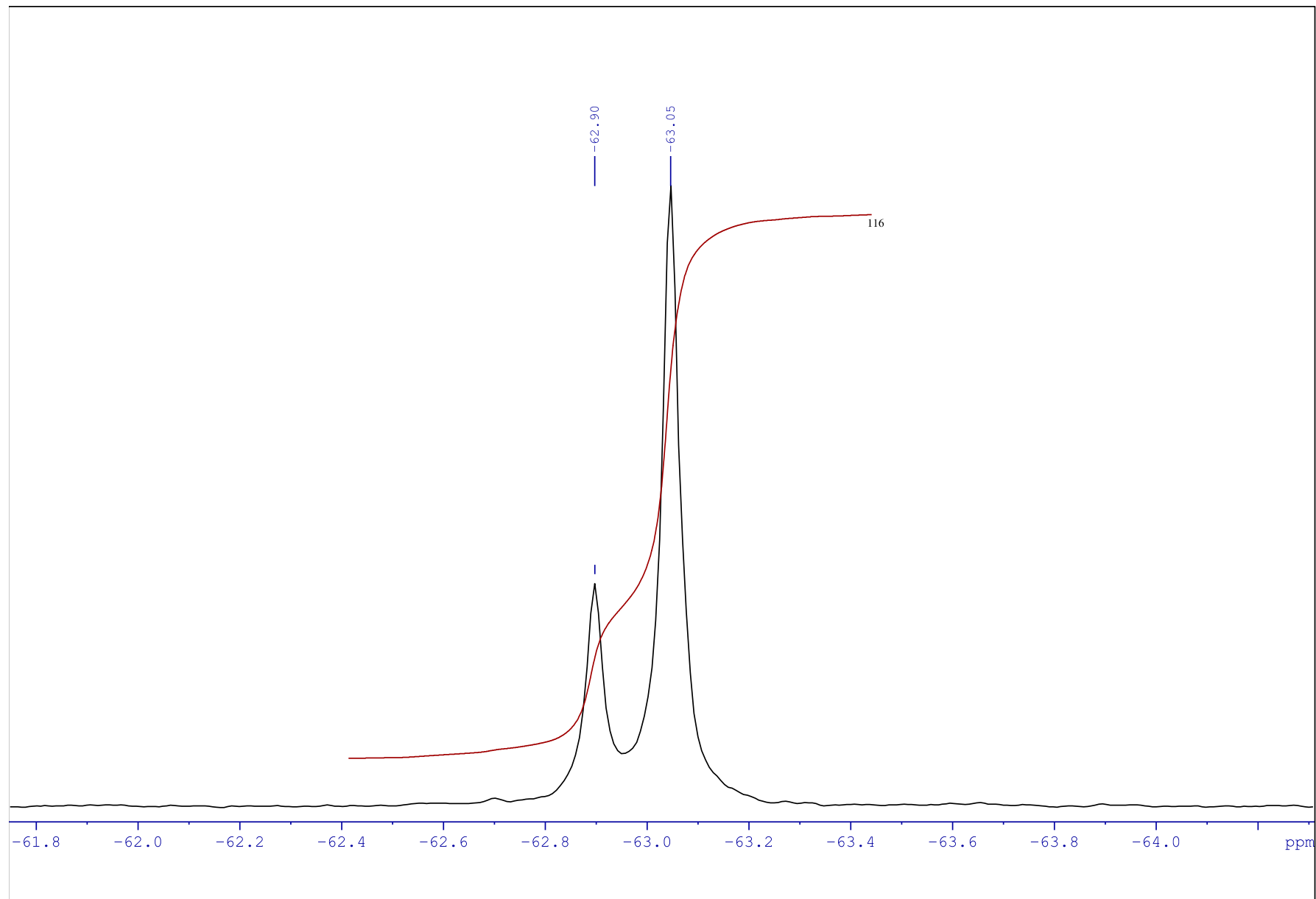
$^1\text{H}$  NMR spectrum of **3b** ( $\text{CDCl}_3$ )



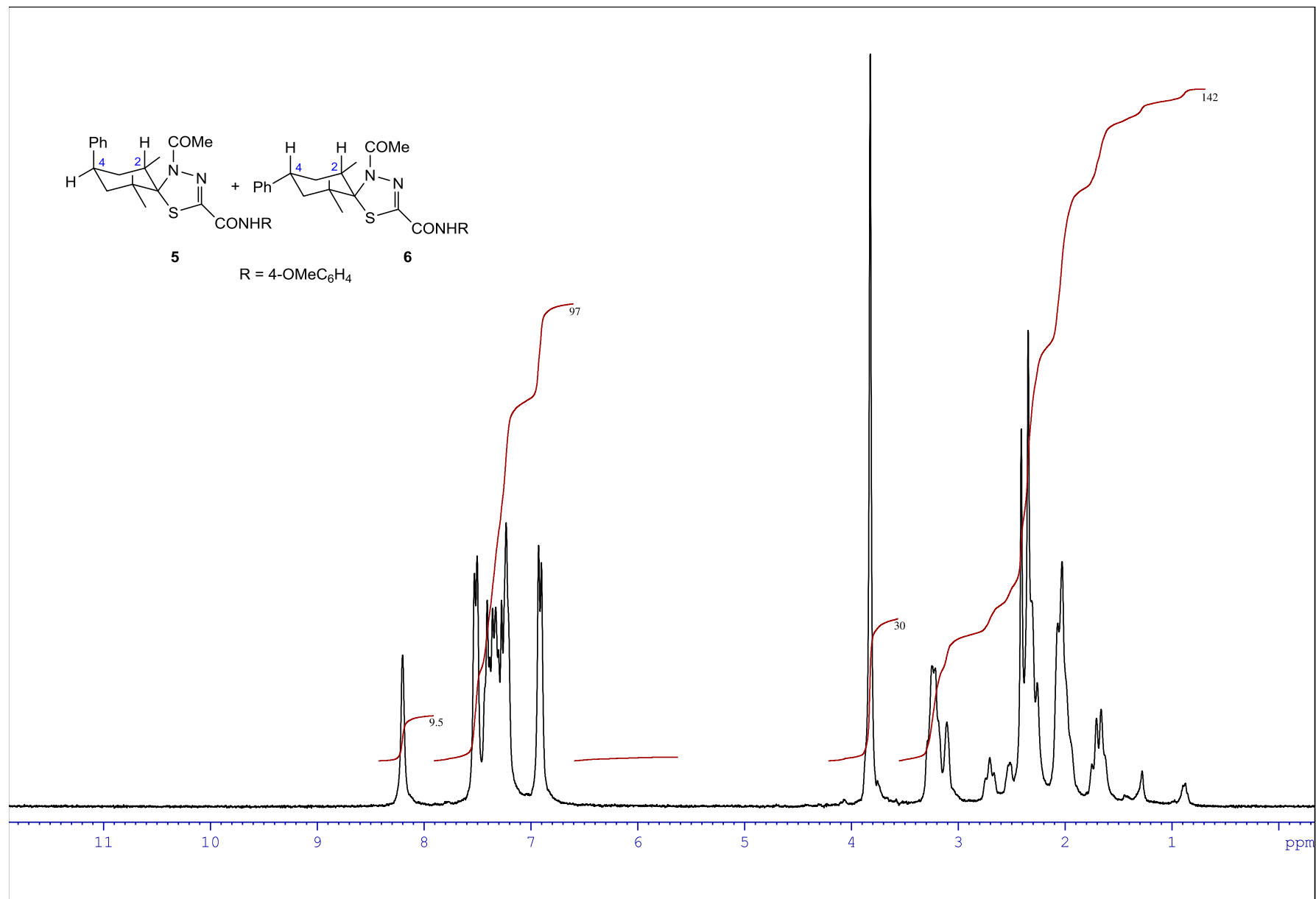
$^{19}\text{F}$  NMR spectrum of **3i** ( $\text{CDCl}_3$ ).



$^1\text{H}$  NMR spectrum of **3j** ( $\text{CDCl}_3$ ).

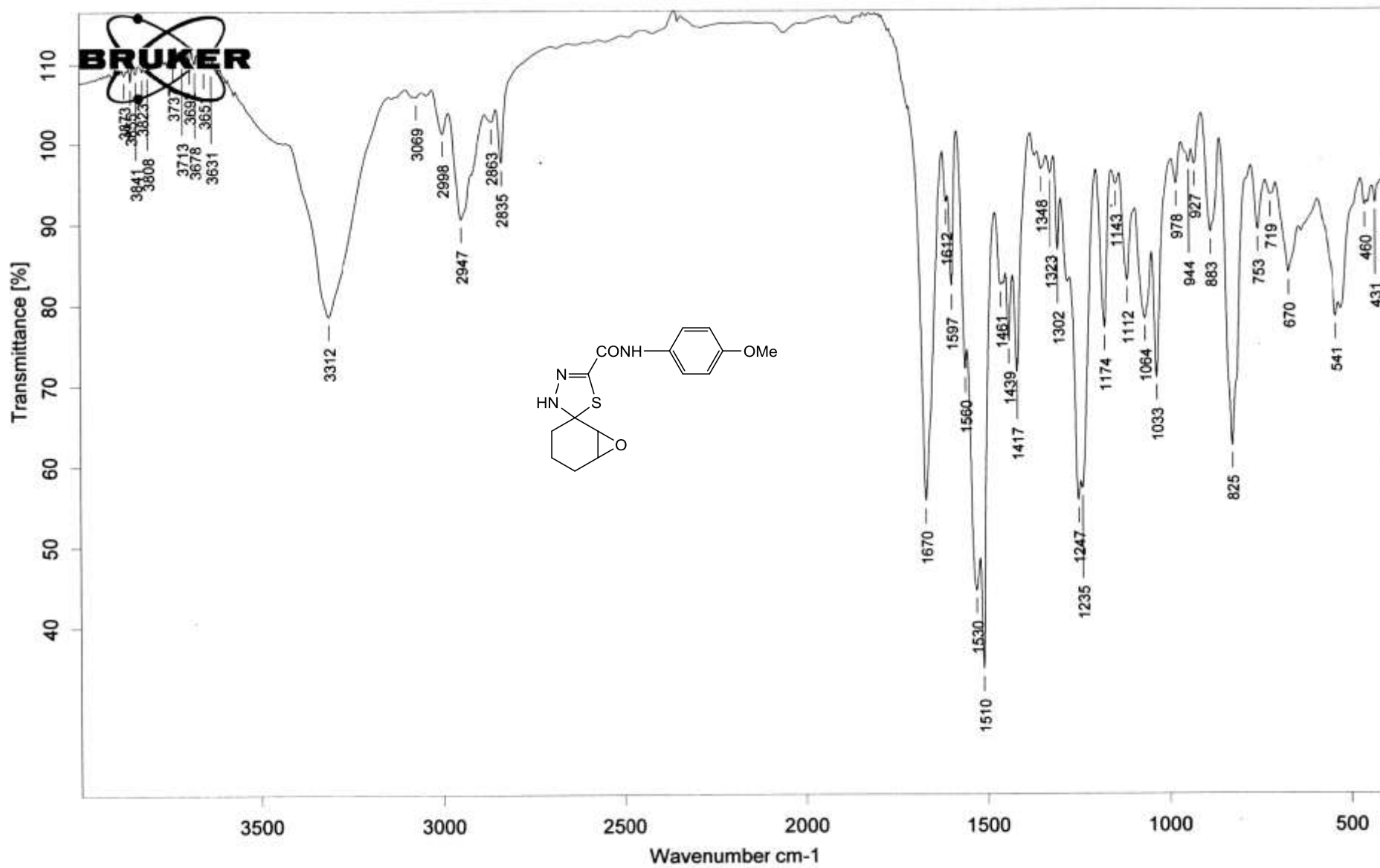


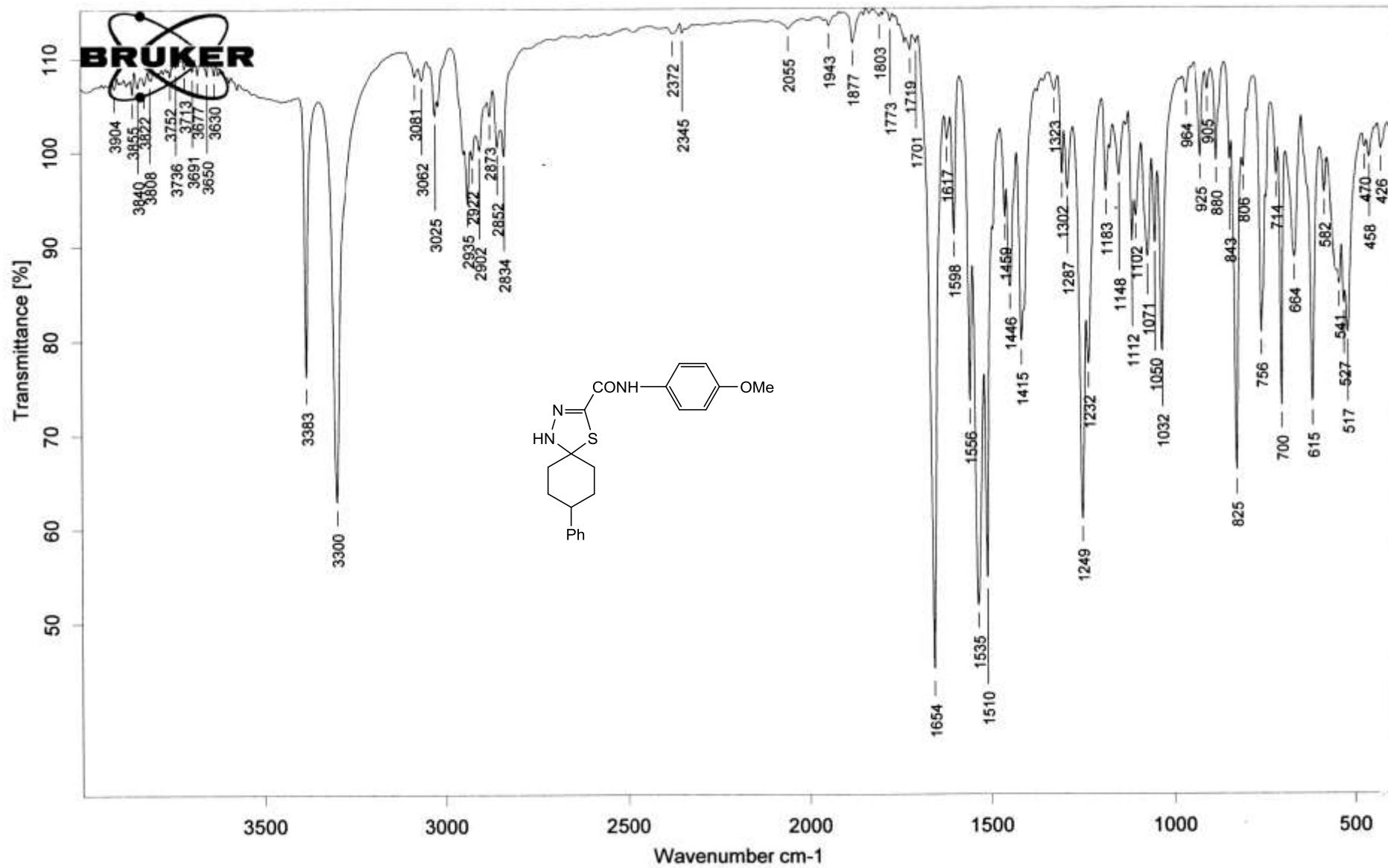
$^{19}\text{F}$  NMR spectrum of **3i** ( $\text{CDCl}_3$ ).



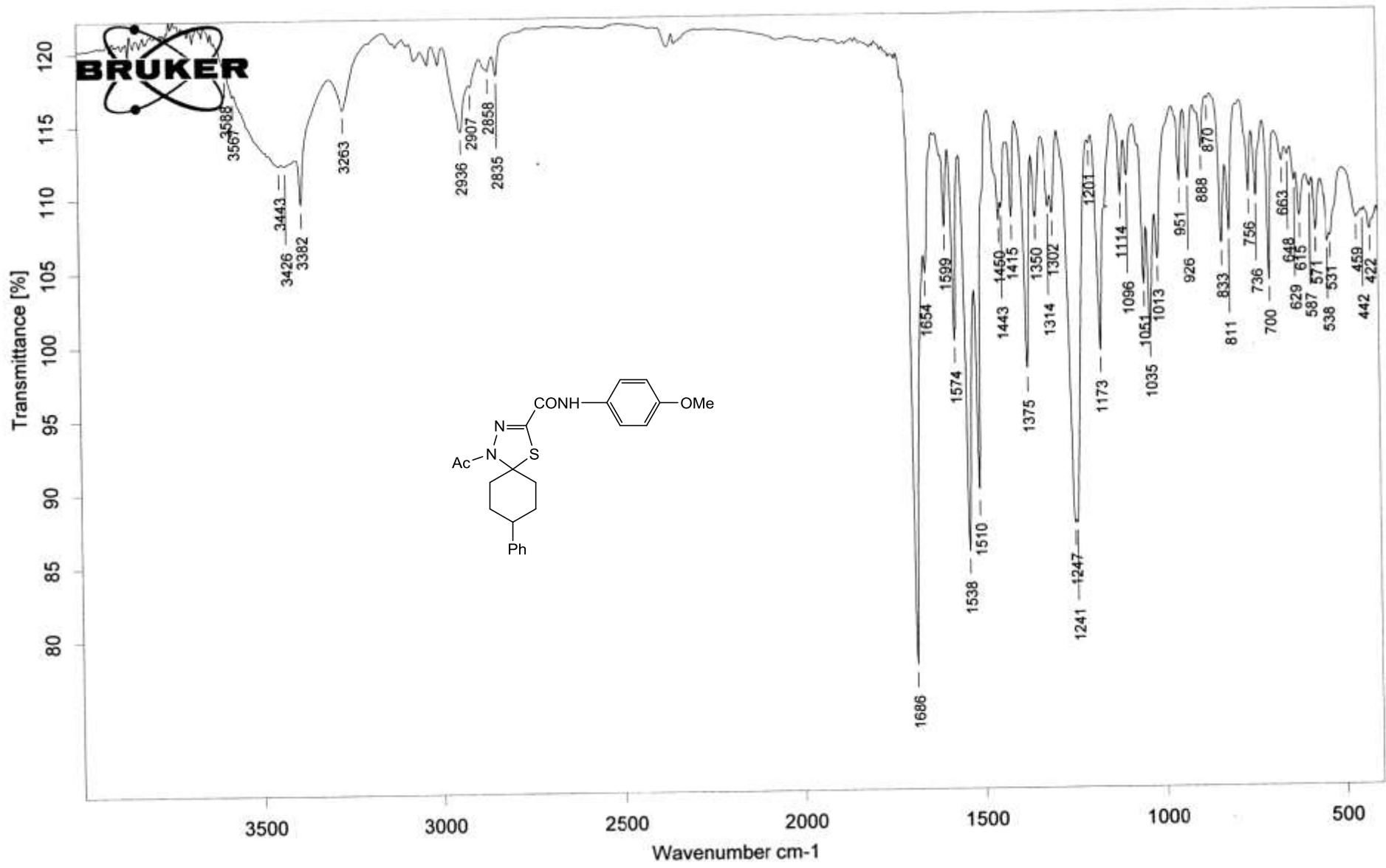
<sup>1</sup>H NMR spectrum of mixture **5** and **6**.

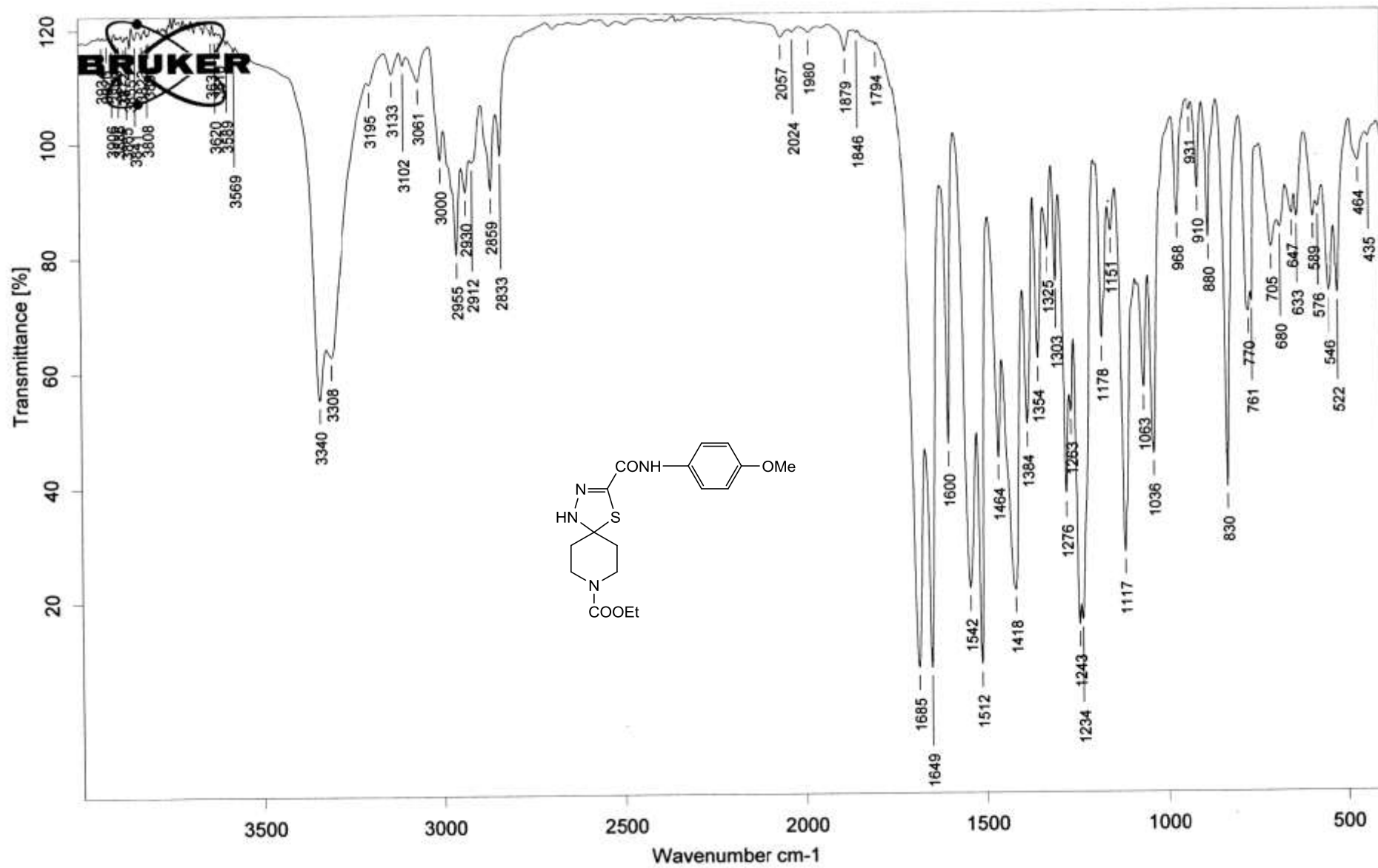
### 3. IR spectra

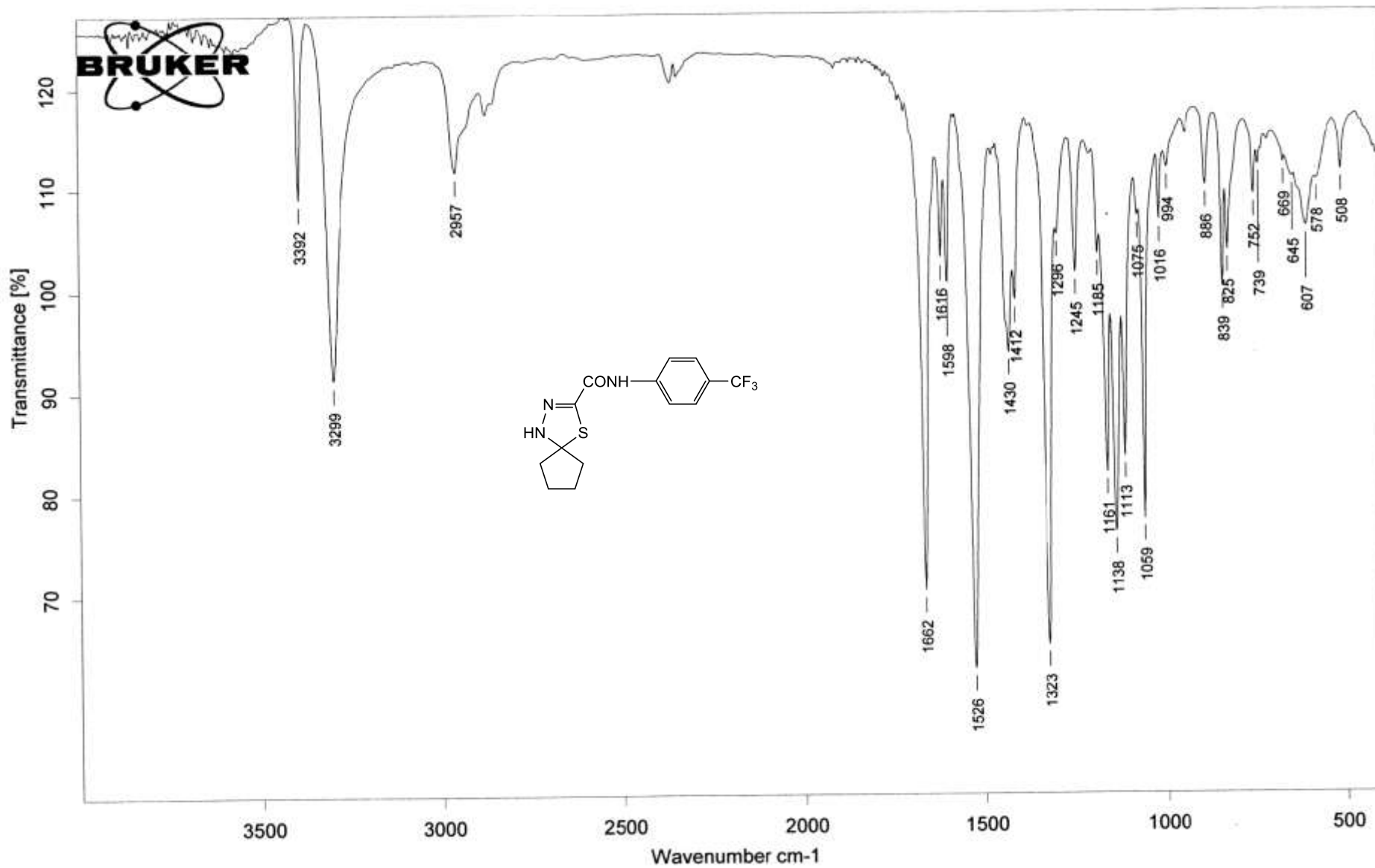


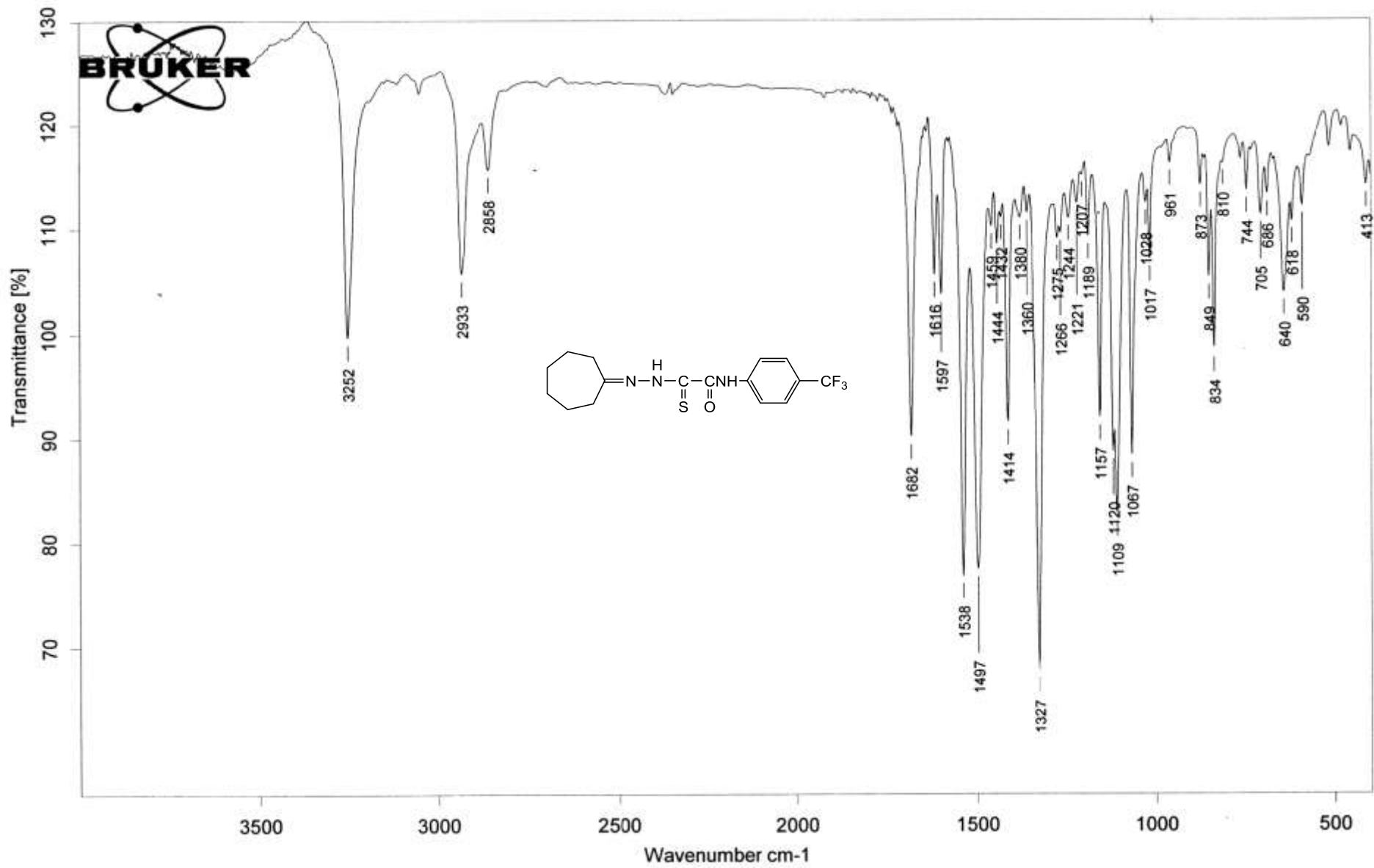


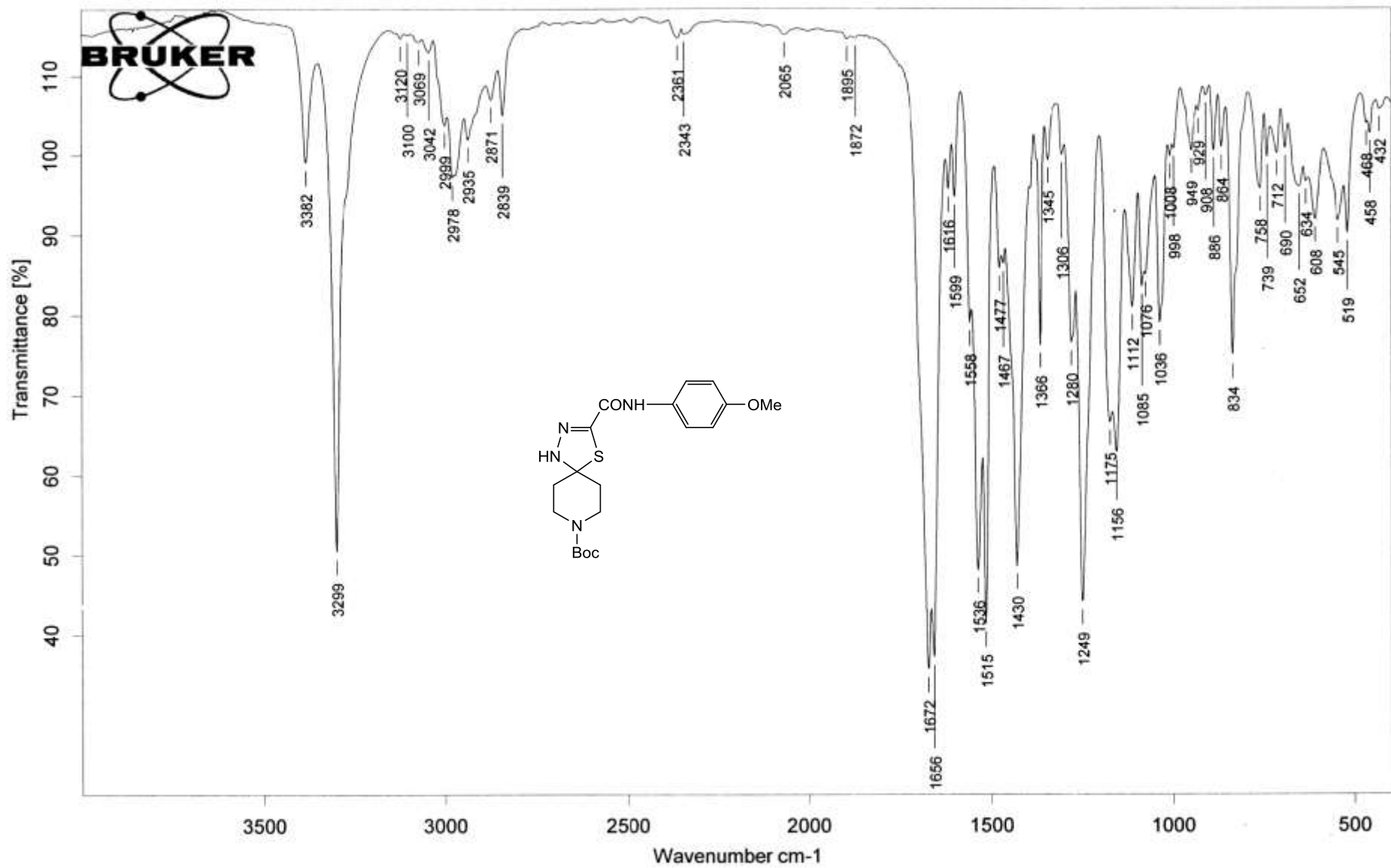


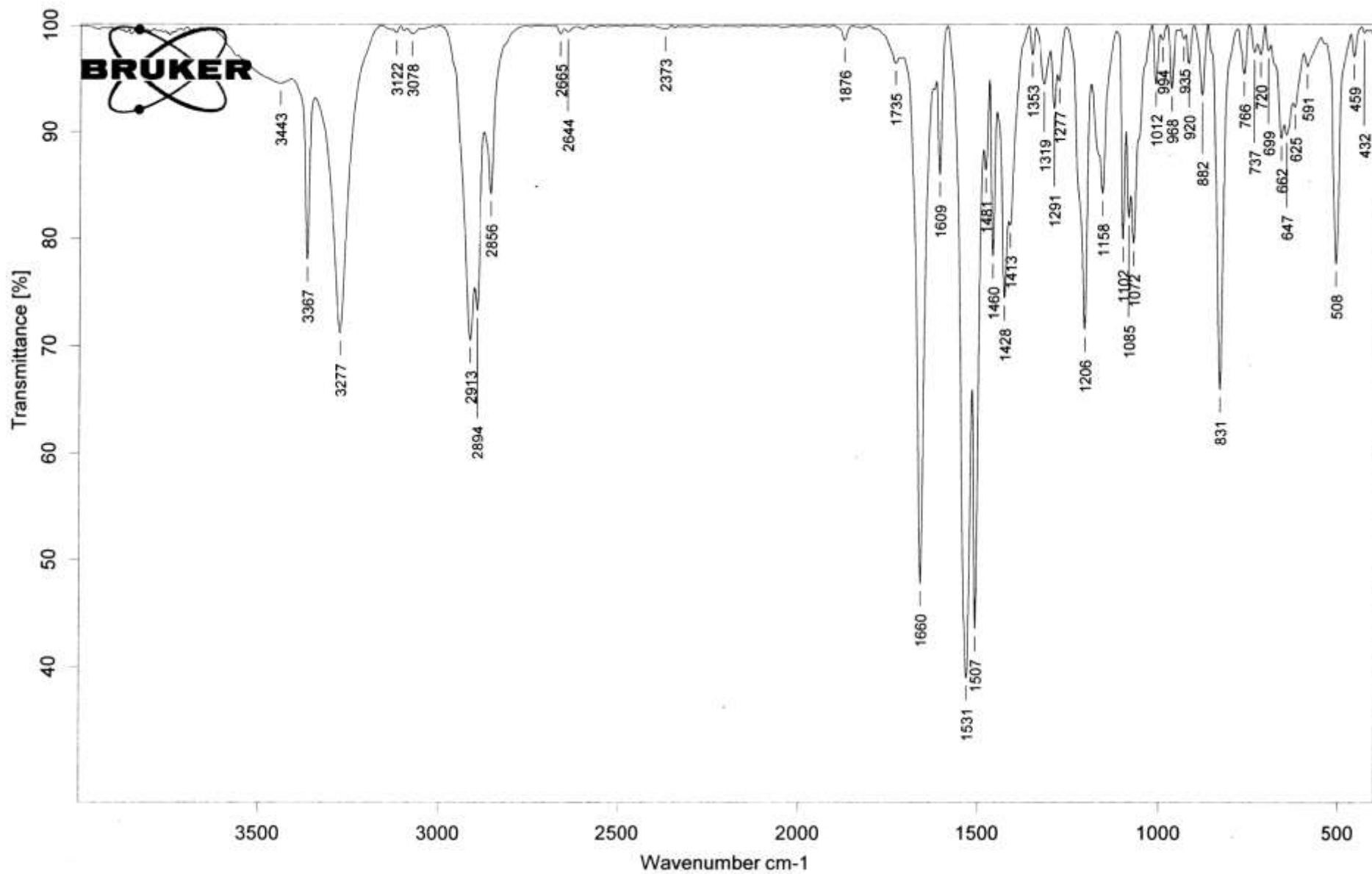












## 4. Mass spectra

### Display Report

#### Analysis Info

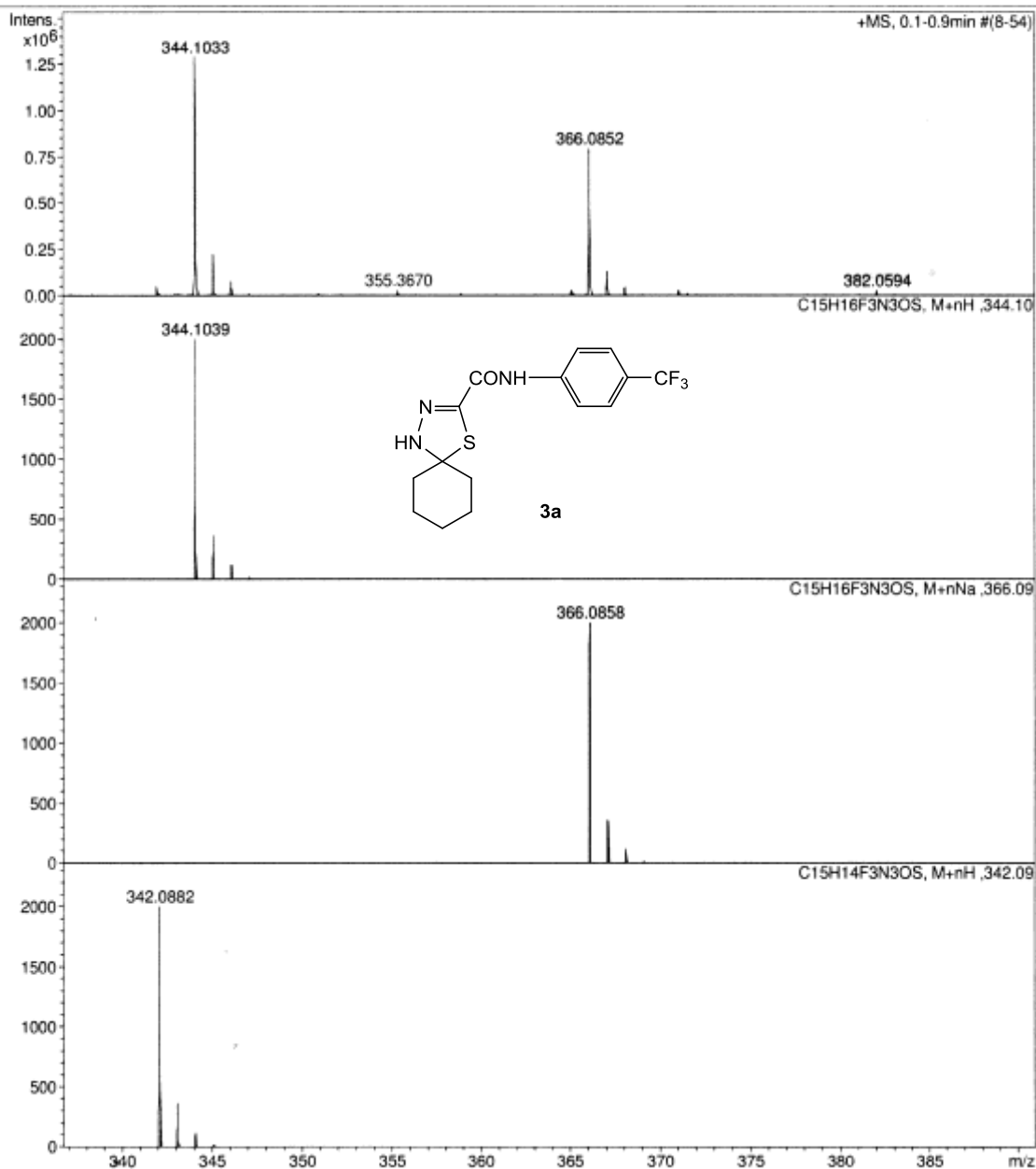
Method tune\_50-1600.m

Instrument / Ser# microTOF 10248

Comment C15H16F3N3OS mH 344.1038 calibrant added CH3OH

#### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	1.0 Bar
Focus	Not active			Set Dry Heater	200 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	1600 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of **3a**.

# Display Report

## Analysis Info

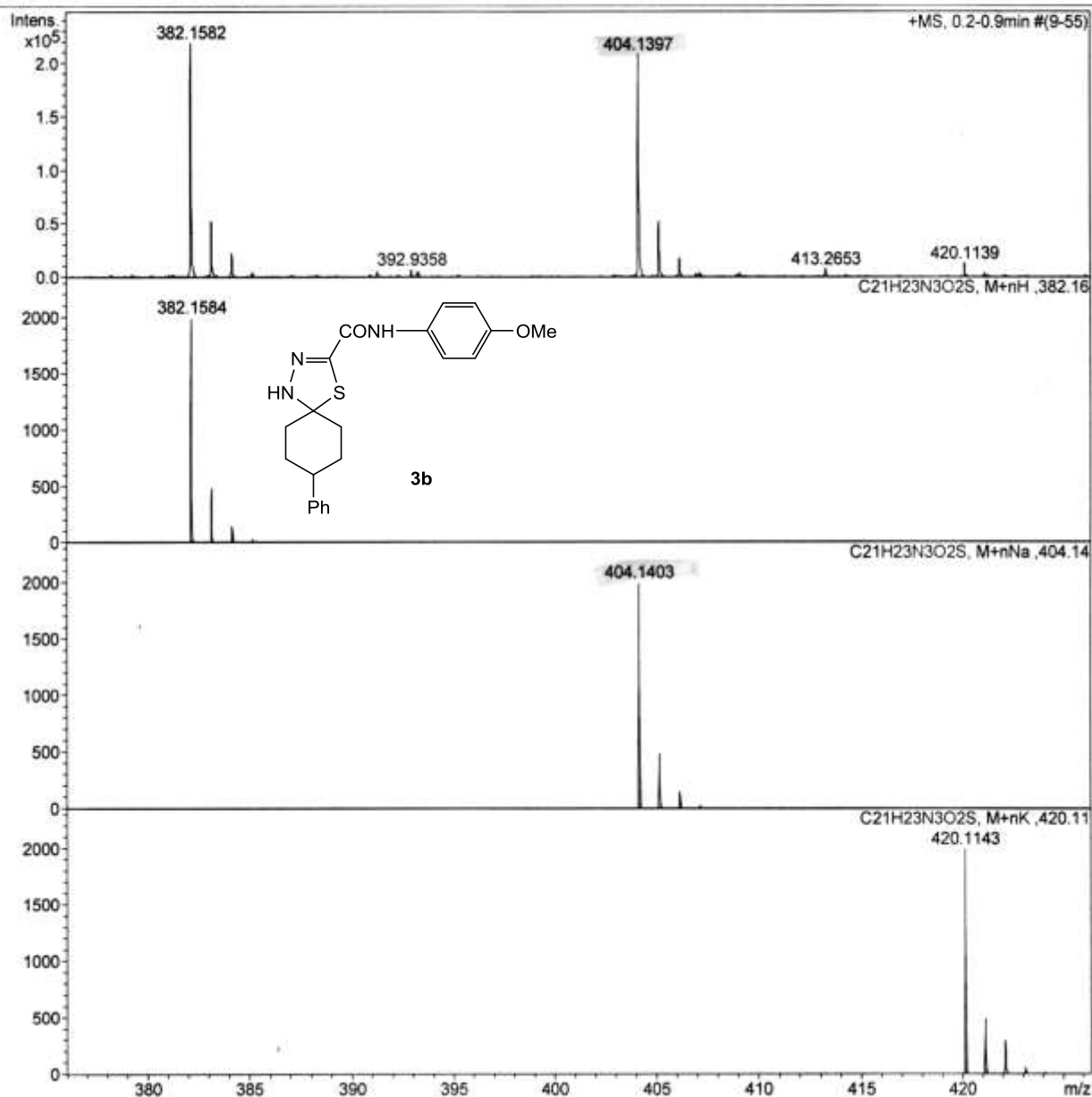
Method tune\_50-1600.m

Instrument / Ser# micrOTOF 10248

Comment C21H23N3O2S mH 382.1583 calibrant added, CH3OH

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	1.0 Bar
Focus	Not active			Set Dry Heater	200 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	1600 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of 3b.



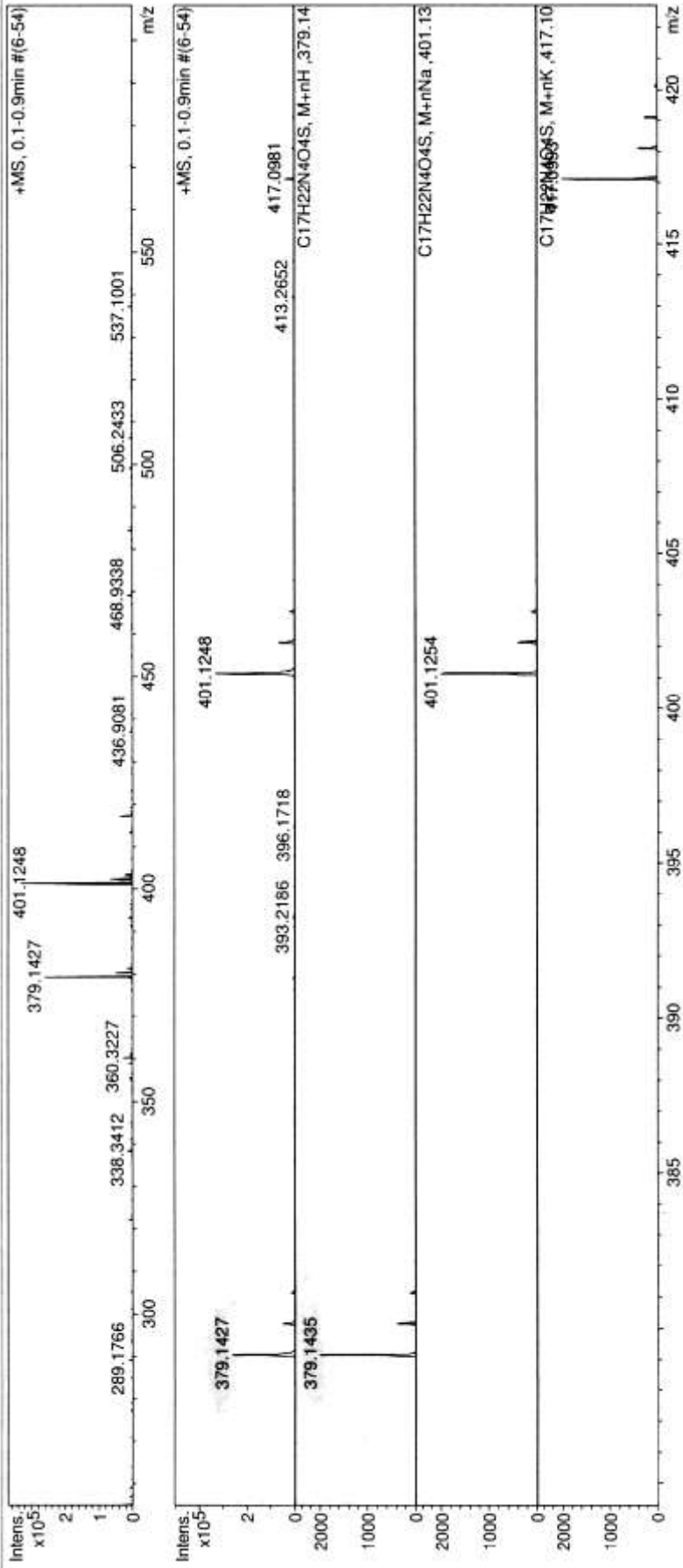
# Display Report

## Analysis Info

Method: tune\_50-1600.m  
 Instrument / Ser#: microTOF 10248  
 Comment: C17H22N4O4S mH 379.1434 calibrant added

## Acquisition Parameter

Source Type: ESI  
 Focus: Not active  
 Scan Begin: 50 m/z  
 Scan End: 1600 m/z  
 Ion Polarity: Positive  
 Set Nebulizer: 1.0 Bar  
 Set Dry Heater: 200 °C  
 Set Dry Gas: 4.0 l/min  
 Set Divert Valve: Waste



# Display Report

## Analysis Info

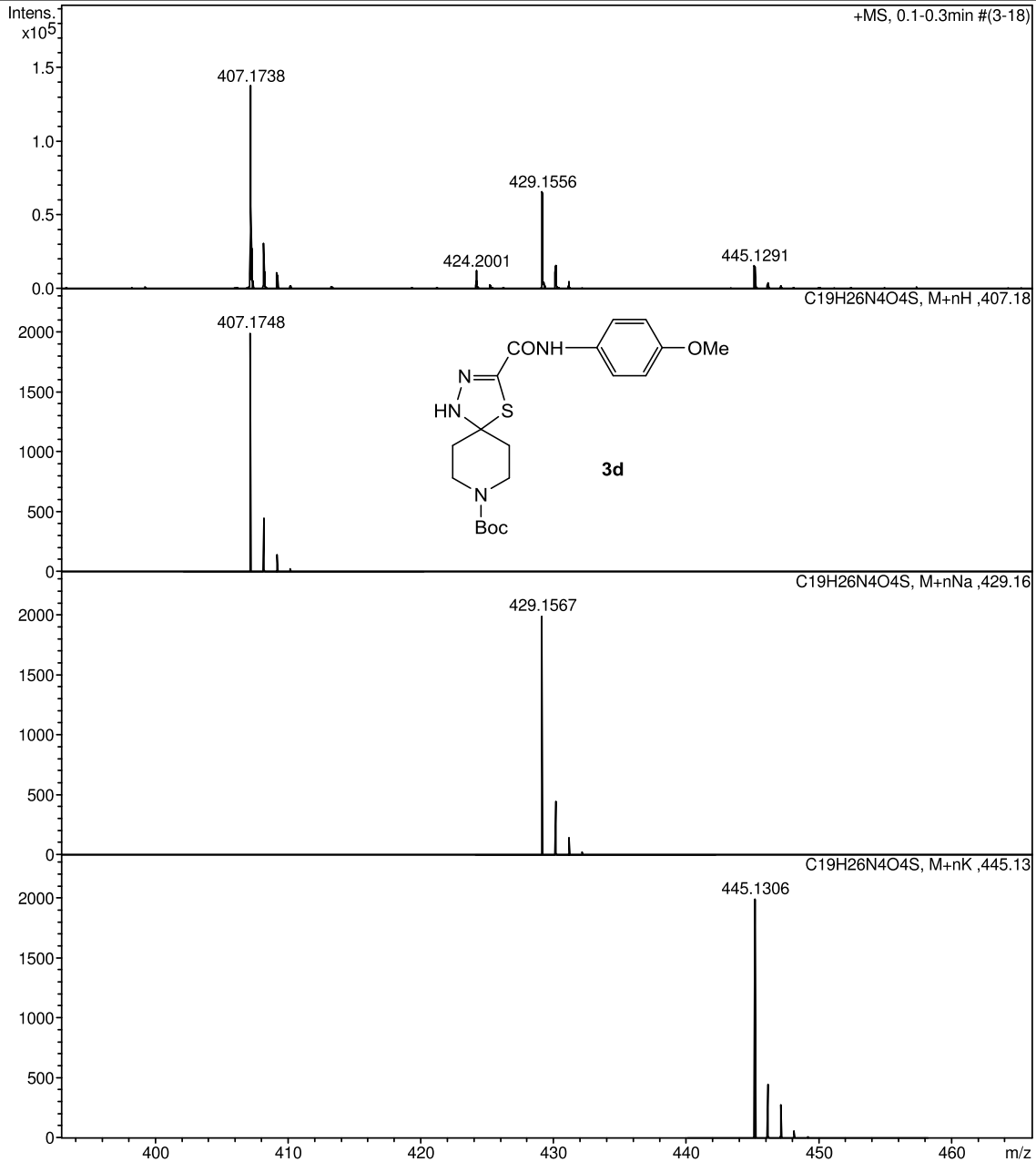
Method tune\_50-1600.m

Instrument / Ser# microTOF 10248

Comment C19H26N4O4S mH 407.1757 clb added CH3OH

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	1.0 Bar
Focus	Not active			Set Dry Heater	200 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	1600 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of **3d**.

# Display Report

## Analysis Info

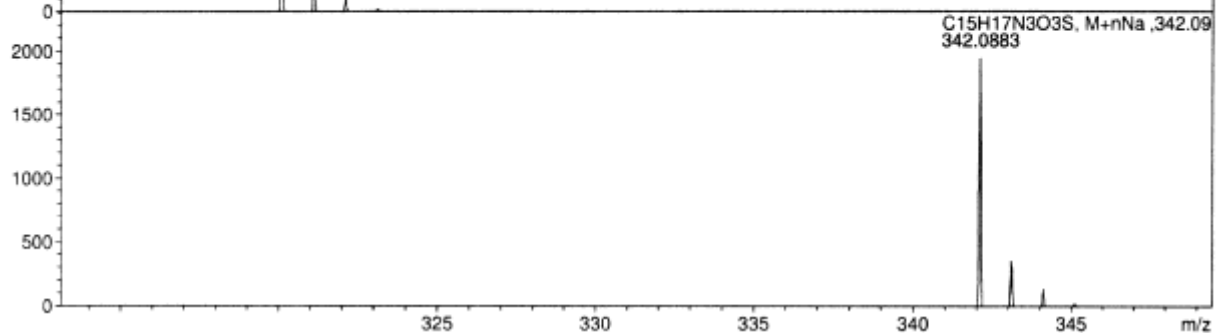
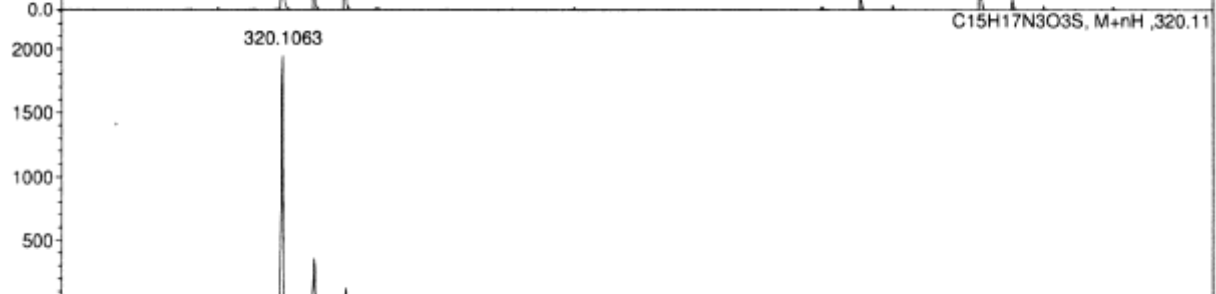
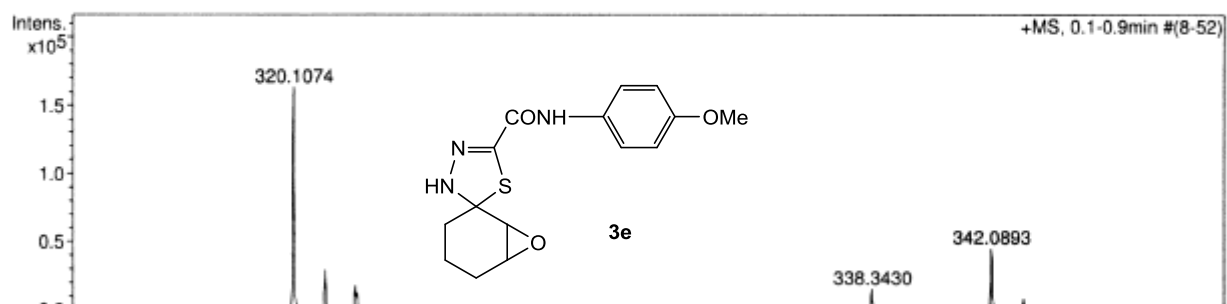
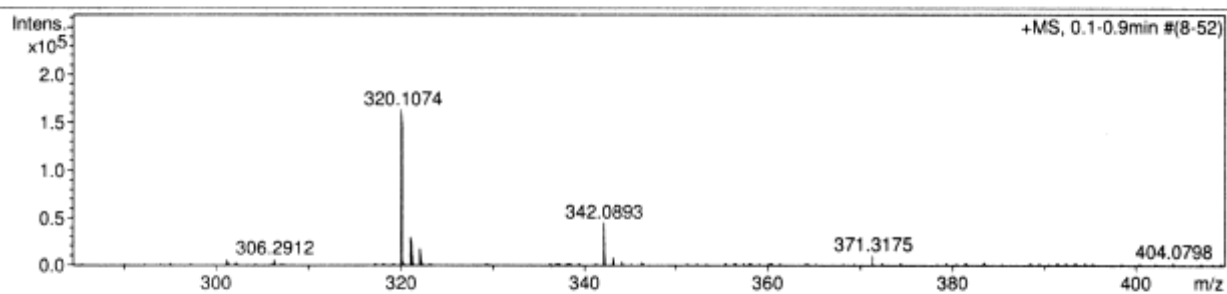
Method tune\_50-1600.m

Instrument / Ser# microTOF 10248

Comment C15H17N3O3S mH 320.1063 clb added.

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	1.0 Bar
Focus	Not active			Set Dry Heater	200 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	1600 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of **3e**.

# Display Report

## Analysis Info

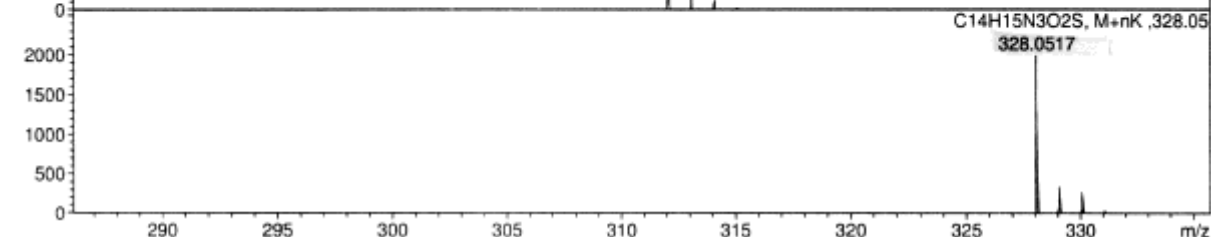
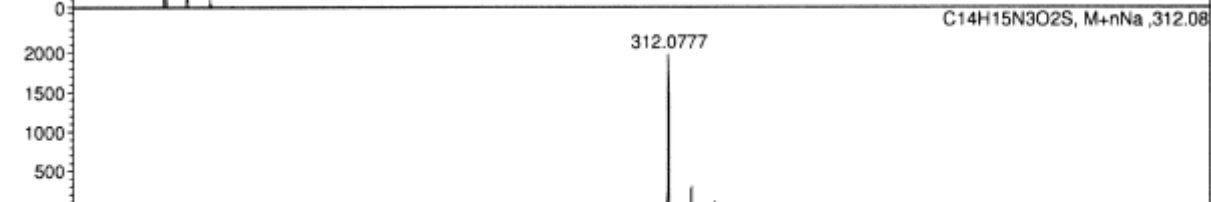
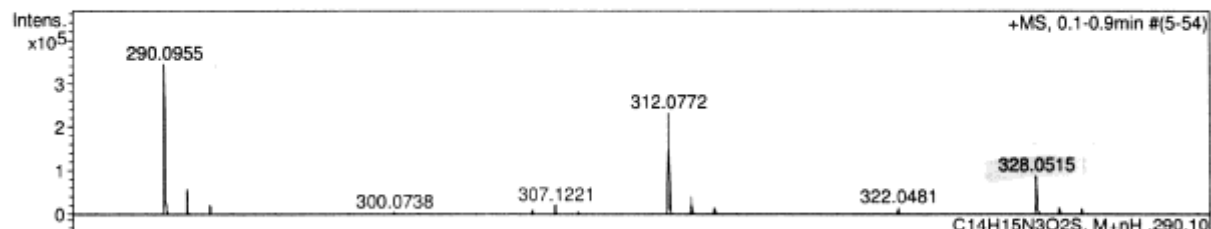
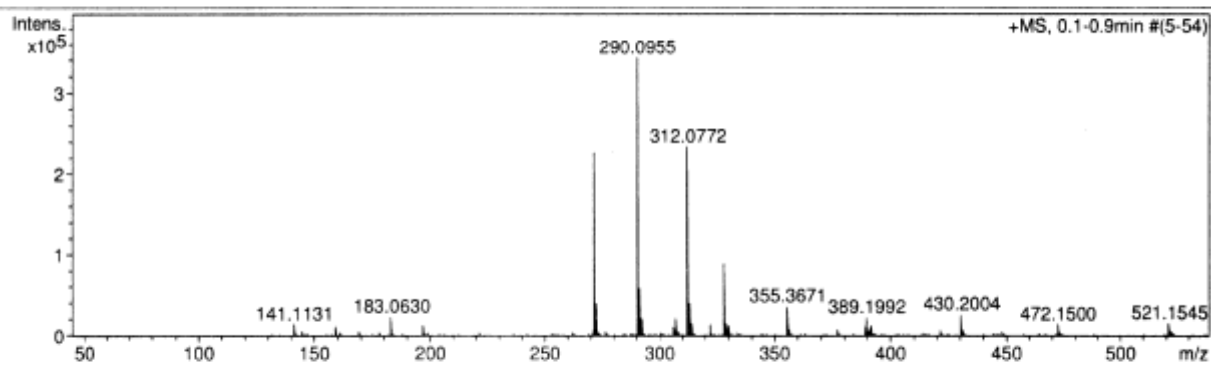
Method tune\_50-1600.m

Instrument / Ser# micrOTOF 10248

Comment C14H15N3O2S mH 290.0957 calibrant added

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	1.0 Bar
Focus	Not active			Set Dry Heater	200 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	1600 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of 3f.

# Display Report

## Analysis Info

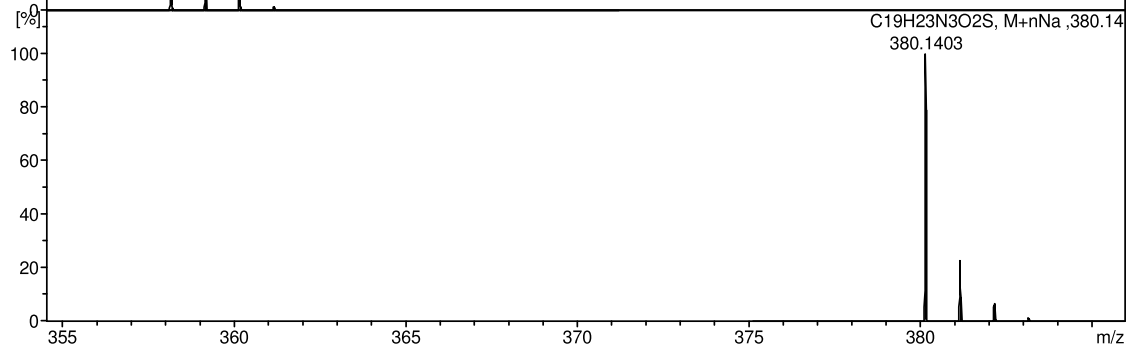
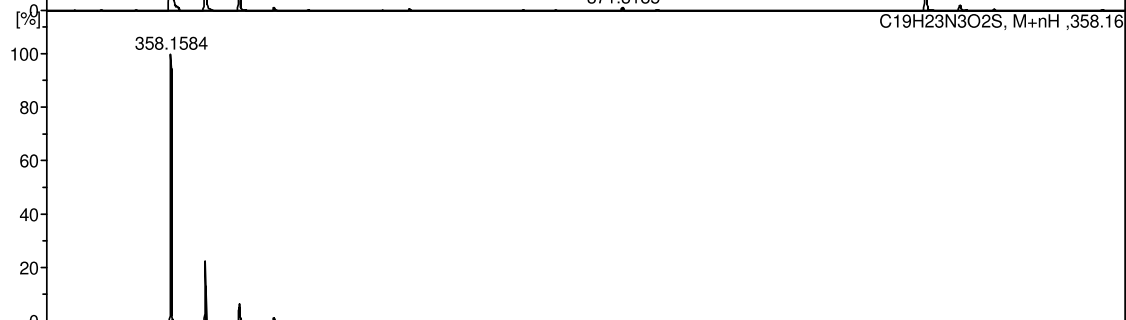
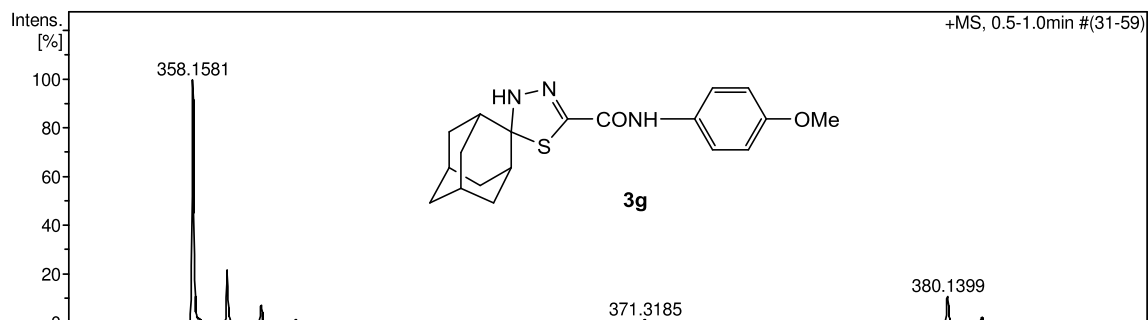
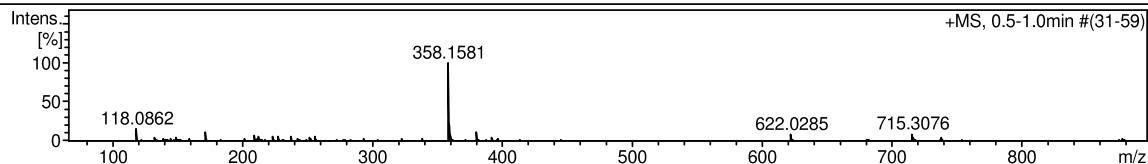
Method tune\_low.m

Instrument / Ser# micrOTOF 10248

Comment C19H25N3O2S mH 360.1740 calibrant added CH3CN

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of **3g**.

# Display Report

## Analysis Info

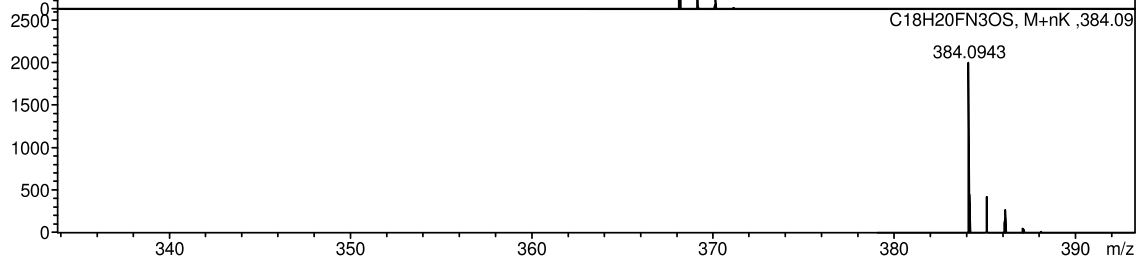
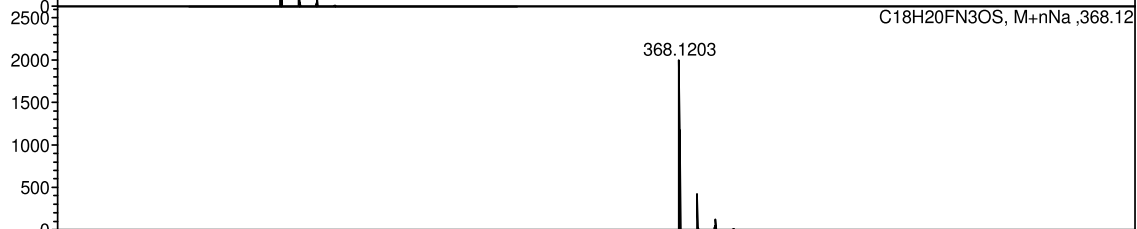
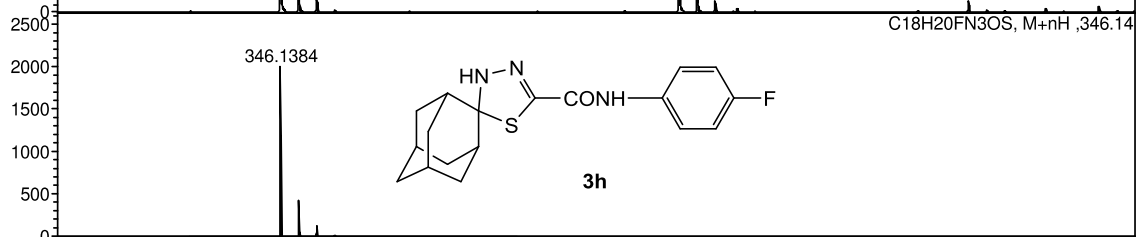
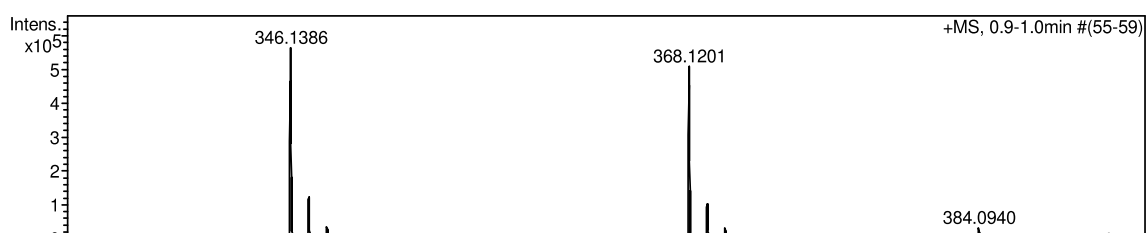
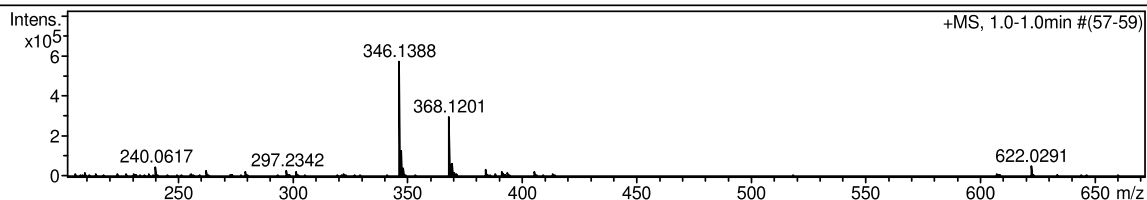
Method tune\_low.m

Instrument / Ser# micrOTOF 10248

Comment C18H20FN3OS mH 346.1383 calibrant added CH3OH

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of **3h**.

# Display Report

## Analysis Info

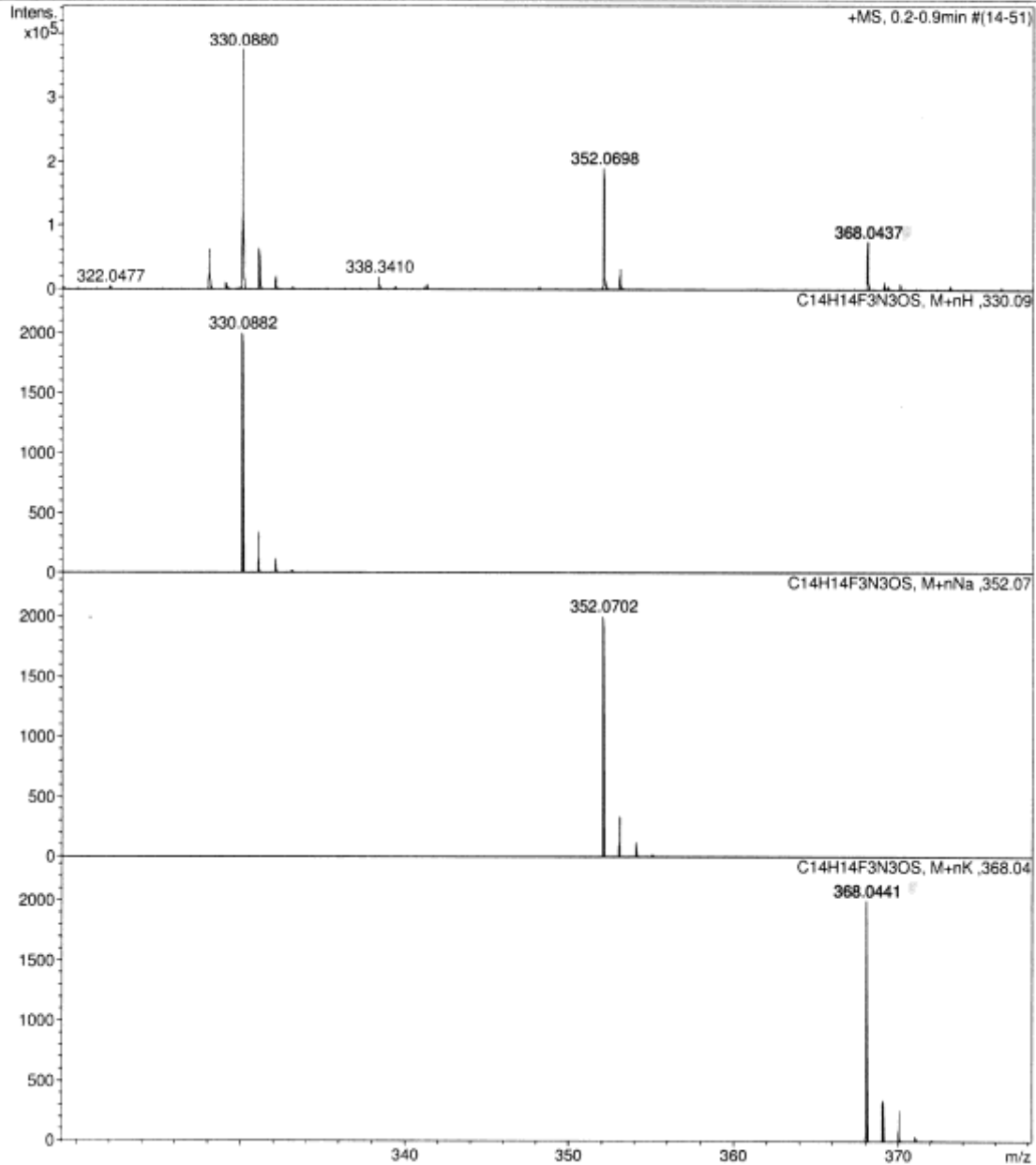
Method tune\_50-1600.m

Instrument / Ser# micrOTOF 10248

Comment C14H14F3N3OS mH 330.0882 calibrant added

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	1.0 Bar
Focus	Not active			Set Dry Heater	200 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	1600 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of **3i**.

# Display Report

## Analysis Info

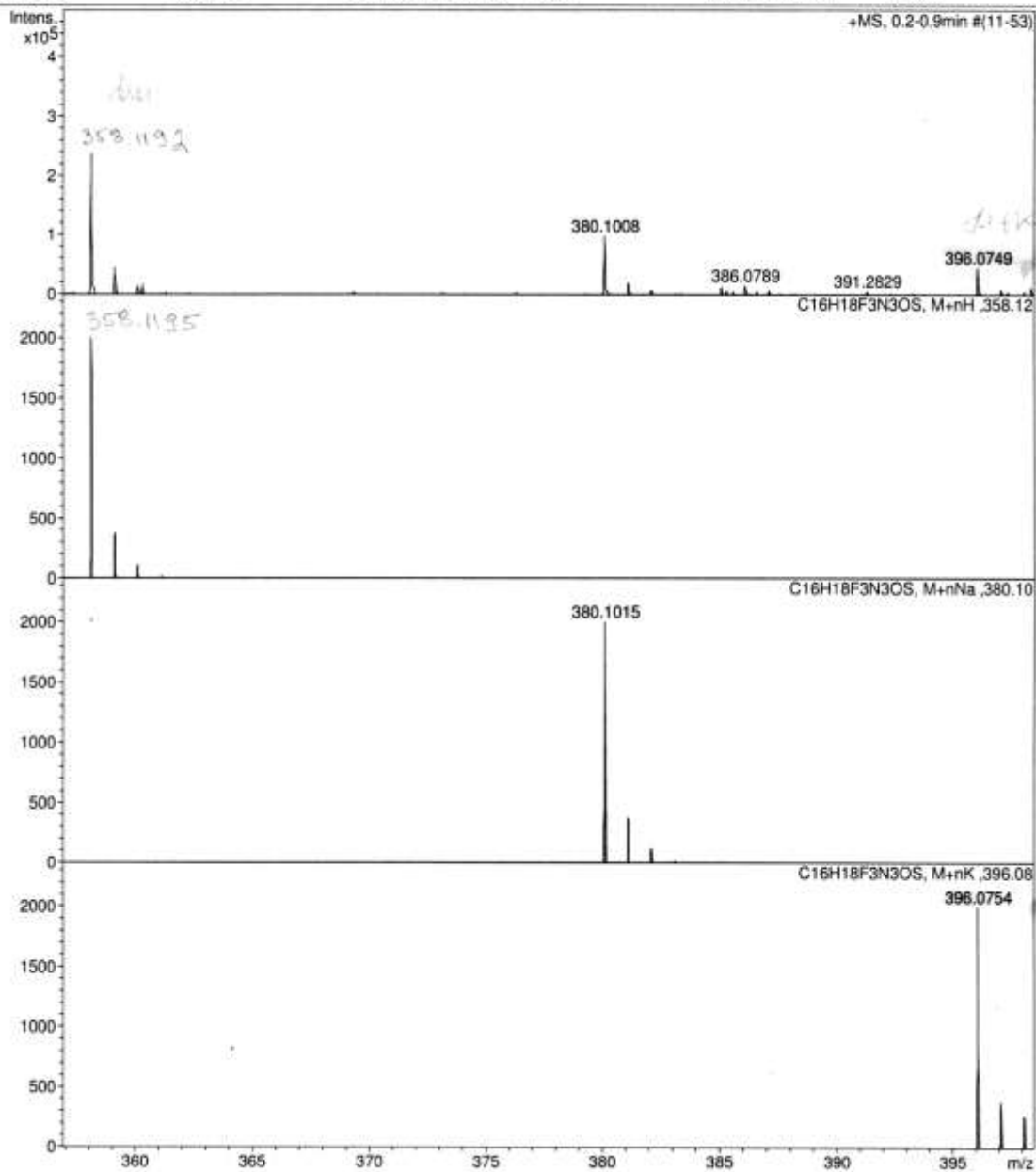
Method tune\_50-1600.m

Instrument / Ser# micrOTOF 10248

Comment C16H18F3N3OS mH 358.1195 calibrant added

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	1.0 Bar
Focus	Not active			Set Dry Heater	200 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	1600 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



Mass-spectra of **3j-4j**.



# Display Report

## Analysis Info

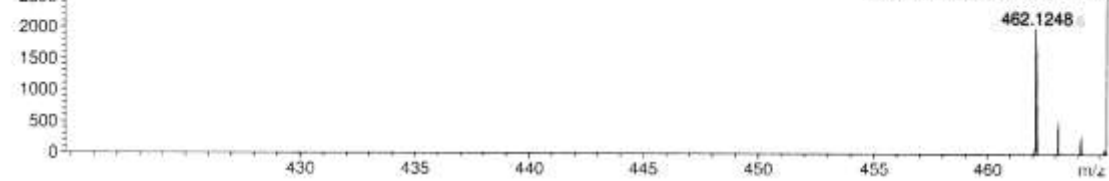
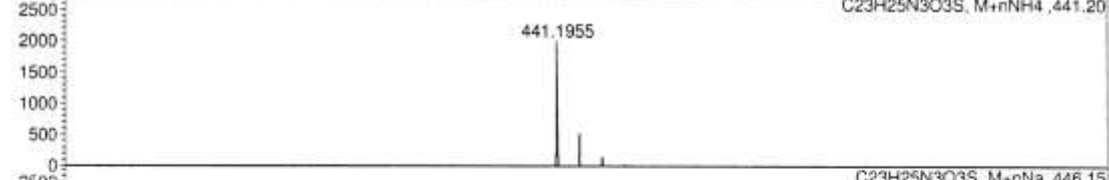
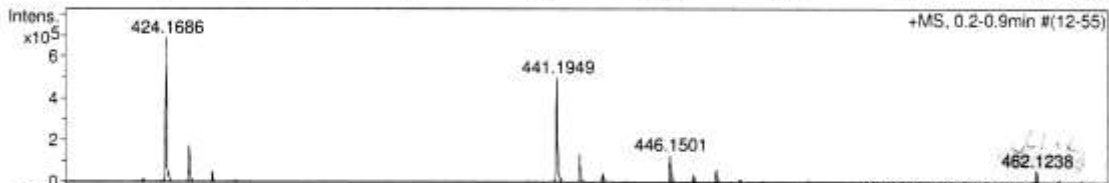
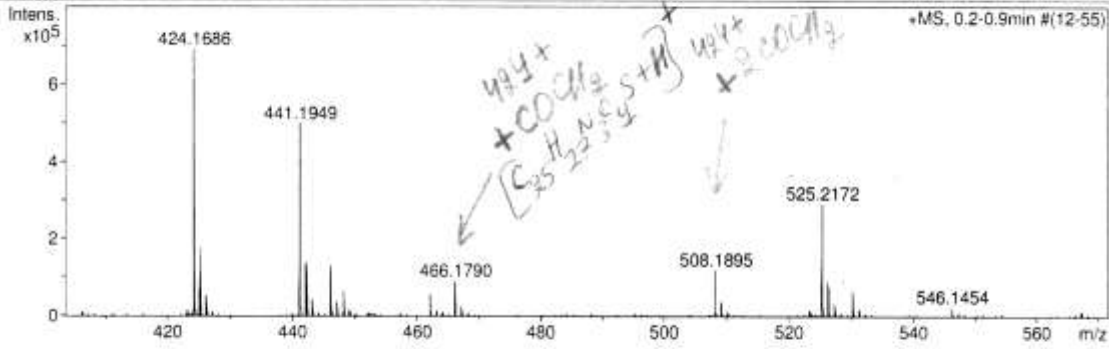
Method tune 50-1600.m

Instrument / Ser# microTOF 10248

Comment C23H25N3O3S MH 424.1689 calibrant added

## Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	1.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	1600 m/z			Set Divert Valve	Waste



Mass-spectra of 5/6.